

# PD-L1-related lncRNAs are associated with malignant characteristics and immune microenvironment in glioma

Zhiwei Xia<sup>1,\*</sup>, Ruxin Tu<sup>2,4,\*</sup>, Fangkun Liu<sup>3</sup>, Hao Zhang<sup>3,5</sup>, Ziyu Dai<sup>3</sup>, Zeyu Wang<sup>3,6</sup>, Peng Luo<sup>7</sup>, Shiqing He<sup>8</sup>, Gelei Xiao<sup>3</sup>, Jie Feng<sup>2,4,9</sup>, Quan Cheng<sup>3,4</sup>

<sup>1</sup>Department of Neurology, Hunan Aerospace Hospital, Changsha 410205, Hunan, P.R. China

<sup>2</sup>Department of Neurology, Xiangya Hospital, Central South University, Changsha 410008, Hunan Province, P.R. China

<sup>3</sup>Department of Neurosurgery, Xiangya Hospital, Central South University, Changsha 410008, Hunan Province, P.R. China

<sup>4</sup>National Clinical Research Center for Geriatric Disorders, Xiangya Hospital, Central South University, Changsha 410008, Hunan, P.R. China

<sup>5</sup>Department of Neurosurgery, The Second Affiliated Hospital, Chongqing Medical University, Chongqing 400010, P.R. China

<sup>6</sup>MRC Centre for Regenerative Medicine, Institute for Regeneration and Repair, University of Edinburgh, Little France, Edinburgh, EH16 4UU, UK

<sup>7</sup>Department of Oncology, Zhujiang Hospital, Southern Medical University, Guangzhou 510280, P.R. China

<sup>8</sup>Department of Neurosurgery, Affiliated Nanhua Hospital, Hengyang Medical College, University of South China, Hengyang 421001, Hunan, P.R. China

<sup>9</sup>Hunan Clinical Research Center for Cerebrovascular Disease, Changsha 410008, Hunan Province, P.R. China

\*Equal contribution and shared first authorship

**Correspondence to:** Jie Feng, Quan Cheng; **email:** [4010009@csu.edu.cn](mailto:4010009@csu.edu.cn), [chengquan@csu.edu.cn](mailto:chengquan@csu.edu.cn)

**Keywords:** glioma, PD-L1, LINC01271, risk score, prognosis

**Received:** March 14, 2023

**Accepted:** August 21, 2023

**Published:** October 12, 2023

**Copyright:** © 2023 Xia et al. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/3.0/) (CC BY 3.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

## ABSTRACT

**Background:** The expression of long non-coding RNA (lncRNA) can function as diagnostic and therapeutic biomarker for tumors. This research explores the role of PD-L1-related lncRNAs in affecting malignant characteristics and the immune microenvironment of glioma.

**Methods:** Downloading gene expression profiles and clinicopathological information of glioma from TCGA and CGGA databases, 6 PD-L1-related lncRNAs were identified through correlation analysis, Cox and LASSO regression analysis, establishing the risk score model based on them. Bioinformatics analysis and cell experiments *in vitro* were adopted to verify the effects of LINC01271 on glioma.

**Results:** Risk scores based on 6 PD-L1-related lncRNAs (AL355974.3, LINC01271, AC011899.3, MIR4500HG, LINC02594, AL357055.3) can reflect malignant characteristics and immunotherapy response of glioma. Patients with high LINC01271 expression had a worse prognosis, a higher abundance of M1 subtype macrophages in the immune microenvironment, and a higher degree of tumor malignancy. Experiments *in vitro* confirmed its positive regulatory effect on the proliferation and migration of glioma cells.

**Conclusions:** The risk score model based on 6 PD-L1-related lncRNAs can reflect the malignant characteristics and prognosis of glioma. LINC01271 can independently be used as a new target for prognosis evaluation and therapy.

## INTRODUCTION

Glioma is of great concern because of its high fatality rate. According to the degree of malignancies, lower-grade gliomas (LGG) are classified as World Health Organization (WHO) II and III, including diffuse low-grade and intermediate-grade gliomas. In contrast, glioblastomas (GBM) belong to WHO grade IV [1], with the highest incidence and lowest survival [2]. Surgical resection is mainly applied to treat glioma, combined with radiotherapy, chemotherapy, and other comprehensive methods [3]. Nevertheless, the therapies are inefficacious due to the infiltration and invasion of glioma. The median overall survival (OS) of GBM patients is only 12-18 months after diagnosis, even under the optimal treatment [4]. For decades, researchers have actively explored glioma's pathological features to discover biomarkers for early diagnosis, providing targets for gene therapy and immunotherapy [3–6], aiming to provide better health care and personalized medicine for glioma patients.

Immunotherapy has greatly advanced the treatment of malignant tumors, with immune checkpoint inhibitors (ICIs) playing a crucial role. Classical immune checkpoints include programmed cell death protein 1 (PD-1), PD-L1 and cytotoxic T lymphocyte antigen 4 (CTLA-4). The ICIs can inhibit the interaction between the ligand and immunosuppressive receptors, thereby preventing immune evasion [7]. Nowadays, ICIs have shown significant advantages in lung cancer and breast cancer [5, 8]. This study starts with PD-L1-related lncRNAs, elucidating the immunization characteristics of glioma and their prognostic implications.

In the last decade, lncRNAs have gained recognition for their significant biological effects. They are defined as RNA molecules longer than 200 nucleotides that cannot encode proteins [6]. They play a critical role in many life activities such as epigenetics, cell cycle, and differentiation. Their expression is strictly regulated under physiological conditions [9]. lncRNAs' function as biomarkers or therapeutic targets for tumors has received extensive attention [10–13]. Lu et al. found that N6-methyladenosine-related non-coding RNAs showed excellent performance in predicting prognosis and immunotherapy response in bladder cancer [14]. Lu et al. proved the potential of lncRNAs as prognostic markers and personalized therapeutic targets for ovarian cancer treatment [15, 16]. The abnormal expression of lncRNAs is associated with glioma occurrence, progression, invasiveness, and recurrence [17–20], through diverse molecular mechanisms that directly or indirectly regulate gene expression [21, 22]. However,

there is a paucity of studies investigating PD-L1-related lncRNAs in gliomas.

Herein, for the first time, we established a risk score model containing 6 PD-L1-related lncRNAs to reflect individual heterogeneity of gliomas and provide a tool for prognostic stratification. LINC01271 was chosen as the target lncRNA to validate its predictive efficacy on the clinical and pathological features, and prognosis of glioma (Figure 1 Flow Chart).

## RESULTS

### Identification of lncRNAs related to PD-L1

The genetic information, RNA transcriptome data, and related clinical data of glioma patient samples were acquired from the TCGA and CGGA database. PD-L1 is encoded by gene *CD274*. Through correlation analysis and univariate logistic regression analysis, 23 lncRNAs were selected from 13895 lncRNAs associated with *CD274* expression. Then multivariate cox analysis was used to figure out lncRNAs independently related to *CD274*. With  $p < 0.05$  as the standard, 6 lncRNAs (AL355974.3, LINC01271, AC011899.3, MIR4500HG, LINC02594, AL357055.3) were screened. Finally, further dimension reduction and model construction were performed through LASSO analysis (Figure 2A–2C and Supplementary Table 1).

### Risk score model can predict the prognosis of glioma patients

We observed the effects of the 6 PD-L1-related lncRNAs on the prognosis of glioma in TCGA database. Kaplan-Meier survival curve analysis showed AL355974.3 (HR=4.06,  $p < 0.001$ ), LINC01271 (HR=3.51,  $p < 0.001$ ), and AC011899.3 (HR=2.66,  $p < 0.001$ ) were significantly related to a poorer prognosis; MIR4500HG (HR=0.39,  $p < 0.001$ ), LINC02594 (HR=0.48,  $p < 0.001$ ), and AL357055.3 (HR=0.28,  $p < 0.001$ ) were relevant to a better prognosis (Supplementary Figure 1). The findings suggest that using a prediction model based on these 6 lncRNAs can enhance the prognostic accuracy of glioma.

We established a risk score model based on the 6 PD-L1-related lncRNAs. The correlation analysis revealed a positive association between AL355974.3, LINC01271, and AC011899.3 with the risk score, whereas MIR4500HG, LINC02594 and AL357055.3 exhibited an inverse correlation (Supplementary Figure 2).

The prognostic outcome of glioma is intricately linked to the risk score. The survival analysis of the TCGA data revealed that, among all glioma patients, the low-risk score group exhibited significantly superior

prognosis compared to the high-risk score group ( $p < 0.0001$ ) (Figure 3A–3C). However, in terms of tumor pathological classification, only the PFI shows a significant difference among GBM patients with varying risk scores ( $p = 0.018$ ). In contrast, the three prognostic indicators of LGG patients are all significant differences (OS, DSS:  $p < 0.0001$ , PFI:

$p = 0.00094$ ) (Figure 3D–3I). The survival analysis in the CGGA validation dataset, which contains 306 patients, yields similar results. Significantly different OS between two risk groups exists among both all glioma and separate LGG patients (OS<sub>ALL</sub>:  $P < 0.0001$ ; OS<sub>LGG</sub>:  $P = 0.00024$ ), but not in GBM subgroup ( $p = 0.37$ ) (Figure 3J–3L).

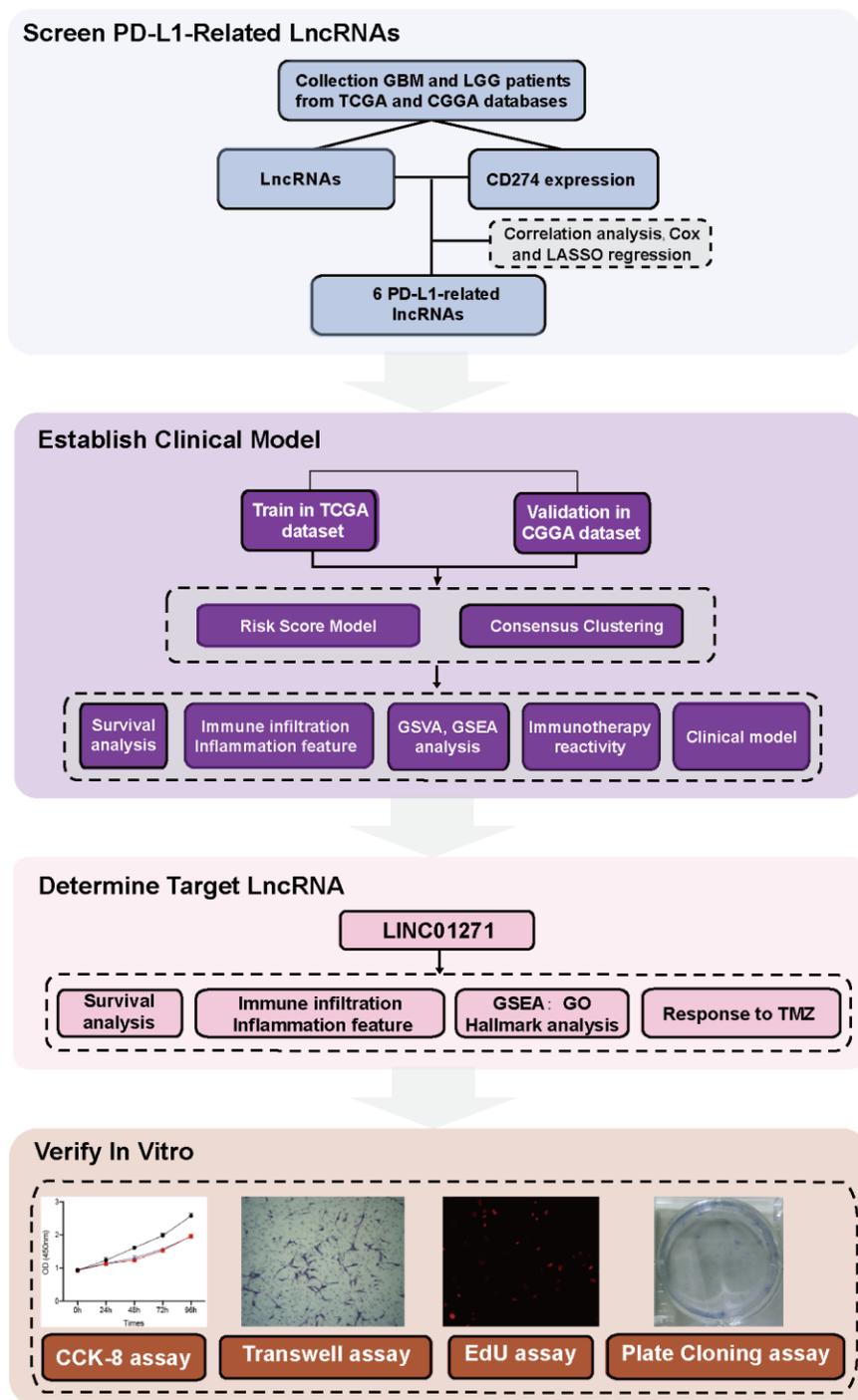
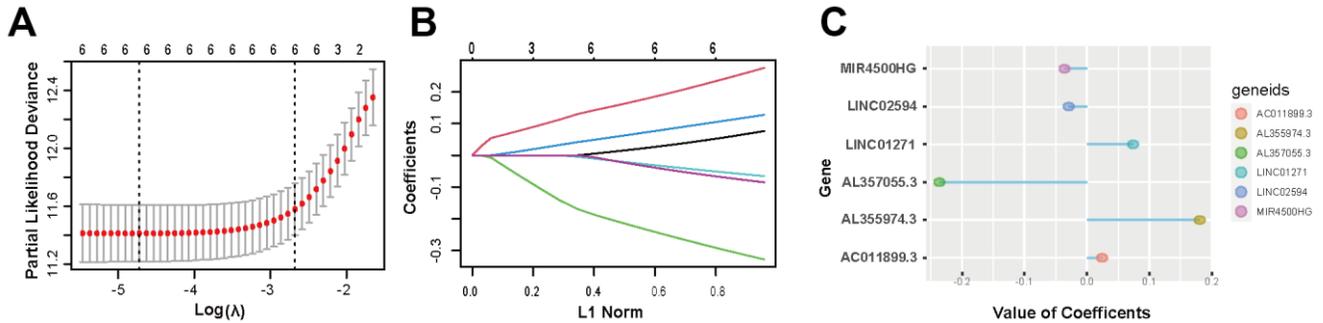


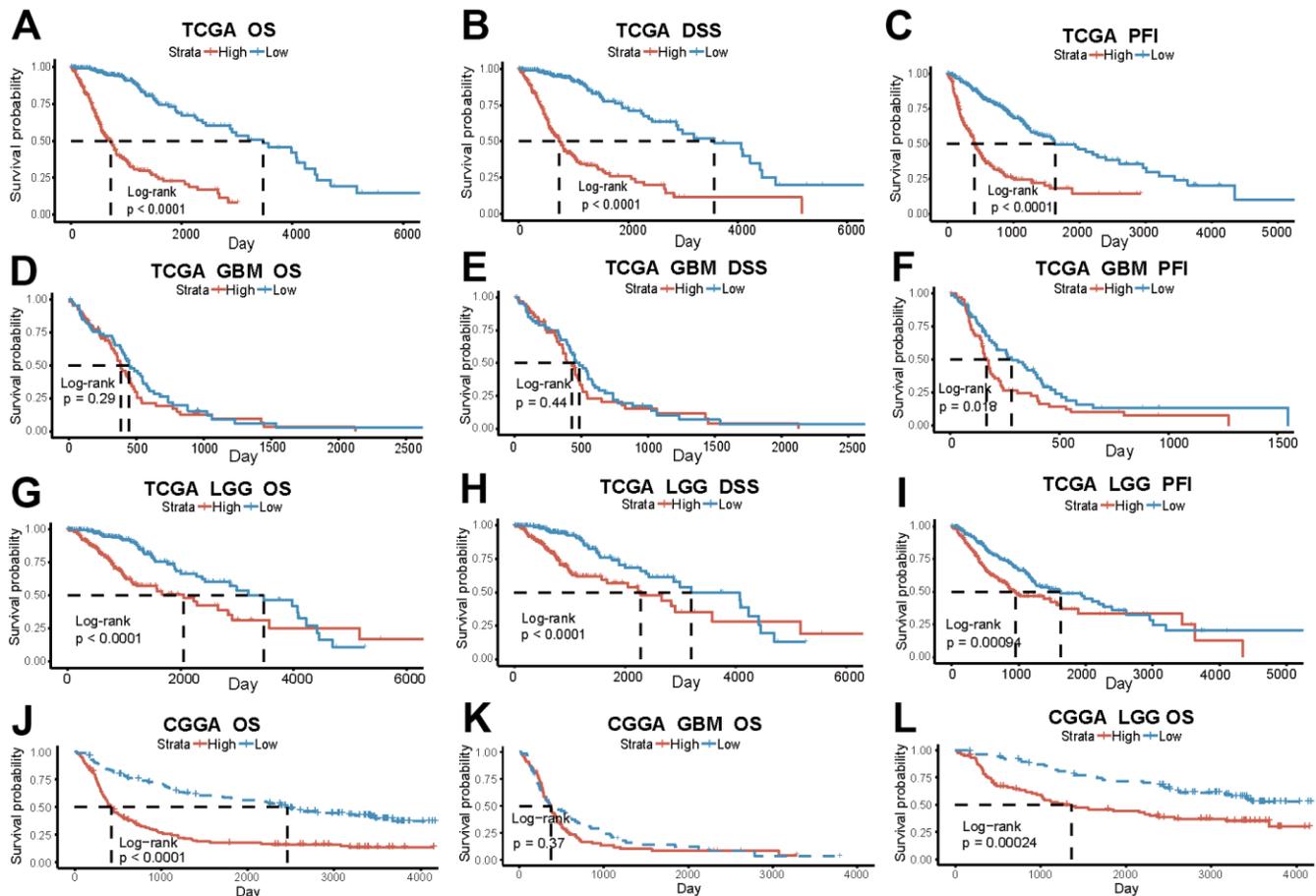
Figure 1. Flow chart.

To further explore the prognostic efficiency of PD-L1-related lncRNAs, we use consensus cluster analysis to group samples, a clear distinction between the groups was observed when  $k=2$  (Supplementary Figure 3A–3C). Cluster1 and Cluster2 can be distinguished by

principal component analysis (PCA), proving the clustering rationality (Supplementary Figure 3D, 3E). The risk score for Cluster1 generated by cluster analysis is significantly higher than that of Cluster2 ( $p < 2.2 \times 10^{-16}$ ) (Supplementary Figure 3F). Survival analysis using



**Figure 2. Screening of 6 PD-L1-related lncRNAs.** (A, B) LASSO regression analysis is employed to determine the optimal penalty coefficient ( $\lambda$ ). (C) Regression coefficients of 6 PD-L1-related lncRNAs.



**Figure 3. The risk score based on PD-L1-related lncRNAs indicates glioma patients' prognosis.** (A–C) OS, DSS, PFI in the high and low-risk group of all glioma patients in TCGA. (D–F) OS, DSS, PFI of GBM patients in TCGA. (G–I) OS, DSS, PFI of LGG patients in TCGA. (J–L) OS of all patients, GBM and LGG patients in CGGA.

TCGA data showed worse prognosis of Cluster 1 among all glioma patients and LGG subgroup. But in GBM subgroup, only PFI had a statistical difference (Supplementary Figure 3G–3O). In the CGGA database, poorer prognosis of Cluster 1 patients was showed both in the overall glioma patients and in the subgroup analysis (Supplementary Figure 3P–3R).

These results suggest that the risk score model, which is based on 6 PD-L1-related lncRNAs, exhibits favorable predictive efficacy in glioma patient prognosis, especially in the LGG patient group.

### **Risk score is correlated with the clinical, pathological, and genetic characteristics of glioma**

The association between risk scores and clinical features of glioma was investigated. 672 patients enrolled from TCGA database were categorized into two groups based on the median risk score, with high and low expression groups identified. Patients with high-risk score tended to be older and more frequently diagnosed with GBM, which exhibited IDH wild type, non-codel of 1p/19q, and unmethylated MGMT pathological features (Table 1). Heatmap and box plots provide a more intuitive and accurate representation, patients with high-risk scores had higher levels of *CD274* expression and more malignant glioma pathologic features, regardless of gender (Figure 4A and Supplementary Figure 4A–4H). Among GBM and LGG subgroups, IDH wild type was more common in high-risk groups, there was no significant difference in MGMT and 1p/19q status (Supplementary Figure 4I–4L).

Genomic heterogeneity leads to prognostic differences in tumors. Patients with higher tumor mutation burden (TMB) have higher risk scores ( $p < 0.001$ , Figure 4B). In the TCGA set, 292 (91.82%) of the 318 patients with high-risk score and 303 (96.19%) of the 315 patients with low-risk score had somatic mutations (Figure 4C, 4D). Genome variation analysis shows that low-risk patients have higher mutation frequency of *IDH1*, *ATRX*, *TP53*, *NOTCH1*, *IDH2*, and *CIC* ( $p < 0.001$ ), while *EGFR*, *PTEN*, *TTN*, *RB1*, and *NF1* gene mutation frequency is higher in a high-risk group ( $p < 0.001$ ) (Figure 4E).

### **Risk score correlates with the immune inflammatory microenvironment and immunotherapy responsiveness of glioma**

Immune and inflammatory cells play a crucial role in the composition of tumor tissue, with the tumor immune microenvironment (TIME) exerting significant influence on both disease progression and response to treatment. We use GSVA to identify the immune infiltration and

inflammation profiles associated with characteristic lncRNAs and draw correlation maps and heatmaps.

To clarify whether PD-L1-related lncRNAs affect prognosis by influencing the tumor immunity and inflammation, we compared the degree of immune cell infiltration. Patients in the high-risk group exhibited a greater diversity of immune cell infiltration compared to those in the low-risk group. In TCGA data, a significant increase in macrophages (M0, M1), neutrophils, CD8<sup>+</sup> T cells, T follicular helper cells and eosinophils within the high-risk score group was observed ( $p < 0.001$ ). While patients in the low-risk score group exhibited a marked elevation in monocytes ( $P < 0.001$ ) (Figure 5A, 5B and Supplementary Figure 5A). Consistently, in the CGGA set, macrophages had a higher abundance and relevance of risk score in the high-risk score group and Cluster 1 (Supplementary Figure 5B, 5C).

In TCGA set, high-risk group and Cluster 1 were associated with higher expression of MHC-I, MHC-II, LCK, HCK, interferon and particularly *STAT1* among inflammation-related molecules. Meanwhile, IgG showed increased expression in low-risk group and Cluster 2 (Figure 5C, 5D). The results of the CGGA dataset verification reveal slight variations in the relevance degrees of 7 molecules, with MHC-I exhibiting the most robust positive association with high-risk scores (Supplementary Figure 5D, 5E). This suggests that PD-L1-related lncRNAs regulate lymphocyte activation, activation of antigen-presenting cells and interferon signalling in gliomas.

The growth and progression of cancer are in connection with immunosuppression. ICIs have been considered revolutionary immunotherapy for a variety of tumors, including glioma; the tumor microenvironment influences the response of the tumor to immune checkpoint inhibitor therapies. We analyzed the expression of five common immune checkpoints (*CTLA-4*, *HAVCR2*, *IDO1*, *LAG3*, *PDCD1*) in patients, and found that people with high-risk scores had higher expression of these five immune checkpoints ( $P < 0.001$ , Figure 5E). We further forecasted the patient's response to anti-PD-1 and anti-CTLA-4 therapies. We observed a higher likelihood of sensitivity to anti-PD-1 treatment among patients in the high-risk group ( $p < 0.05$ ), while the response predicted to anti-CTLA-4 treatments of the two groups is similar (Figure 5F). Risk scores based on PD-L1-related lncRNAs can predict response to immunotherapy in glioma patients and thus guide personalized medicine.

For a profound understanding of the potential biological mechanisms of PD-L1-related lncRNAs, GSEA analysis

**Table 1. Clinical characteristics of patients with high and low risk score.**

Characteristic	N	High, N=336 <sup>1</sup>	Low, N=336 <sup>1</sup>	p-value <sup>2</sup>
<b>Age</b>	672	54(42,63)	38(30,48)	<0.001
<b>Gender</b>	672			>0.9
Female		143(43%)	142(42%)	
Male		193(57%)	194(58%)	
<b>Cancer</b>	672			<0.001
GBM		143(43%)	7(2.1%)	
LGG		193(57%)	329(98%)	
<b>IDH</b>	662			<0.001
Mutant		122(37%)	312(94%)	
WT		207(63%)	21(6.3%)	
<b>1p/19q</b>	668			<0.001
codel		50(15%)	121(36%)	
non-codel		282(85%)	215(64%)	
<b>MGMT</b>	635			<0.001
Methylated		184(61%)	294(88%)	
Unmethylated		117(39%)	40(12%)	

<sup>1</sup>Median(IQR); n(%).

<sup>2</sup>Wilcoxon rank sum test; Pearson's Chi-squared test.

was conducted. The results revealed that the biological processes of PD-L1-related lncRNAs were enriched in neutrophil activation involved in immune response, neutrophil degranulation, and T cell activation. Cell components were mainly concentrated at the leading edge of the cell, on the outer side of both the plasma membrane and synaptic membrane. The molecular functions were focused on passive transmembrane transporter activity, channel activity and phospholipid binding (Figure 5G).

#### **Clinical models based on PD-L1-related lncRNAs and clinical characteristics can predict the prognosis of glioma patients**

Based on the TCGA dataset, we developed a clinical model that combines risk scores and clinical case characteristics, and created a nomogram (Figure 6A). The model's predicted 3-year and 5-year OS closely align with the observed outcomes (Figure 6B), suggesting that the model has a specific predictive value. Taking account of the nomogram, the patients were divided into two new risk groups. Survival analysis showed a conspicuously different prognosis in the two groups ( $P < 0.0001$ ) (Figure 6C). The 3-year AUC under the ROC curve is 0.915, while the 5-year AUC is 0.881. The robust specificity and sensitivity of our model demonstrate its superior predictive power compared to using risk score alone

when incorporating clinicopathological characteristics (Figure 6D).

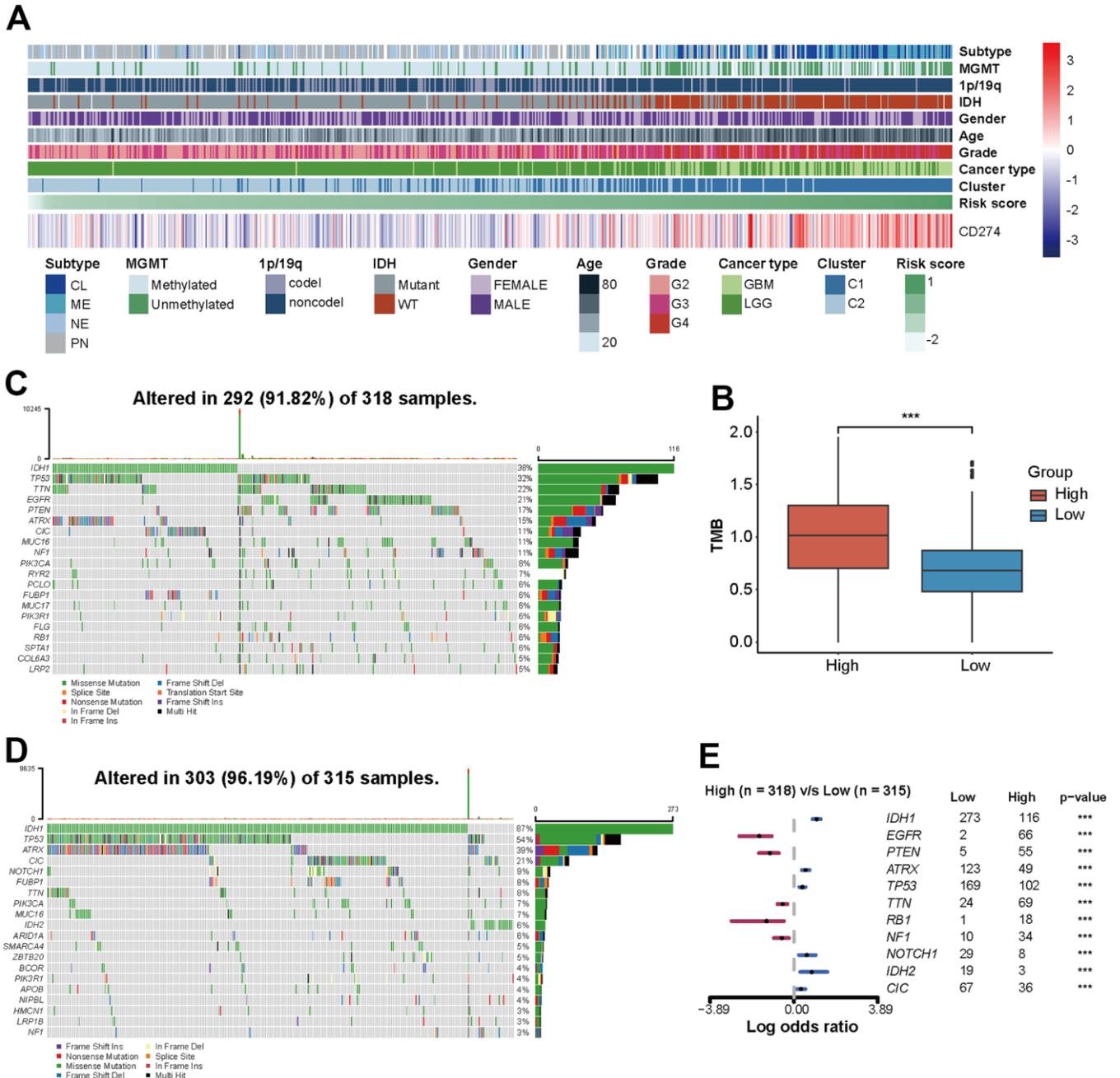
#### **LINC01271 is significantly associated with features of glioma**

According to previous studies, Diermeier's team found that lncRNA Mammary Tumor-associated RNA 25 (MaTAR25) contributed to the malignancy of breast tumor cells *in vitro* and *in vivo*. LINC01271 was identified as a homologous human lncRNA of MaTAR25, and they proved that increased LINC01271 expression was relevant to poor prognosis and metastasis [23, 24]. The role of LINC01271 in gliomas has not been reported, so we selected LINC01271 as the target to investigate its impact on glioma development and prognosis.

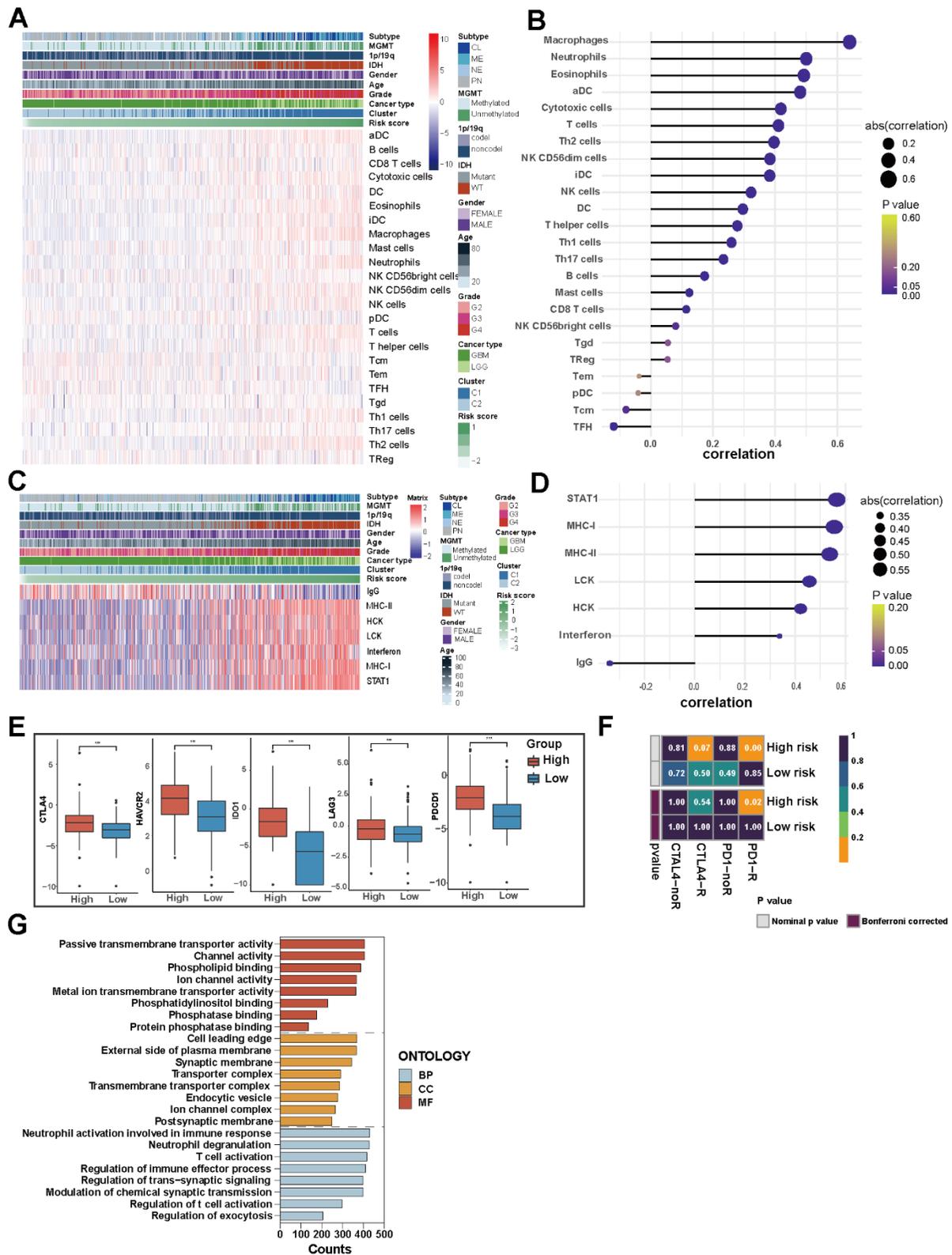
The glioma patients in two data sets were separated into two different groups according to LINC01271 expression level. In the TCGA dataset, the overall OS, DSS, and PFI of patients with high LINC01271 expression were worse than another group ( $p < 0.0001$ ) (Figure 7A and Supplementary Figure 6A, 6D), but from the perspective of cancer type, the prognostic difference was only reflected in LGG (LGG: OS  $p = 0.0042$ , DSS  $p = 0.0083$ , PFI  $p < 0.0001$ ; GBM: OS  $p = 0.27$ , DSS  $p = 0.33$ , PFI  $p = 0.46$ ) (Figure 7B, 7C and Supplementary Figure 6B, 6C, 6E, 6F), which is in line with the results in the CGGA set (Supplementary Figure 6G–6I).

Then we studied the correlation between LINC01271 expression and the characteristics of immunity, inflammation, and clinicopathology. The results showed more infiltrates of M1 subtype macrophages, CD8<sup>+</sup> T cells and T follicular helper cells in the high LINC01271 expression group (Figure 7D and Supplementary Figure 6J), and the infiltration degree of eosinophils, macrophages, neutrophils and T cells

was positively correlated with the LINC01271 expression, while pDC was negatively correlated with the expression of LINC01271 (Figure 7E). Low LINC01271 expression group showed more infiltration of NK cells and Mast cells (Supplementary Figure 6J). The same analysis was conducted on CGGA database samples, yielding a heatmap with comparable findings (Supplementary Figure 6K).



**Figure 4. The risk score of PD-L1-related lncRNAs reflects the clinical, pathological, and genetic characteristics of glioma. (A)** Heatmap displaying risk scores, clinical information, and pathological features of glioma patients. **(B)** Analysis of tumor mutation burden (TMB). **(C, D)** 20 mutation sites in high-risk and low-risk groups. **(E)** Risk score-related somatic mutations.



**Figure 5. Immune and inflammatory characteristics of glioma and the biological mechanism of PD-L1-related lncRNAs. (A)** Heatmap displaying the characteristics of immune cells panel. **(B)** The correlation between immune cell infiltration and risk score. **(C)** Heatmap displaying the characteristics of inflammation panel. **(D)** The correlation between inflammation characteristics and risk score. **(E)** 5 immune checkpoints' expression in high-risk and low-risk groups in TCGA. **(F)** Patients' predicted response to anti-CTLA-4 and anti-PD-1 therapy. **(G)** GO analysis of PD-L1-related lncRNAs.

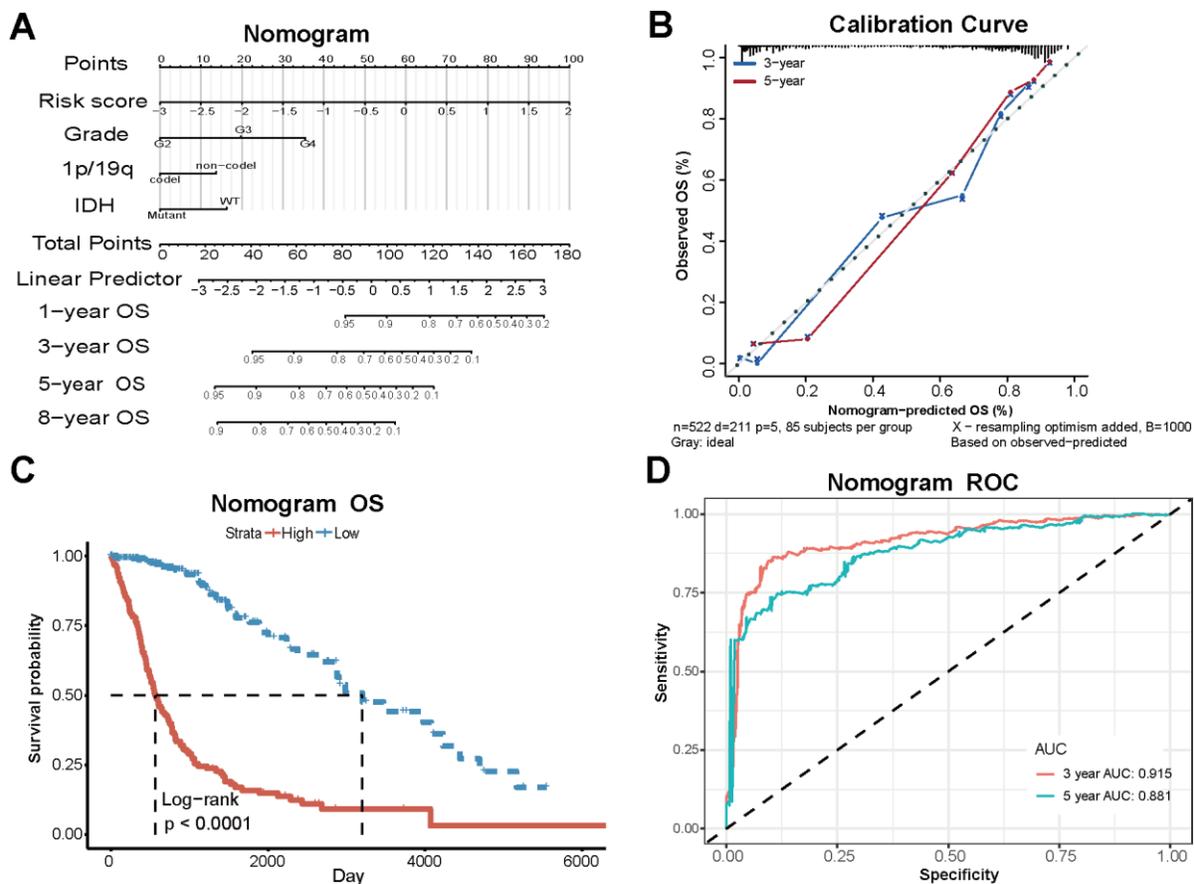
Inflammatory characteristics analysis shows that STAT1, MHC-II, LCK, MHC-I, interferon, and HCK are associated with high LINC01271 expression. In contrast, patients with low LINC01271 expression have higher IgG expression (Figure 7F, 7G). These findings are consistent with the previous analysis results based on the 6 PD-L1-related lncRNAs.

As for clinicopathological characteristics, we found that CL, ME subtype, MGMT non-methylation, 1p19q non-codel, IDH wild type, higher age, higher WHO grade, and higher GBM patients' proportion are associated with high LINC01271 expression, which illustrates that LINC01271 is a risk factor for prognosis (Figure 7D, 7F).

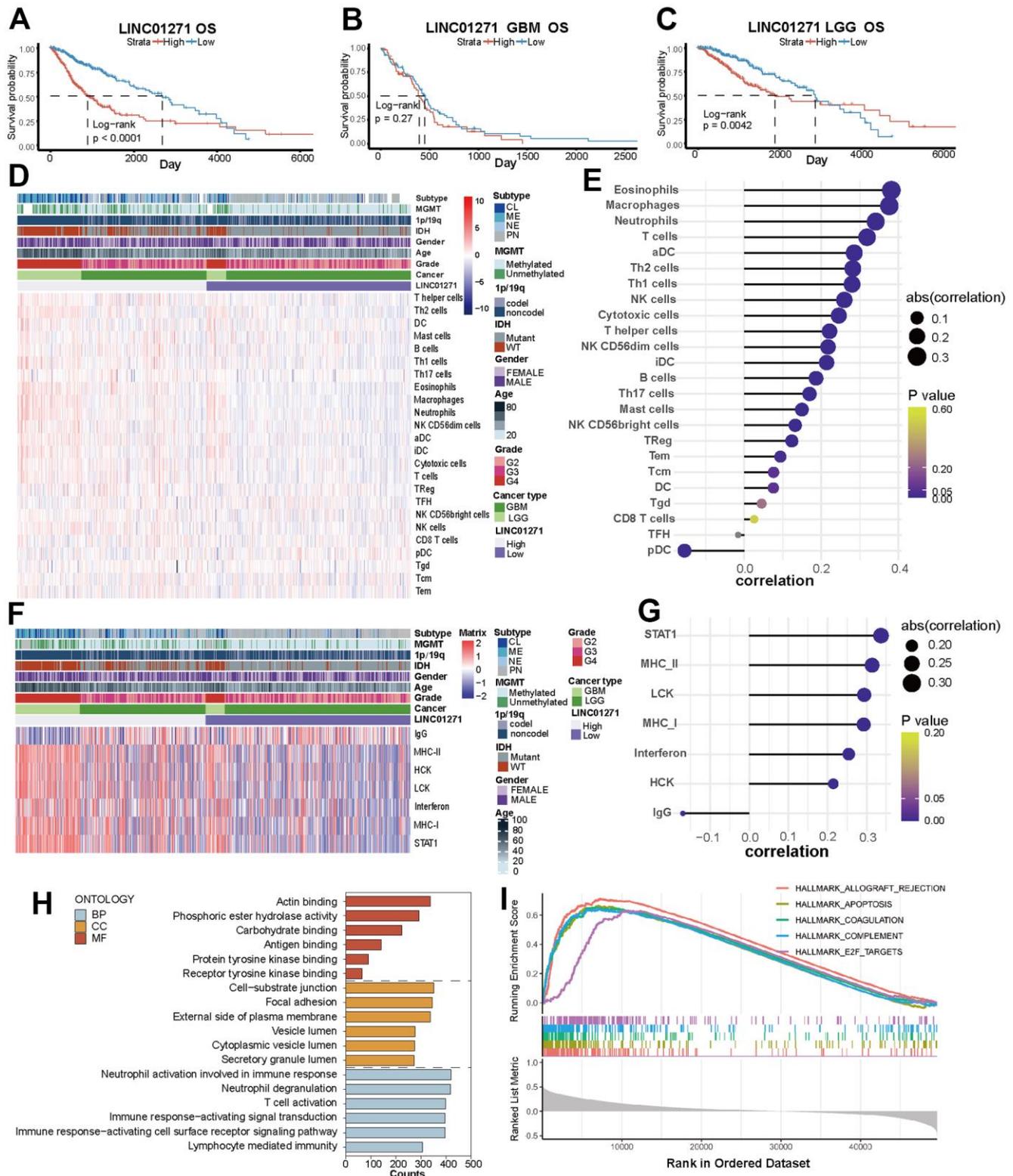
GSEA analysis revealed that LINC01271 mainly participated in the same biological process as the 6 PD-L1-related lncRNAs model, but concentrated on different cellular components, including the cell-substrate junction, focal adhesion, external side of the plasma membrane and others, and were related to

molecular functions like acting binding and phosphoric ester hydrolase activity (Figure 7H). The expression of LINC01271 is also in connection with some hallmark gene sets, such as APOPTOSIS, COAGULATION, COMPLEMENT, and E2F TARGETS (Figure 7I), suggesting the immune pathways related to characteristic genes, and perhaps related to the mechanism of LINC01271's biological role in glioma.

We further conducted a prediction of the patients' sensitivity to TMZ treatment, which is currently one of the most commonly used chemotherapy drugs for glioma. With data derived from the GDSC database, we built a predictive model and estimated the half-maximal inhibitory concentration (IC50) value of TMZ in each patient. Unfortunately, there was no statistical difference in IC50 when grouping according to LINC01271 expression (Supplementary Figure 6L). Finally, we compared the LINC01271 expression in tumor and normal tissues using the TCGA database, but no statistically significant difference was observed (Supplementary Figure 6M).



**Figure 6. Clinical models based on PD-L1-related lncRNAs and clinical characteristics.** (A) The nomogram combining risk scores and clinical case characteristics according to TCGA data. (B) The calibration curves showing the predicted and actual observed OS rates. (C) Survival analysis of new high-risk group and low-risk group. (D) The ROC curves and AUC values of the nomogram.



**Figure 7. Correlations between LINC01271 expression and glioma prognosis, immune and inflammatory features.** (A–C) OS of glioma patients with high and low LINC01271 expression in TCGA. (D) Heatmap displaying the characteristics of immune cells panel. (E) Correlation between immune cell infiltration and LINC01271 level. (F) Heatmap displaying the characteristics of inflammation panel. (G) Correlation between inflammation characteristics and LINC01271 level. (H) GO analysis of LINC01271 related genes. (I) Hallmark analysis of LINC01271 related genes.

## Knockout of LINC01271 effectively suppressed the proliferation and migration of glioma cells

We silence *LINC01271* gene expression in U251 and U87 to detect proliferation and migration. RT-qPCR confirmed the silencing efficiency of siRNA, si-LINC01271-334 and si-LINC01271-1564 significantly reduced the expression of *LINC01271*, si-LINC01271-1196 also had the same trend (Figure 8A). CCK-8 assay, plate cloning assay, and Edu assay showed that tumor cell proliferation and colony formation were inhibited after silencing *LINC01271* (Figure 8B–8D). Transwell migration assay suggested inhibition of tumor cell migration after *LINC01271* knockout (Figure 8E). In conclusion, knockout of *LINC01271* visibly suppressed the proliferation and migration of glioma cells, proving that LINC01271 positively regulates the progression of glioma and lncRNA LINC01271 is expected to become a new target for the treatment.

## DISCUSSION

The high fatality rate, ease of development, and propensity for metastasis make glioma a formidable threat to human health. It is of great clinical significance to search for prognostic biomarkers and immunotherapy targets for glioma. The present study utilized data from public databases TCGA and CGGA to collect glioma patients' information, and identified 6 PD-L1-related lncRNAs for the construction of a risk score model. The findings indicated that glioma patients with a high-risk score exhibited significantly poorer prognosis and more malignant tumor pathological characteristics and microenvironment. Considering previous research results, we chose LINC01271 for further study separately. We revealed that the high LINC01271 expression level was relevant to poor glioma prognosis (especially LGG), aggressive clinicopathological features, and immunoinflammatory characteristics. Subsequent *in vitro* cell experiments have confirmed that knocking out LINC01271 effectively suppresses the proliferation and migration of glioma cells, indicating that LINC01271 holds great potential as a biomarker for glioma and as an immunotherapy target.

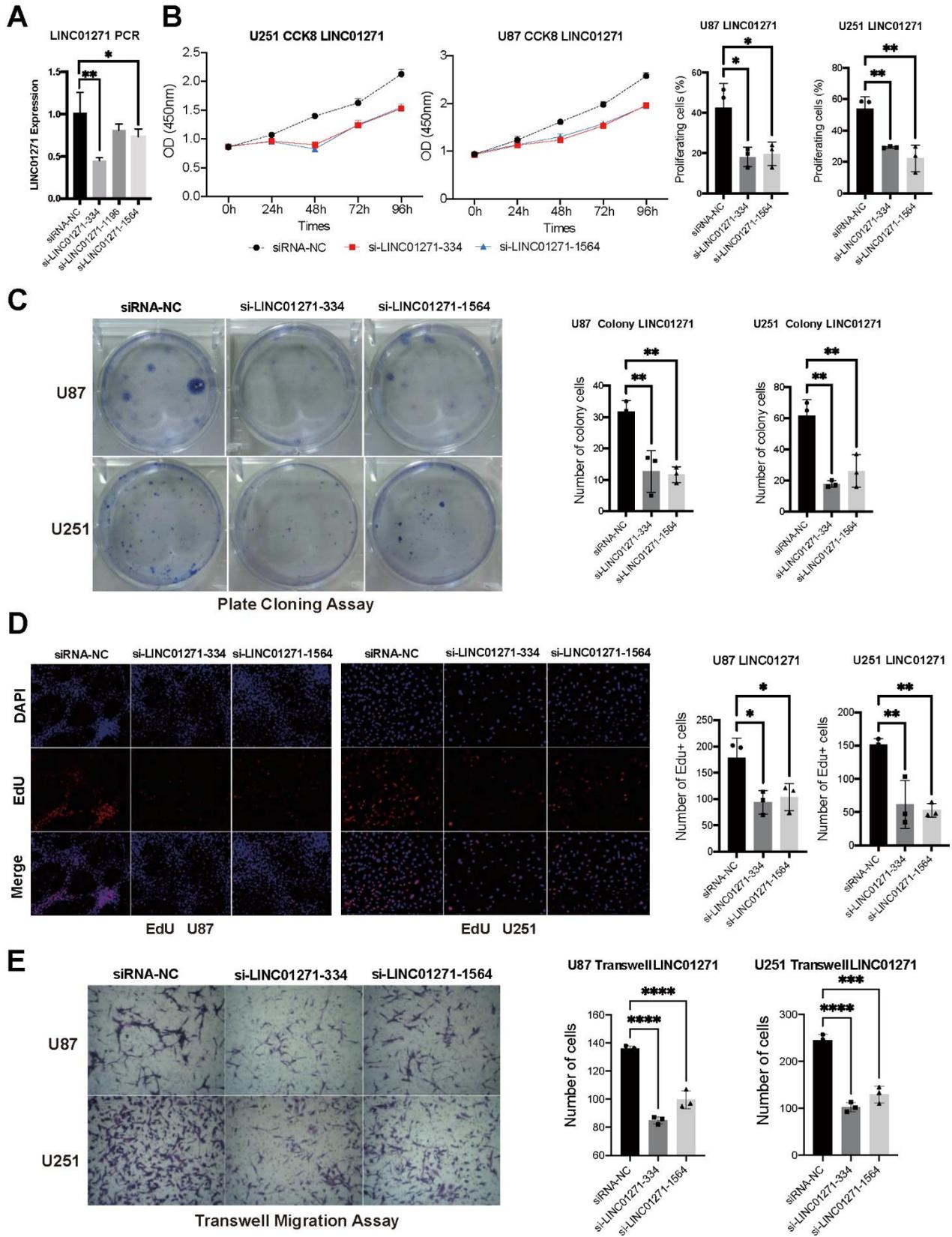
The PD-1 receptor is encoded by the *PDCDI* gene, while PD-L1 is encoded by the *CD274* gene; both are expressed on activated T cells. Many tumors use PD-L1/PD-1 signals to achieve immune escape [25]. The usage of ICIs to block PD-L1/PD-1 has made significant progress in a variety of tumors, including lung cancer. However, due to differences in the immune microenvironment and tumor cell immunogenicity, there is no encouraging progress in the treatment of glioblastoma with ICIs [26]. Currently ongoing research

confirm that PD-L1 expression is associated with the glioma prognosis. Zhu et al. found that the expression of PD-L1 in gliomas tended to be dependent on tumor grade, with higher levels associated with shorter overall survival [25]. Previous studies have also demonstrated that PD-1 and PD-L1 expression are negative predictors of GBM prognosis [27]. Our research showed that patients with high-risk scores exhibited elevated expression levels of 5 immune checkpoints and displayed a greater propensity to respond favorably to anti-PD-1 treatment, which is consistent with previous studies. This may be related to the more malignant GBM patients, especially those with the mesenchymal subtype [28], as they tend to express higher levels of PD-L1 in their tumor cells.

The matrix components in the TIME influence the effect of PD-L1 expression on prognosis. Qian et al. found that the level of IFN- $\gamma$  in mouse gliomas is positively correlated with the PD-L1 expression, and scores based on IFN- $\gamma$ -induced genes can serve as supplementary prognostic indicators for anti-PD-1/PD-L1 therapy [29]. Moreover, PD-L1 can also be expressed in GBM extracellular vesicles, playing an immunosuppressive role through monocytes rather than T cells [30]. This suggests that the immunosuppressive signalling pathway involved in PD-L1 is complex.

The expression of CTLA-4, another immune checkpoint, competes with the costimulatory receptor CD28 for binding its ligands CD80 and CD86 [31], which also obviously increased after T cell activation. Reardon et al. revealed that the combination therapy targeting CTLA-4 and PD-1 cured 70% of glioma mice, higher than the cure rate of PD-1 inhibitor alone, and induced tumor-specific memory effect to prevent recurrence [32]. This demonstrated that CTLA-4 inhibitory therapy might also improve the prognosis of patients with glioma. Still, unfortunately, there is no significant difference in response to the anti-CTLA-4 treatment of the two groups in our experiment. The above research suggests that immune checkpoint suppression therapy for glioma may involve more internal mechanisms.

Our previous research found that the prognosis of LGG patients with epilepsy surrounded by a diverse immune microenvironment is different [33]. The interaction between immune cells and inflammatory factors within the microenvironment modulates tumor progression [34]. The microenvironment of glioma promotes angiogenesis and inflammation, leading to high permeability of the blood-brain barrier (BBB) and facilitating immune cell recruitment. Our study found that macrophages (M0, M1) were more highly expressed in the high-risk score group. Macrophages



**Figure 8. Validation of LINC01271 in positively regulating glioma cell proliferation and migration *in vitro*.** (A) *LINC01271* expression after silencing. (B–D) CCK-8, plate clone assay, EdU verified the proliferation of glioma cells after *LINC01271* silencing. (E) Glioma cell migration after *LINC01271* silencing.

are traditionally divided into M1 and M2 subtypes: M1, which is pro-inflammatory and secretes inflammatory factors such as IL-6, IL-8, and TNF- $\alpha$ , while M2 releases anti-inflammatory factors such as IL-10 and TGF- $\beta$ . M1 macrophage has anti-tumor properties while M2 type positively regulates the growth and migration of tumor cells [35]. In previous studies, M2 macrophages have been found to promote tumor growth and are associated with shorter survival times [36]. M2 polarization of macrophages was also significantly correlated with PD-L1 high expression [36]. Our study showed that although M2 macrophages in different risk score groups had a high expression, there was no significant difference. This may be because lncRNA regulation of gliomas does not confine to the polarization of macrophages. The glioma microenvironment comprises non-tumor cells, such as infiltrating or resident immune cells, other glial cells, vascular cells, etc., particularly tumor-associated macrophages (TAMs) [37]. TAM promotes the development of tumors through activating inflammatory response, destructing the BBB [38, 39], regulating glioma metabolism [40], and promoting immunosuppression [41]. GBM often resists ICIs treatment; the therapy that targets PD-L1 expression may overcome resistance to immune checkpoint blockade [42]. In addition to TAMs, T cells occupy most of the lymphocytes. Han et al. found that the abundance of CD8<sup>+</sup> T cells was negatively correlated with tumor grade in gliomas, which is contrary to CD4<sup>+</sup> T cells. The high presence of CD4<sup>+</sup> T cells and low presence of CD8<sup>+</sup> T cells represent poor patient prognosis [43]. As a subset of CD4<sup>+</sup> T cells, Treg has high diversity and plasticity, and plays a vital role in immune tolerance through the CTLA-4 and PD-L1 pathways [44]. However, in this study, both CD8<sup>+</sup> T cells and Treg cells existed more in the high-risk group, while the latter group had higher naive CD4<sup>+</sup> T cells. The observed phenomenon could potentially be attributed to the stratification based on the risk score of lncRNAs associated with PD-L1 and the complexity of immunity.

In terms of inflammatory characteristics, we found that irrespective of whether it is based on the clinical model comprising 6 PD-L1-related lncRNAs or solely LINC01271 expression, the high-scoring group exhibited heightened expression levels of STAT1, MHC, HCK, LCK, and Interferon molecules. Studies have shown that high expression of signal transducer and activator of transcription 1 (STAT1) in brain gliomas is associated with poorer OS in patients [45]. In terms of mechanism, STAT1 may mediate epithelial-mesenchymal transition through the wnt/ $\beta$ -catenin signalling pathway to increase glioblastoma growth and migration [46]. Moreover, STAT1 production decreased in LGG with IDH mutant,

and the expression of cytotoxic T lymphocyte-related genes and IFN- $\gamma$  inducible chemokines is reduced [47], which may partially explain that the prognosis of IDH mutant is better than wild-type patients. The immunogenicity of tumors can be determined by the expression of major histocompatibility complex (MHC)-I. Previous studies revealed that gliomas reduce T cells' recognition and killing by downregulating MHC-I and antigen-processing machinery (APM) components, thus evading immune surveillance [48, 49]. However, TMZ increases MHC-I expression through the NF- $\kappa$ B pathway to achieve a therapeutic effect [50].

Recent studies have shown that gene mutations regulated by epigenetics have become a significant feature of glioma subtypes [51]. Previous studies have used 1p/19q chromosomal codeletion, mutations of IDH1/2, and MGMT promoter methylation as three molecular biomarkers to guide the classification and treatment decisions of glioma [52]. Verhaak et al. classified gliomas into neural (NE), proneural (PN), classical (CL), and mesenchymal (MES) based on the histological type [53]. Among them, patients in the PN subtype have the best median survival time; MES subtypes have the worst prognosis with the most apparent inflammation features (genes of TNF and NF- $\kappa$ B pathways are significantly up-regulated) [54]. MGMT promoter unmethylated makes tumor cells less likely to benefit from DNA alkylation agents such as temozolomide or nitrosourea, and such patients have a shorter median survival [55]. 1p/19q non-codeletion, IDH wild type, MGMT promoter unmethylated, plus CL and MES subtype and higher WHO grade are associated with poor outcomes of glioma [56]. Currently, molecular biology markers are recommended for routine examinations in the pathological diagnosis of gliomas according to guidelines, and targeted therapies based on genetic and immune characteristics are also being studied with enthusiasm. This study found that whether it was based on the risk-scoring model of 6 PD-L1-related lncRNAs or grouped only by LINC01271 expression level, the results showed the same results as previous studies, which illustrated the rationality and feasibility of the model.

As a non-coding regulatory RNA, lncRNA has been proven to be a robust biomarker in many diseases in recent years. The present study selected 6 PD-L1-related lncRNAs, based on which a clinical model was developed with improved efficiency in predicting glioma prognosis. Diermeier's team screened a series of lncRNAs that were overexpressed in mouse breast tumors compared to normal mammary epithelial cells and named them Mammary Tumor-associated RNA (MaTAR) [24]. Among them, MaTAR25 functions by regulating the expression of the gene Tensin1 via purine-rich element binding protein B (PURB),

affecting tumor proliferation, migration and invasion. Subsequently, they identified LINC01271 as a direct homolog of MaTAR25 in humans and demonstrated that increased LINC01271 expression was associated with higher levels of malignancy [23]. Here, we show that LINC01271 expression in human gliomas is also associated with poor prognosis, which may be a new therapeutic target.

Unfortunately, few studies on the other 5 lncRNAs. Wang et al. found that MIR4500HG, among five immune-related genes, was a predictor of poor prognosis and early recurrence in stage Ia-b non-small cell lung cancer (NSCLC) [34]. Liang et al. reported the association of AC011891.3 with poor prognosis of prostate adenocarcinoma, which may serve as a potential immunotherapeutic target [57]. LINC02594, also known as CTC-501O10.1, exhibits up-regulation in the plasma of patients diagnosed with gastric cancer and thus holds potential as a biomarker for detecting this malignancy [58]. AL357055.3(lnc-SLC16A1-2:1) has not been reported, but its homologous protein SLC16A1 is a confirmed poor prognosis marker in various malignant tumors, possibly due to its involvement in glycolysis and glucose metabolism synthesis [59, 60]. AL355974.3, also referred to as lnc-COL4A1-7:1 in the human non-coding RNA library, lacks functional characterization in existing literature. There is still much to be unveiled regarding the mechanism by which lncRNA contributes to tumor progression.

GSEA analysis showed that LINC01271 is associated with the enrichment of hallmark gene sets, such as APOPTOSIS, COAGULATION, COMPLEMENT and E2F TARGETS. This suggests that LINC01271 may play a role in tumors through immune pathways. E2F transcription factors (E2Fs) are a set of genes encoding transcription factor families. They participate in the core transcription that drives the cell cycle process through the cyclin-dependent kinase (CDK)- retinoblastoma (RB)-E2F axis [61]. E2Fs have been implicated in the progression of breast cancer [62], lung cancer [63], chromophobe renal cell carcinoma [64], among others, however, the specific contributions of individual family members remain unclear. Liao et al. found that E2Fs expression increased in the human brain and central nervous system tumors and was associated with poorer OS of LGG and GBM and increased immune cell infiltration in the two tumors, proving that E2Fs may become promising prognostic biomarkers and immunotherapy targets of gliomas [65], which may also have a connection with the mechanism of LINC01271 in glioma.

This study has the following limitations: the sample data only comes from the TCGA and CGGA databases, and

the sample size is not large, which may not objectively reflect the characteristics of glioma. Only bioinformatics analysis and cell experiments *in vitro* were obtained to verify the effect of LINC01271 on glioma without exploring specific downstream mechanisms. We will improve the *in vitro* experiment and in-depth study of possible molecular mechanisms in the next step.

## CONCLUSIONS

This study established a glioma risk score model based on 6 PD-L1-related lncRNAs from an immunological perspective, which can effectively reflect the clinical characteristics, prognosis, pathological features, immunoinflammatory microenvironment and immunotherapy reactivity of glioma patients. The application value of LINC01271 in predicting the prognosis and treatment of glioma is evident. In conclusion, we have identified a potential risk stratification approach and biomarker that could offer early prognosis prediction and personalized immunotherapy guidance for glioma, thereby contributing to the optimization of secondary and tertiary prevention.

## MATERIALS AND METHODS

### Data collection

RNA transcript data (Workflow Type: HTSeq-Counts) with glioma were respectively downloaded from the TCGA database and CGGA database [66], along with baseline clinical characteristics (age, gender, WHO grade, cancer type and subtype) and clinical pathological features (isocitrate dehydrogenase (IDH) Mutation status, 1p/19q gene status, DNA methylation of O6-methylguanine-DNA-methyltransferase (MGMT) and somatic mutation). The data was trained in the TCGA database and validated in the CGGA database.

### Immune and inflammatory characteristics

The CIBERSORT deconvolution algorithm was performed to explore the immune microenvironment [67]. Inflammation characteristics were reflected by gene set variation analysis (GSVA) with the expression of 7 kinds of inflammation-related molecules, including MHC-I, MHC-II, STAT1, LCK, HCK, IgG and interferon [68]. Beyond that, draw correlograms and heatmaps respectively by the “corrgram” and the “pheatmap” R packages to confirm the correlation.

### Risk score model and consensus clustering analysis

According to the expression of 6 PD-L1-related lncRNAs in samples, the risk score model was established with the

formula:  $risk\ score = \sum_{i=1}^n (Gene\ exp\ r_i \times \beta_i)$ . Patients were divided into high/low-risk score groups with the median risk score. The R package “rms” was adopted to construct a nomogram for a new risk model; then we evaluated the efficiency of which with the AUC value of the receiver operating curve (ROC). Consensus Clustering Analysis was taken on to further verify the rationality of the risk score model [69].

### Functional enrichment analysis

To further analyze the function of signature lncRNAs, gene ontology (GO) analysis was applied to study the essential biological functions of signature lncRNAs that are conspicuously related to risk scores. GSEA determined the immune pathways related to signature lncRNAs based on the hallmark gene set [70, 71].

### Response to immunotherapy

The response of glioma to ICIs was predicted by the tumor immune dysfunction and exclusion (TIDE) algorithm [72]. The R package “pRRophetic” was adopted to predict the sensitivity to temozolomide (TMZ) treatment with data from the Genomics of Drug Sensitivity in Cancer (GDSC) database [73].

### Cell transfection

Human glioma cells U251 and human glioblastoma cells U87 were purchased from the iCell Bioscience Inc. (Shanghai, China) company. According to the transcription of the LINC01271 gene, three kinds of siRNA (si-LINC01271-334, si-LINC01271-1196, si-LINC01271-1564) were designed to silence LINC01271 gene expression in cell transfection experiments using Lip2000. Examine the silencing efficiency through quantitative real-time PCR (PCR primer information for LINC01271 and the synthesis information for related siRNA are shown in Supplementary Tables 2, 3) [74].

### Cell counting kit-8 assay

Cells were inoculated into a 96-well plate at a density of  $5 \times 10^3$  cells per well overnight and were incubated for 4 hours after adding 30ul CCK-8 solution. A spectrophotometer (Bio-Tek) measured the absorbance at 450nm to evaluate cell proliferation.

### EdU image kit

Cells were incubated in 50uM 5-Ethynyl-2'-deoxyuridine (EdU) medium for 2 hours, fixed with 4% paraformaldehyde, and permeated with 0.5% TritonX-100 penetrant. Apollo and DAPI were used for staining,

imageJ merge images were used to determine cell proliferation, and quantitative analysis was performed by flow cytometry.

### Colony formation and cell migration assay

For clone formation assay, the treated cells were inoculated in 6-well plates (200 cells per well) for 2-3 weeks at 37° C, with 5% CO<sub>2</sub> and saturated humidity, rinsed twice with phosphate buffered saline (PBS), fixed with 4% paraformaldehyde for 15min, stained with crystal violet staining solution after fixed and counted colony. To evaluate cell migration ability, the cells were added to the Transwell upper chamber by  $2 \times 10^6$  cells/ml and suspended in 0.2ml serum-free medium for 48h. The migrating cells were fixed, stained, and counted after incubation.

### Statistical analysis

All statistical analysis was accomplished with R software (version 3.5.3). The Student's t-test, Wilcoxon rank sum test and Pearson's Chi-squared test were used for comparing differences in continuous or categorical variables between groups. The Least Absolute Shrinkage and Selection Operator (LASSO) analysis, and multivariate and univariate Cox regression analysis were performed to screen for variables of interest.  $P < 0.05$  was considered statistically significant. Survival analysis is carried out by the Kaplan-Meier method with the “survival” R package (version 3.6) and calculated OS, Disease-Specific Survival (DSS), and Progression-Free Interval (PFI) as prognostic indicators.

### Consent for publication

All authors give their consent to publish this manuscript.

### Availability of data

All data are available in public repositories, which are listed in the main context.

### Abbreviations

lncRNA: Long non-coding RNAs; TCGA: The Cancer Genome Atlas; CGGA: Chinese Glioma Genome Atlas; TIDE: Tumor immune dysfunction and exclusion; GDSC: Genomics of Drug Sensitivity in Cancer; LASSO: Least Absolute Shrinkage and Selection Operator; GSVA: Gene set variation analysis; GO: Gene ontology; CCK-8: Cell counting kit-8; EdU: 5-Ethynyl-2'-deoxyuridine; Rt-qPCR: Real-time polymerase chain reaction; PCA: Principal component analysis; LGG: Lower-grade gliomas; GBM: Glioblastomas; WHO: World Health Organization; OS: Overall Survival; DSS:

Disease-Specific Survival; PFI: Progression-Free Interval; ICI: Immune checkpoint inhibitors; PD-1/PDCD1: Programmed cell death protein 1; PD-L1: Programmed cell death ligand-1; CTLA-4: Cytotoxic T lymphocyte antigen 4; HAVCR2: Hepatitis A virus cellular receptor 2; IDO1: Indolamine 2,3-dioxygenase 1; LAG3: Lymphocyte activation gene 3; MGMT: O6-methylguanine-DNA-methyltransferase; IDH: Isocitrate dehydrogenase; ROC: Receiver operating curve; AUC: Area under curve; TMZ: Temozolomide; TIME: Tumor immune microenvironment; TMB: Tumor mutation burden; TAM: Tumor-associated macrophages; PFS: Progression-free survival; IC50: Half-maximal inhibitory concentration; BBB: Blood-brain barrier; HR: Hazard ratio; APM: Antigen-processing machinery; NE: Neural; PN: Proneural; CL: Classical; MES: Mesenchymal; NSCLC: non-small cell lung cancer; E2Fs: E2F transcription factors; CDK: Cyclin-dependent kinase; RB: Retinoblastoma.

## AUTHOR CONTRIBUTIONS

ZX, RT: Conceptualization, Methodology, Software, Investigation, Formal Analysis, Writing-Original Draft; FL, HZ, ZD, ZW: Data Curation, Resources, Visualization, Investigation, Writing-Original Draft; PL, HS, GX: Data Curation, Software, Validation, Writing-Original Draft; JF, QC: Supervision, Funding Acquisition, Writing-Review and Editing. All authors read and approved the final manuscript.

## ACKNOWLEDGMENTS

We are grateful to the High Performance Computing Center of Central South University for partial support of this work. The authors express gratitude to the public databases, websites, and software used in the paper.

The content of this manuscript has previously appeared online in a preprint [75].

## CONFLICTS OF INTEREST

The authors declare that the research was conducted without any commercial or financial relationships that could be construed as a potential conflict of interest.

## FUNDING

This work was supported by the National Natural Science Foundation of China Youth Project (No. 82001392), the Science Foundation of AMHT Group (NO.2022YK04), the Hunan Provincial Natural Science Foundation of China (NO.2022JJ30544) and the Fundamental and Applied Research Project of Hengyang Science and Technology Bureau (202250045302).

## REFERENCES

1. Chen R, Smith-Cohn M, Cohen AL, Colman H. Glioma Subclassifications and Their Clinical Significance. *Neurotherapeutics*. 2017; 14:284–97. <https://doi.org/10.1007/s13311-017-0519-x> PMID:28281173
2. Ostrom QT, Patil N, Cioffi G, Waite K, Kruchko C, Barnholtz-Sloan JS. CBTRUS Statistical Report: Primary Brain and Other Central Nervous System Tumors Diagnosed in the United States in 2013-2017. *Neuro Oncol*. 2020; 22:iv1–v96. <https://doi.org/10.1093/neuonc/noaa200> PMID:33123732
3. Bush NA, Chang SM, Berger MS. Current and future strategies for treatment of glioma. *Neurosurg Rev*. 2017; 40:1–14. <https://doi.org/10.1007/s10143-016-0709-8> PMID:27085859
4. Glaser T, Han I, Wu L, Zeng X. Targeted Nanotechnology in Glioblastoma Multiforme. *Front Pharmacol*. 2017; 8:166. <https://doi.org/10.3389/fphar.2017.00166> PMID:28408882
5. Iams WT, Porter J, Horn L. Immunotherapeutic approaches for small-cell lung cancer. *Nat Rev Clin Oncol*. 2020; 17:300–12. <https://doi.org/10.1038/s41571-019-0316-z> PMID:32055013
6. DeOcesano-Pereira C, Machado RAC, Chudzinski-Tavassi AM, Sogayar MC. Emerging Roles and Potential Applications of Non-Coding RNAs in Glioblastoma. *Int J Mol Sci*. 2020; 21:2611. <https://doi.org/10.3390/ijms21072611> PMID:32283739
7. Wei SC, Duffy CR, Allison JP. Fundamental Mechanisms of Immune Checkpoint Blockade Therapy. *Cancer Discov*. 2018; 8:1069–86. <https://doi.org/10.1158/2159-8290.CD-18-0367> PMID:30115704
8. Emens LA. Breast Cancer Immunotherapy: Facts and Hopes. *Clin Cancer Res*. 2018; 24:511–20. <https://doi.org/10.1158/1078-0432.CCR-16-3001> PMID:28801472
9. Beermann J, Piccoli MT, Viereck J, Thum T. Non-coding RNAs in Development and Disease: Background, Mechanisms, and Therapeutic Approaches. *Physiol Rev*. 2016; 96:1297–325. <https://doi.org/10.1152/physrev.00041.2015> PMID:27535639
10. Arun G, Diermeier SD, Spector DL. Therapeutic Targeting of Long Non-Coding RNAs in Cancer. *Trends Mol Med*. 2018; 24:257–77.

- <https://doi.org/10.1016/j.molmed.2018.01.001>  
PMID:[29449148](https://pubmed.ncbi.nlm.nih.gov/29449148/)
11. Kelemen E, Danis J, Göblös A, Bata-Csörgő Z, Széll M. Exosomal long non-coding RNAs as biomarkers in human diseases. *EJIFCC*. 2019; 30:224–36. PMID:[31263395](https://pubmed.ncbi.nlm.nih.gov/31263395/)
  12. Singh D, Khan MA, Siddique HR. Emerging role of long non-coding RNAs in cancer chemoresistance: unravelling the multifaceted role and prospective therapeutic targeting. *Mol Biol Rep*. 2020; 47:5569–85. <https://doi.org/10.1007/s11033-020-05609-x> PMID:[32601922](https://pubmed.ncbi.nlm.nih.gov/32601922/)
  13. Zhang G, Pian C, Chen Z, Zhang J, Xu M, Zhang L, Chen Y. Identification of cancer-related miRNA-lncRNA biomarkers using a basic miRNA-lncRNA network. *PLoS One*. 2018; 13:e0196681. <https://doi.org/10.1371/journal.pone.0196681> PMID:[29715309](https://pubmed.ncbi.nlm.nih.gov/29715309/)
  14. Lu M, Zhan H, Liu B, Li D, Li W, Chen X, Zhou X. N6-methyladenosine-related non-coding RNAs are potential prognostic and immunotherapeutic responsiveness biomarkers for bladder cancer. *EPMA J*. 2021; 12:589–604. <https://doi.org/10.1007/s13167-021-00259-w> PMID:[34950253](https://pubmed.ncbi.nlm.nih.gov/34950253/)
  15. Li N, Zhan X. Identification of clinical trait-related lncRNA and mRNA biomarkers with weighted gene co-expression network analysis as useful tool for personalized medicine in ovarian cancer. *EPMA J*. 2019; 10:273–90. <https://doi.org/10.1007/s13167-019-00175-0> PMID:[31462944](https://pubmed.ncbi.nlm.nih.gov/31462944/)
  16. Li N, Zhan X. Anti-parasite drug ivermectin can suppress ovarian cancer by regulating lncRNA-EIF4A3-mRNA axes. *EPMA J*. 2020; 11:289–309. <https://doi.org/10.1007/s13167-020-00209-y> PMID:[32549918](https://pubmed.ncbi.nlm.nih.gov/32549918/)
  17. Huang W, Shi Y, Han B, Wang Q, Zhang B, Qi C, Liu F. lncRNA GAS5-AS1 inhibits glioma proliferation, migration, and invasion via miR-106b-5p/TUSC2 axis. *Hum Cell*. 2020; 33:416–26. <https://doi.org/10.1007/s13577-020-00331-z> PMID:[32072565](https://pubmed.ncbi.nlm.nih.gov/32072565/)
  18. Shen F, Chang H, Gao G, Zhang B, Li X, Jin B. Long noncoding RNA FOXD2-AS1 promotes glioma malignancy and tumorigenesis via targeting miR-185-5p/CCND2 axis. *J Cell Biochem*. 2019; 120:9324–36. <https://doi.org/10.1002/jcb.28208> PMID:[30520141](https://pubmed.ncbi.nlm.nih.gov/30520141/)
  19. Suo ST, Gong P, Peng XJ, Niu D, Guo YT. Knockdown of long non-coding RNA VIM-AS1 inhibits glioma cell proliferation and migration, and increases the cell apoptosis via modulation of WEE1 targeted by miR-105-5p. *Eur Rev Med Pharmacol Sci*. 2020; 24:6834–47. [https://doi.org/10.26355/eurrev\\_202006\\_21673](https://doi.org/10.26355/eurrev_202006_21673) PMID:[32633376](https://pubmed.ncbi.nlm.nih.gov/32633376/)
  20. Tan SK, Pastori C, Penas C, Komotar RJ, Ivan ME, Wahlestedt C, Ayad NG. Serum long noncoding RNA HOTAIR as a novel diagnostic and prognostic biomarker in glioblastoma multiforme. *Mol Cancer*. 2018; 17:74. <https://doi.org/10.1186/s12943-018-0822-0> PMID:[29558959](https://pubmed.ncbi.nlm.nih.gov/29558959/)
  21. Xu CH, Xiao LM, Liu Y, Chen LK, Zheng SY, Zeng EM, Li DH. The lncRNA HOXA11-AS promotes glioma cell growth and metastasis by targeting miR-130a-5p/HMGB2. *Eur Rev Med Pharmacol Sci*. 2019; 23:241–52. [https://doi.org/10.26355/eurrev\\_201901\\_16770](https://doi.org/10.26355/eurrev_201901_16770) PMID:[30657566](https://pubmed.ncbi.nlm.nih.gov/30657566/)
  22. Zhou H, Ma Y, Zhong D, Yang L. Knockdown of lncRNA HOXD-AS1 suppresses proliferation, migration and invasion and enhances cisplatin sensitivity of glioma cells by sponging miR-204. *Biomed Pharmacother*. 2019; 112:108633. <https://doi.org/10.1016/j.biopha.2019.108633> PMID:[30784927](https://pubmed.ncbi.nlm.nih.gov/30784927/)
  23. Chang KC, Diermeier SD, Yu AT, Brine LD, Russo S, Bhatia S, Alsudani H, Kostroff K, Bhuiya T, Brogi E, Pappin DJ, Bennett CF, Rigo F, Spector DL. MaTAR25 lncRNA regulates the Tensin1 gene to impact breast cancer progression. *Nat Commun*. 2020; 11:6438. <https://doi.org/10.1038/s41467-020-20207-y> PMID:[33353933](https://pubmed.ncbi.nlm.nih.gov/33353933/)
  24. Diermeier SD, Chang KC, Freier SM, Song J, El Demerdash O, Krasnitz A, Rigo F, Bennett CF, Spector DL. Mammary Tumor-Associated RNAs Impact Tumor Cell Proliferation, Invasion, and Migration. *Cell Rep*. 2016; 17:261–74. <https://doi.org/10.1016/j.celrep.2016.08.081> PMID:[27681436](https://pubmed.ncbi.nlm.nih.gov/27681436/)
  25. Zhu Z, Zhang H, Chen B, Liu X, Zhang S, Zong Z, Gao M. PD-L1-Mediated Immunosuppression in Glioblastoma Is Associated With the Infiltration and M2-Polarization of Tumor-Associated Macrophages. *Front Immunol*. 2020; 11:588552. <https://doi.org/10.3389/fimmu.2020.588552> PMID:[33329573](https://pubmed.ncbi.nlm.nih.gov/33329573/)
  26. Wang X, Guo G, Guan H, Yu Y, Lu J, Yu J. Challenges and potential of PD-1/PD-L1 checkpoint blockade immunotherapy for glioblastoma. *J Exp Clin Cancer Res*. 2019; 38:87. <https://doi.org/10.1186/s13046-019-1085-3> PMID:[30777100](https://pubmed.ncbi.nlm.nih.gov/30777100/)
  27. Nduom EK, Wei J, Yaghi NK, Huang N, Kong LY, Gabrusiewicz K, Ling X, Zhou S, Ivan C, Chen JQ, Burks

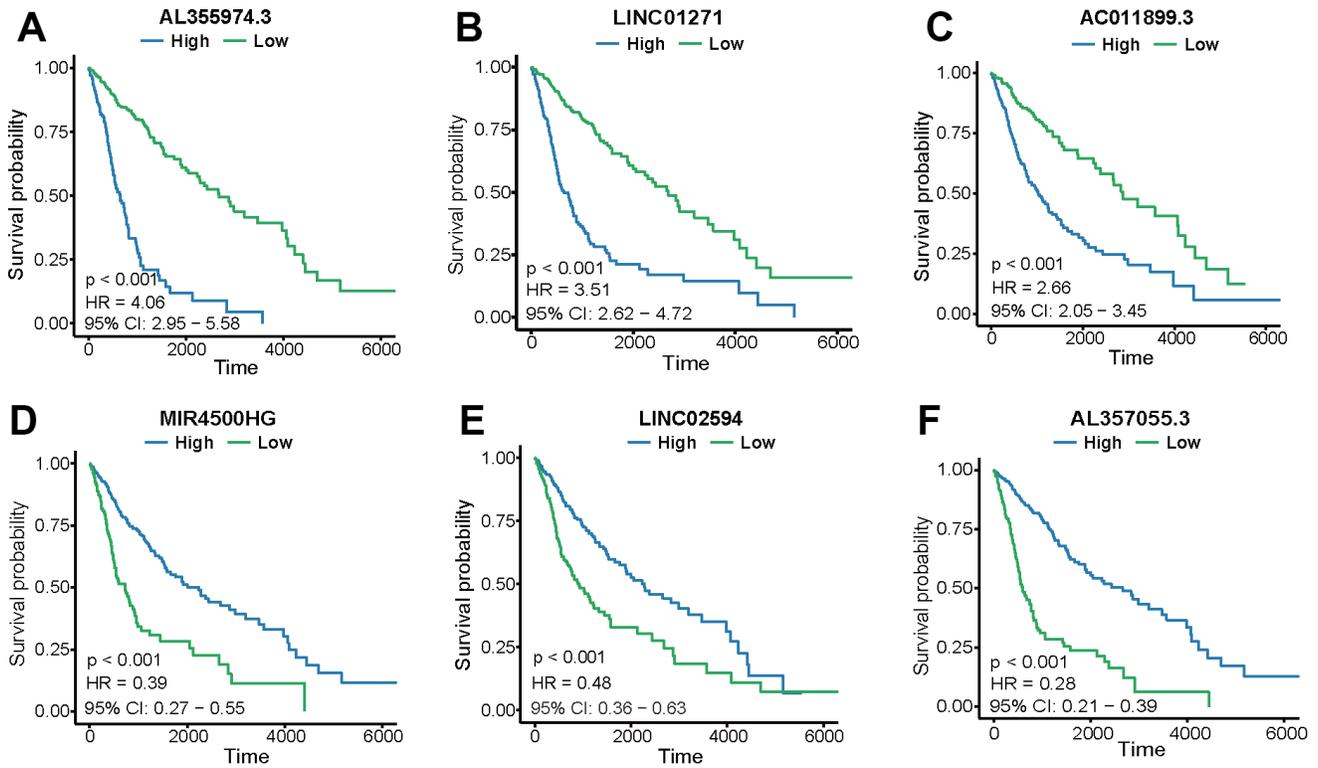
- JK, Fuller GN, Calin GA, et al. PD-L1 expression and prognostic impact in glioblastoma. *Neuro Oncol.* 2016; 18:195–205.  
<https://doi.org/10.1093/neuonc/nov172>  
PMID:[26323609](https://pubmed.ncbi.nlm.nih.gov/26323609/)
28. Berghoff AS, Kiesel B, Widhalm G, Rajky O, Ricken G, Wöhrer A, Dieckmann K, Filipits M, Brandstetter A, Weller M, Kurscheid S, Hegi ME, Zielinski CC, et al. Programmed death ligand 1 expression and tumor-infiltrating lymphocytes in glioblastoma. *Neuro Oncol.* 2015; 17:1064–75.  
<https://doi.org/10.1093/neuonc/nou307>  
PMID:[25355681](https://pubmed.ncbi.nlm.nih.gov/25355681/)
29. Qian J, Wang C, Wang B, Yang J, Wang Y, Luo F, Xu J, Zhao C, Liu R, Chu Y. The IFN- $\gamma$ /PD-L1 axis between T cells and tumor microenvironment: hints for glioma anti-PD-1/PD-L1 therapy. *J Neuroinflammation.* 2018; 15:290.  
<https://doi.org/10.1186/s12974-018-1330-2>  
PMID:[30333036](https://pubmed.ncbi.nlm.nih.gov/30333036/)
30. Himes BT, Peterson TE, de Mooij T, Garcia LMC, Jung MY, Uhm S, Yan D, Tyson J, Jin-Lee HJ, Parney D, Abukhadra Y, Gustafson MP, Dietz AB, et al. The role of extracellular vesicles and PD-L1 in glioblastoma-mediated immunosuppressive monocyte induction. *Neuro Oncol.* 2020; 22:967–78.  
<https://doi.org/10.1093/neuonc/noaa029>  
PMID:[32080744](https://pubmed.ncbi.nlm.nih.gov/32080744/)
31. Saha D, Martuza RL, Rabkin SD. Macrophage Polarization Contributes to Glioblastoma Eradication by Combination Immunovirotherapy and Immune Checkpoint Blockade. *Cancer Cell.* 2017; 32:253–67.e5.  
<https://doi.org/10.1016/j.ccell.2017.07.006>  
PMID:[28810147](https://pubmed.ncbi.nlm.nih.gov/28810147/)
32. Reardon DA, Gokhale PC, Klein SR, Ligon KL, Rodig SJ, Ramkissoon SH, Jones KL, Conway AS, Liao X, Zhou J, Wen PY, Van Den Abbeele AD, Hodi FS, et al. Glioblastoma Eradication Following Immune Checkpoint Blockade in an Orthotopic, Immunocompetent Model. *Cancer Immunol Res.* 2016; 4:124–35.  
<https://doi.org/10.1158/2326-6066.CIR-15-0151>  
PMID:[26546453](https://pubmed.ncbi.nlm.nih.gov/26546453/)
33. Cheng Q, Duan W, He S, Li C, Cao H, Liu K, Ye W, Yuan B, Xia Z. Multi-Omics Data Integration Analysis of an Immune-Related Gene Signature in LGG Patients With Epilepsy. *Front Cell Dev Biol.* 2021; 9:686909.  
<https://doi.org/10.3389/fcell.2021.686909>  
PMID:[34336837](https://pubmed.ncbi.nlm.nih.gov/34336837/)
34. Wang Q, Zhou D, Wu F, Liang Q, He Q, Peng M, Yao T, Hu Y, Qian B, Tang J, Wang X, Liu W, Yu F, Chen C. Immune Microenvironment Signatures as Biomarkers to Predict Early Recurrence of Stage Ia-b Lung Cancer. *Front Oncol.* 2021; 11:680287.  
<https://doi.org/10.3389/fonc.2021.680287>  
PMID:[34395248](https://pubmed.ncbi.nlm.nih.gov/34395248/)
35. Li Q, Barres BA. Microglia and macrophages in brain homeostasis and disease. *Nat Rev Immunol.* 2018; 18:225–42.  
<https://doi.org/10.1038/nri.2017.125>  
PMID:[29151590](https://pubmed.ncbi.nlm.nih.gov/29151590/)
36. Xue N, Zhou Q, Ji M, Jin J, Lai F, Chen J, Zhang M, Jia J, Yang H, Zhang J, Li W, Jiang J, Chen X. Chlorogenic acid inhibits glioblastoma growth through repolarizing macrophage from M2 to M1 phenotype. *Sci Rep.* 2017; 7:39011.  
<https://doi.org/10.1038/srep39011>  
PMID:[28045028](https://pubmed.ncbi.nlm.nih.gov/28045028/)
37. Wang Q, Hu B, Hu X, Kim H, Squatrito M, Scarpace L, deCarvalho AC, Lyu S, Li P, Li Y, Barthel F, Cho HJ, Lin YH, et al. Tumor Evolution of Glioma-Intrinsic Gene Expression Subtypes Associates with Immunological Changes in the Microenvironment. *Cancer Cell.* 2017; 32:42–56.e6.  
<https://doi.org/10.1016/j.ccell.2017.06.003>  
PMID:[28697342](https://pubmed.ncbi.nlm.nih.gov/28697342/)
38. Couto M, Coelho-Santos V, Santos L, Fontes-Ribeiro C, Silva AP, Gomes CM. The interplay between glioblastoma and microglia cells leads to endothelial cell monolayer dysfunction via the interleukin-6-induced JAK2/STAT3 pathway. *J Cell Physiol.* 2019; 234:19750–60.  
<https://doi.org/10.1002/jcp.28575>  
PMID:[30937892](https://pubmed.ncbi.nlm.nih.gov/30937892/)
39. Wang Q, He Z, Huang M, Liu T, Wang Y, Xu H, Duan H, Ma P, Zhang L, Zamvil SS, Hidalgo J, Zhang Z, O'Rourke DM, et al. Vascular niche IL-6 induces alternative macrophage activation in glioblastoma through HIF-2 $\alpha$ . *Nat Commun.* 2018; 9:559.  
<https://doi.org/10.1038/s41467-018-03050-0>  
PMID:[29422647](https://pubmed.ncbi.nlm.nih.gov/29422647/)
40. Zhang Y, Yu G, Chu H, Wang X, Xiong L, Cai G, Liu R, Gao H, Tao B, Li W, Li G, Liang J, Yang W. Macrophage-Associated PGK1 Phosphorylation Promotes Aerobic Glycolysis and Tumorigenesis. *Mol Cell.* 2018; 71:201–15.e7.  
<https://doi.org/10.1016/j.molcel.2018.06.023>  
PMID:[30029001](https://pubmed.ncbi.nlm.nih.gov/30029001/)
41. Wei J, Marisetty A, Schrand B, Gabrusiewicz K, Hashimoto Y, Ott M, Grami Z, Kong LY, Ling X, Caruso H, Zhou S, Wang YA, Fuller GN, et al. Osteopontin mediates glioblastoma-associated macrophage infiltration and is a potential therapeutic target. *J Clin Invest.* 2019; 129:137–49.  
<https://doi.org/10.1172/JCI121266> PMID:[30307407](https://pubmed.ncbi.nlm.nih.gov/30307407/)

42. Aslan K, Turco V, Blobner J, Sonner JK, Liuzzi AR, Núñez NG, De Feo D, Kickingeder P, Fischer M, Green E, Sadik A, Friedrich M, Sanghvi K, et al. Heterogeneity of response to immune checkpoint blockade in hypermutated experimental gliomas. *Nat Commun.* 2020; 11:931.  
<https://doi.org/10.1038/s41467-020-14642-0>  
PMID:[32071302](https://pubmed.ncbi.nlm.nih.gov/32071302/)
43. Han S, Zhang C, Li Q, Dong J, Liu Y, Huang Y, Jiang T, Wu A. Tumour-infiltrating CD4(+) and CD8(+) lymphocytes as predictors of clinical outcome in glioma. *Br J Cancer.* 2014; 110:2560–8.  
<https://doi.org/10.1038/bjc.2014.162> PMID:[24691423](https://pubmed.ncbi.nlm.nih.gov/24691423/)
44. Perng P, Lim M. Immunosuppressive Mechanisms of Malignant Gliomas: Parallels at Non-CNS Sites. *Front Oncol.* 2015; 5:153.  
<https://doi.org/10.3389/fonc.2015.00153>  
PMID:[26217588](https://pubmed.ncbi.nlm.nih.gov/26217588/)
45. Zhang J, Wang F, Liu F, Xu G. Predicting STAT1 as a prognostic marker in patients with solid cancer. *Ther Adv Med Oncol.* 2020; 12:1758835920917558.  
<https://doi.org/10.1177/1758835920917558>  
PMID:[32426049](https://pubmed.ncbi.nlm.nih.gov/32426049/)
46. Zhao L, Li X, Su J, Wang Gong F, Lu J, Wei Y. STAT1 determines aggressiveness of glioblastoma both *in vivo* and *in vitro* through wnt/ $\beta$ -catenin signalling pathway. *Cell Biochem Funct.* 2020; 38:630–41.  
<https://doi.org/10.1002/cbf.3518> PMID:[32390230](https://pubmed.ncbi.nlm.nih.gov/32390230/)
47. Yang W, Li Y, Gao R, Xiu Z, Sun T. MHC class I dysfunction of glioma stem cells escapes from CTL-mediated immune response via activation of Wnt/ $\beta$ -catenin signaling pathway. *Oncogene.* 2020; 39:1098–111.  
<https://doi.org/10.1038/s41388-019-1045-6>  
PMID:[31591480](https://pubmed.ncbi.nlm.nih.gov/31591480/)
48. Kohanbash G, Carrera DA, Shrivastav S, Ahn BJ, Jahan N, Mazor T, Chheda ZS, Downey KM, Watchmaker PB, Beppler C, Warta R, Amankulor NA, Herold-Mende C, et al. Isocitrate dehydrogenase mutations suppress STAT1 and CD8+ T cell accumulation in gliomas. *J Clin Invest.* 2017; 127:1425–37.  
<https://doi.org/10.1172/JCI90644> PMID:[28319047](https://pubmed.ncbi.nlm.nih.gov/28319047/)
49. Morrison BJ, Steel JC, Morris JC. Reduction of MHC-I expression limits T-lymphocyte-mediated killing of Cancer-initiating cells. *BMC Cancer.* 2018; 18:469.  
<https://doi.org/10.1186/s12885-018-4389-3>  
PMID:[29699516](https://pubmed.ncbi.nlm.nih.gov/29699516/)
50. Zhang D, Qiu B, Wang Y, Guan Y, Zhang L, Wu A. Temozolomide increases MHC-I expression via NF- $\kappa$ B signaling in glioma stem cells. *Cell Biol Int.* 2017; 41:680–90.  
<https://doi.org/10.1002/cbin.10773> PMID:[28403532](https://pubmed.ncbi.nlm.nih.gov/28403532/)
51. Gussyatiner O, Hegi ME. Glioma epigenetics: From subclassification to novel treatment options. *Semin Cancer Biol.* 2018; 51:50–8.  
<https://doi.org/10.1016/j.semcancer.2017.11.010>  
PMID:[29170066](https://pubmed.ncbi.nlm.nih.gov/29170066/)
52. Weller M, Stupp R, Hegi ME, van den Bent M, Tonn JC, Sanson M, Wick W, Reifenberger G. Personalized care in neuro-oncology coming of age: why we need MGMT and 1p/19q testing for malignant glioma patients in clinical practice. *Neuro Oncol.* 2012 (Suppl 4); 14:iv100–8.  
<https://doi.org/10.1093/neuonc/nos206>  
PMID:[23095825](https://pubmed.ncbi.nlm.nih.gov/23095825/)
53. Verhaak RG, Hoadley KA, Purdom E, Wang V, Qi Y, Wilkerson MD, Miller CR, Ding L, Golub T, Mesirov JP, Alexe G, Lawrence M, O’Kelly M, et al, and Cancer Genome Atlas Research Network. Integrated genomic analysis identifies clinically relevant subtypes of glioblastoma characterized by abnormalities in PDGFRA, IDH1, EGFR, and NF1. *Cancer Cell.* 2010; 17:98–110.  
<https://doi.org/10.1016/j.ccr.2009.12.020>  
PMID:[20129251](https://pubmed.ncbi.nlm.nih.gov/20129251/)
54. Daubon T, Hemadou A, Romero Garmendia I, Saleh M. Glioblastoma Immune Landscape and the Potential of New Immunotherapies. *Front Immunol.* 2020; 11:585616.  
<https://doi.org/10.3389/fimmu.2020.585616>  
PMID:[33154756](https://pubmed.ncbi.nlm.nih.gov/33154756/)
55. Khasraw M, Reardon DA, Weller M, Sampson JH. PD-1 Inhibitors: Do they have a Future in the Treatment of Glioblastoma? *Clin Cancer Res.* 2020; 26:5287–96.  
<https://doi.org/10.1158/1078-0432.CCR-20-1135>  
PMID:[32527943](https://pubmed.ncbi.nlm.nih.gov/32527943/)
56. Han S, Liu Y, Cai SJ, Qian M, Ding J, Larion M, Gilbert MR, Yang C. IDH mutation in glioma: molecular mechanisms and potential therapeutic targets. *Br J Cancer.* 2020; 122:1580–89.  
<https://doi.org/10.1038/s41416-020-0814-x>  
PMID:[32291392](https://pubmed.ncbi.nlm.nih.gov/32291392/)
57. Liang L, Xia W, Yao L, Wu Q, Hua L, Cheng G, Wang Z, Zhao R. Long non-coding RNA profile study identifies an immune-related lncRNA prognostic signature for prostate adenocarcinoma. *Int Immunopharmacol.* 2021; 101:108267.  
<https://doi.org/10.1016/j.intimp.2021.108267>  
PMID:[34740081](https://pubmed.ncbi.nlm.nih.gov/34740081/)
58. Liu J, Wang J, Song Y, Ma B, Luo J, Ni Z, Gao P, Sun J, Zhao J, Chen X, Wang Z. A panel consisting of three novel circulating lncRNAs, is it a predictive tool for gastric cancer? *J Cell Mol Med.* 2018; 22:3605–13.  
<https://doi.org/10.1111/jcmm.13640>  
PMID:[29700972](https://pubmed.ncbi.nlm.nih.gov/29700972/)

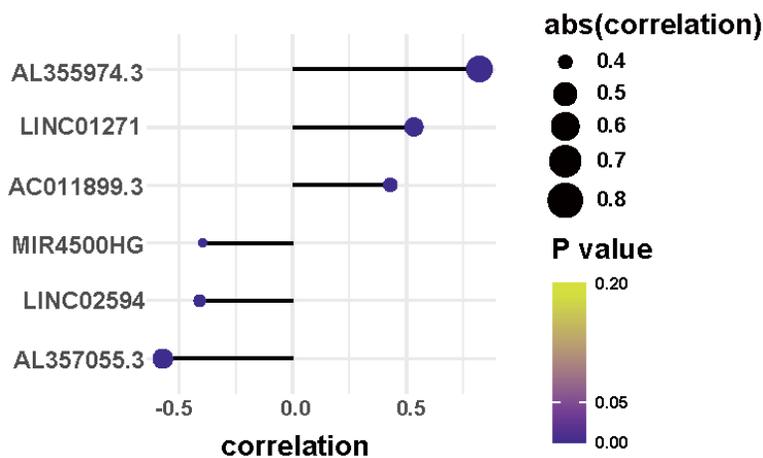
59. Silva A, Cerqueira MC, Rosa B, Sobral C, Pinto-Ribeiro F, Costa MF, Baltazar F, Afonso J. Prognostic Value of Monocarboxylate Transporter 1 Overexpression in Cancer: A Systematic Review. *Int J Mol Sci.* 2023; 24:5141. <https://doi.org/10.3390/ijms24065141> PMID:36982217
60. Chen L, Li Y, Deng X. Comprehensive analysis of pan-cancer reveals the potential of SLC16A1 as a prognostic and immunological biomarker. *Medicine (Baltimore).* 2023; 102:e33242. <https://doi.org/10.1097/MD.00000000000033242> PMID:36930112
61. Kent LN, Leone G. The broken cycle: E2F dysfunction in cancer. *Nat Rev Cancer.* 2019; 19:326–38. <https://doi.org/10.1038/s41568-019-0143-7> PMID:31053804
62. Johnson J, Thijssen B, McDermott U, Garnett M, Wessels LFA, Bernards R. Targeting the RB-E2F pathway in breast cancer. *Oncogene.* 2016; 35:4829–35. <https://doi.org/10.1038/onc.2016.32> PMID:26923330
63. Trikha P, Sharma N, Pena C, Reyes A, Pécot T, Khurshid S, Rawahneh M, Moffitt J, Stephens JA, Fernandez SA, Ostrowski MC, Leone G. E2f3 in tumor macrophages promotes lung metastasis. *Oncogene.* 2016; 35:3636–46. <https://doi.org/10.1038/onc.2015.429> PMID:26549026
64. Hu D, Meng N, Lou X, Li Z, Teng Y, Tu B, Zou Y, Wang F. Prognostic Values of E2F1/2 Transcriptional Expressions in Chromophobe Renal Cell Carcinoma Patients: Evidence from Bioinformatics Analysis. *Int J Gen Med.* 2021; 14:3593–609. <https://doi.org/10.2147/IJGM.S321585> PMID:34295182
65. Liao P, Han S, Qu H. Expression, Prognosis, and Immune Infiltrates Analyses of E2Fs in Human Brain and CNS Cancer. *Biomed Res Int.* 2020; 2020:6281635. <https://doi.org/10.1155/2020/6281635> PMID:33381564
66. Zhao Z, Zhang KN, Wang Q, Li G, Zeng F, Zhang Y, Wu F, Chai R, Wang Z, Zhang C, Zhang W, Bao Z, Jiang T. Chinese Glioma Genome Atlas (CGGA): A Comprehensive Resource with Functional Genomic Data from Chinese Glioma Patients. *Genomics Proteomics Bioinformatics.* 2021; 19:1–12. <https://doi.org/10.1016/j.gpb.2020.10.005> PMID:33662628
67. Newman AM, Liu CL, Green MR, Gentles AJ, Feng W, Xu Y, Hoang CD, Diehn M, Alizadeh AA. Robust enumeration of cell subsets from tissue expression profiles. *Nat Methods.* 2015; 12:453–7. <https://doi.org/10.1038/nmeth.3337> PMID:25822800
68. Hänzelmann S, Castelo R, Guinney J. GSVA: gene set variation analysis for microarray and RNA-seq data. *BMC Bioinformatics.* 2013; 14:7. <https://doi.org/10.1186/1471-2105-14-7> PMID:23323831
69. Wilkerson MD, Hayes DN. ConsensusClusterPlus: a class discovery tool with confidence assessments and item tracking. *Bioinformatics.* 2010; 26:1572–3. <https://doi.org/10.1093/bioinformatics/btq170> PMID:20427518
70. Liberzon A, Birger C, Thorvaldsdóttir H, Ghandi M, Mesirov JP, Tamayo P. The Molecular Signatures Database (MSigDB) hallmark gene set collection. *Cell Syst.* 2015; 1:417–25. <https://doi.org/10.1016/j.cels.2015.12.004> PMID:26771021
71. Yu G, Wang LG, Han Y, He QY. clusterProfiler: an R package for comparing biological themes among gene clusters. *OMICS.* 2012; 16:284–7. <https://doi.org/10.1089/omi.2011.0118> PMID:22455463
72. Jiang P, Gu S, Pan D, Fu J, Sahu A, Hu X, Li Z, Traugh N, Bu X, Li B, Liu J, Freeman GJ, Brown MA, Wucherpennig KW, Liu XS. Signatures of T cell dysfunction and exclusion predict cancer immunotherapy response. *Nat Med.* 2018; 24:1550–8. <https://doi.org/10.1038/s41591-018-0136-1> PMID:30127393
73. Geeleher P, Cox N, Huang RS. pRRophetic: an R package for prediction of clinical chemotherapeutic response from tumor gene expression levels. *PLoS One.* 2014; 9:e107468. <https://doi.org/10.1371/journal.pone.0107468> PMID:25229481
74. Mayer G, Müller J, Lünse CE. RNA diagnostics: real-time RT-PCR strategies and promising novel target RNAs. *Wiley Interdiscip Rev RNA.* 2011; 2:32–41. <https://doi.org/10.1002/wrna.46> PMID:21956968
75. Feng J, Tu R, Liu F. PD-L1-related LncRNAs are Associated with Immune Microenvironment and Prognosis in Glioma. *Research Square [Preprint].* 2022. <https://doi.org/10.21203/rs.3.rs-2327559/v1>

SUPPLEMENTARY MATERIALS

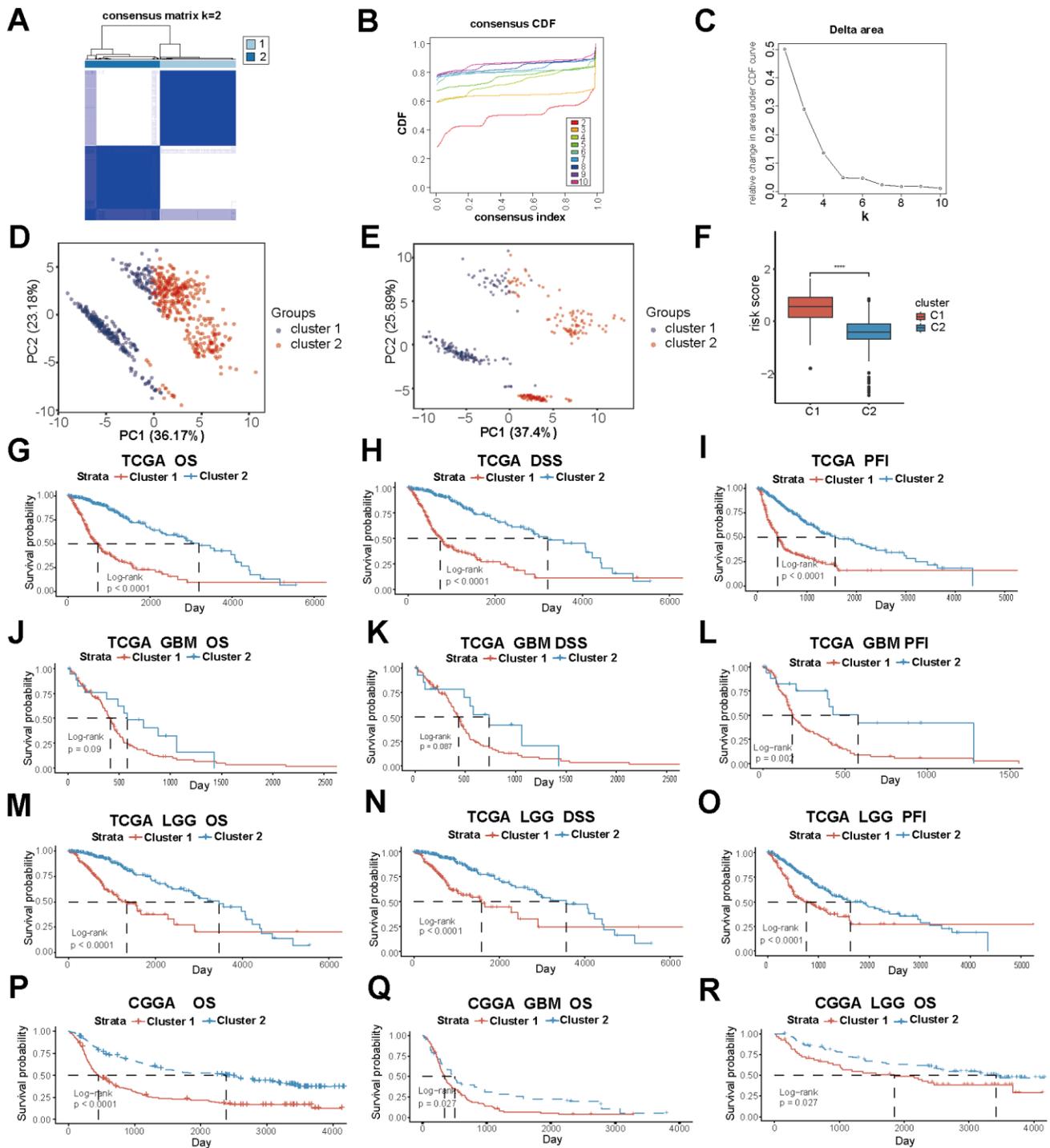
Supplementary Figures



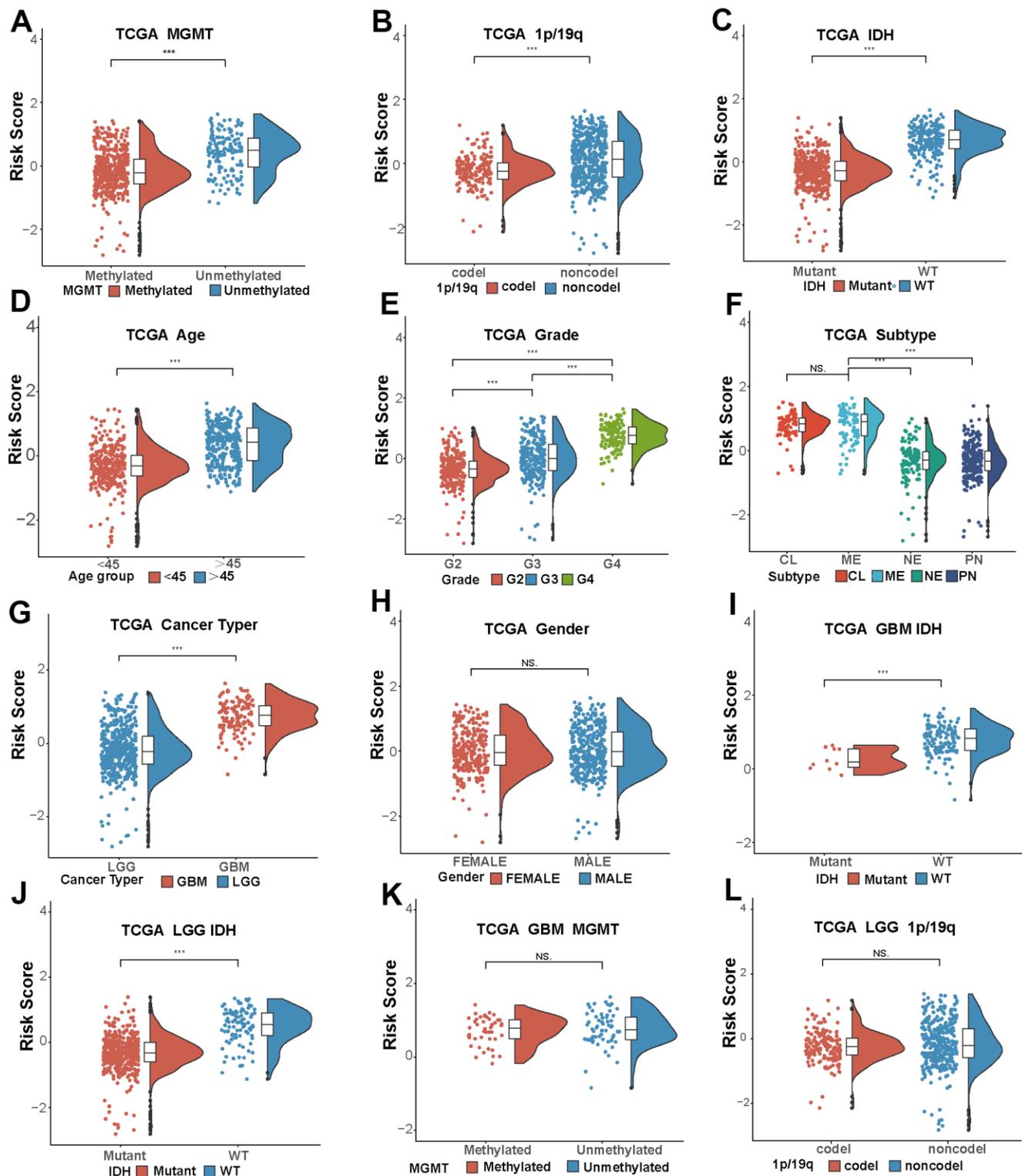
**Supplementary Figure 1. The expression of PD-L1-related lncRNAs is associated with glioma patients' prognosis. (A–F)** Survival analysis curves of AL355974.3 (A), LINC01271 (B), AC011899.3 (C), MIR4500HG (D), LINC02594 (E), and L357055.3 (F) with high and low expression level.



**Supplementary Figure 2. Correlation between 6 PD-L1-related lncRNAs and risk score.**

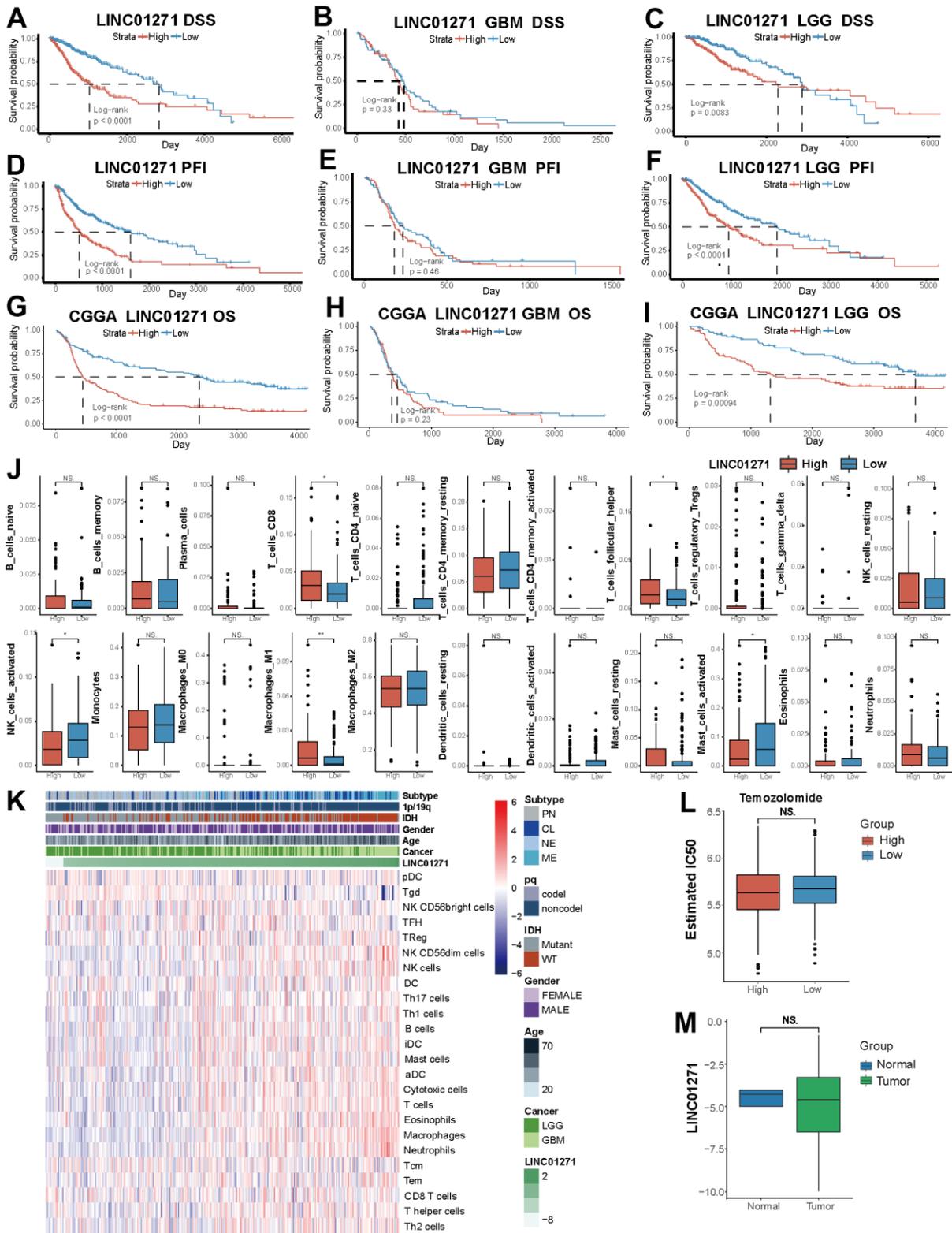


**Supplementary Figure 3. Consensus cluster analysis reflects glioma patients' prognosis.** (A–C) Consensus cluster analysis divided samples into two groups. (D, E) Principal component analysis. (F) Risk scores of two clusters. (G–I) OS, DSS, PFI in two clusters of all glioma patients in TCGA dataset. (J–O) OS, DSS, PFI of GBM and LGG subtype patients of two clusters in TCGA dataset. (P–R) OS of all patients, GBM and LGG patients in CGGA dataset of two clusters.



**Supplementary Figure 4. Quantitative analysis of clinical and glioma pathological characteristics in different risk score groups in TCGA dataset.** (A–H) Comparison of clinical and pathological features in all glioma patients. (I, J) IDH type of different risk score groups among GBM and LGG patients. (K) MGMT methylation status of different risk score groups among GBM patients. (L) 1p/19q co-deletion status of different risk score groups among LGG patients.





**Supplementary Figure 6. Correlations between LINC01271 expression and the prognosis, immune and inflammatory features of glioma.** (A–F) DSS and PFI among all glioma patients, GBM and LGG patients with different LINC01271 expression in TCGA dataset. (G–I) OS of all glioma patients, GBM and LGG patients with different LINC01271 expression in CGGA dataset. (J) Quantitative analysis of the expression of immune cells in different LINC01271 expression groups. (K) Heatmap displaying the characteristics of immune cells panel in CGGA dataset. (L) Sensitivity to TMZ treatment of patients with different LINC01271 expression. (M) LINC01271 expression in tumor and normal tissues in TCGA dataset.

## Supplementary Tables

**Supplementary Table 1. Multivariate cox regression of PD-L1-related lncRNAs.**

Characteristics	Hazard.Ratio	CI95	P.value
A1BG-AS1	1.1	0.95-1.28	0.198
AC009041.3	0.98	0.91-1.05	0.52
AC009955.1	1.03	0.96-1.12	0.393
AC011899.3	1.07	1-1.15	<b>0.038</b>
AC093726.2	1.06	0.9-1.24	0.492
AC131157.1	0.96	0.89-1.03	0.218
AC148476.1	1	0.95-1.07	0.897
ADAMTSL4-AS1	0.93	0.84-1.04	0.199
AL157702.2	1	0.95-1.05	0.97
AL355974.3	1.19	1.08-1.31	<b>0.001</b>
AL357055.3	0.78	0.65-0.93	<b>0.007</b>
AL442067.1	0.99	0.95-1.02	0.416
EPB41L4A-DT	1.12	0.94-1.35	0.212
LACTB2-AS1	1.09	0.99-1.19	0.071
LINC00887	1.06	1-1.12	0.051
LINC01271	1.07	1-1.15	<b>0.046</b>
LINC01504	1.06	0.97-1.15	0.208
LINC02594	0.95	0.91-0.99	<b>0.026</b>
LINC02612	0.99	0.94-1.04	0.684
MIR4500HG	0.93	0.88-0.98	<b>0.009</b>
SCAANT1	1.02	0.99-1.06	0.171
SNHG7	0.95	0.75-1.2	0.666
VLDLR-AS1	0.96	0.86-1.08	0.487

**Supplementary Table 2. Primer sequences for LINC01271.**

Name	Primer sequence (5'-3')
LINC01271-F	CGTCTCTGCACATTGTATGACC
LINC01271-R	CCTGGGATCTAGGGAGCTGAC

**Supplementary Table 3. Synthesis information for siRNAs.**

siRNA	Sense (5'-3')	Antisense (5'-3')
si-LINC01271-334	AGGAAAGACUGUAAAGAAUTT	AUUCUUUACAGUCUUUCCUTT
si-LINC01271-1196	GGACCAAUCACUAGAGCAATT	UUGCUCUAGUGAUUGGUCCTT
si-LINC01271-1564	GGACGGAAGUGGAGCAUCATT	UGAUGCUCACUCCGUCCTT