

A multi-omics approach based on ¹H-NMR metabonomics combined with target protein analysis to reveal the mechanism of RIAISs on cervical carcinoma patients

Chai Yanlan^{1,*}, Aibibai Jielili^{1,*}, Juan Wang², Haiying Tang³, Zi Liu^{2,*,&}, Ping Wang^{1,*}

¹Tianjin Medical University Cancer Institute and Cancer Hospital, National Clinical Research Center for Cancer, Key Laboratory of Cancer Prevention and Therapy, Tianjin's Clinical Research Center for Cancer, Tianjin 300060, P.R. China

²Department of Radiotherapy Oncology, The First Affiliated Hospital of Xi'an Jiao Tong University, Xi'an 710061, Shaanxi, P.R. China

³The Gao Qing People's Hospital of Zibo, Zibo 255000, Shandong, P.R. China

*Equal contribution

Correspondence to: Zi Liu, Ping Wang; email: liuzmail@163.com, <https://orcid.org/0000-0002-4689-261X>; wangping@tjmuch.com

Keywords: cervical carcinoma, RIAISs, ¹H-NMR metabonomic, multi-omics analysis

Abbreviations: CC: Cervical carcinoma; RIAISs: Radiation-induced acute intestinal symptoms; RIID: Radiation-induced intestinal damage; NMR: Nuclear magnetic resonance; FDR: False discovery rate

Received: June 9, 2022

Accepted: September 1, 2022

Published: September 27, 2022

Copyright: © 2023 Yanlan et al. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/3.0/) (CC BY 3.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

ABSTRACT

Cervical carcinoma (CC) is the fourth most common cancer in females and radiotherapy is always as the definitive therapy for cervical cancer patients who are not suitable for surgery. Radiation-induced acute intestinal symptoms (RIAISs) occur in 50-80% of cervical cancer patients. Some research shows that RIAISs may relate to inflammatory reaction by radiotherapy but the action mechanism is also not clearly and the details of the molecular mechanism are still urgently needed. In this paper, basing on ¹H-NMR metabonomic and bioinformatics analysis, an integrated multi-omics analysis including metabonomics and bioinformatics was performed. We propose a hypothesis about pathogenic mechanism on RIAISs and proofed it through western-blot. Our results indicated significant dysregulation of metabolic pathways in RIAIS patients. Most importantly, we found that RIAISs were associated p53 and PI3K-AKT pathway.

INTRODUCTION

During the therapeutic process of patients with cervical cancer and patients who are adverse surgical candidates, radiotherapy is always the ultimate treatment [1]. With advances in radiotherapy and related treatments, significant progress has been made in high-accuracy radial delivery and radiation therapy dose distribution. However, complication and the influence to normal tissue remains an issue [2].

The intestine is particularly sensitive to ionizing radiation and causes side effects such as vomiting, anorexia, weight loss, diarrhea, dehydration, and

infection [3]. Patients receiving radiation to the cervix, pelvis, abdomen, and colorectal system often experience short-term enteritis, but most experience chronic bowel problems [4]. When exposed to ionizing radiation, it causes mucosal inflammation and other side effects in pelvic radiotherapy. Radiation induced intestinal damage (RIID) will affect the treatment schedule and decrease patients' quality of life [5].

Cervical carcinoma (CC) is the fourth most common cancer in females, especially in developing countries [6]. However, radiotherapy can almost cause radiation-induced acute intestinal symptoms (RIAISs) in patients with cervical cancer. Some researches

show that RIAISs may related to inflammatory reaction by radiotherapy but the action mechanism is also not clearly and the details of the molecular mechanism are still urgently needed [7]. Currently, the treatment of RIIAS is mainly focused on supportive and complementary treatment, and the current treatment goal is still only to reduce the severity of symptoms and exacerbations [8].

Metabolomics is becoming an important component of systems biology [9]. Currently, metabolomics has been successfully applied to disease diagnosis, biomarker screening, and biological pathway characterization [10]. Except the mass spectrometry, the technological advances in nuclear magnetic resonance (NMR) spectroscopy have further improved the sensitivity and spectral resolution of metabolomics studies. NMR has irreplaceable advantages such as allowing non-destructive methods in untargeted metabolic analysis with little or no sample preparation [11, 12]. Now NMR spectroscopy is a powerful tool that is widely applied in metabolite identification and quantification [13].

In this study, feces, serum, and urine of CC patients were used as samples. An ^1H NMR based metabolomic analyses was employed to detect the change of metabolins before and after radiotherapy between different groups. The data integrated with bioinformatics analysis results were used to identify potential targets or pathways and novel therapeutic regimens. As a result, we built a profile of RIAISs which based on metabolins and bioinformatics. We found that radiotherapy mainly affected the PI3K-AKT and P53 pathway. The activation of these pathways will lead to an inflammatory reaction and these results could reveal the nosogenesis of RIAISs. The result will aid in our understanding of the pathobiology of the disease.

MATERIALS AND METHODS

Reagents and materials

Chemiluminescent HRP substrates were purchased from Millipore Corporation (MA, USA). Primary antibodies against AKT (33748), p-AKT (11054), NF κ B (48498), p-NF κ B (11166), GAPDH (40493), goat anti-rabbit IgG secondary antibody (L3012) were purchased from Signalway Antibody (MA, USA).

Patients' treatment, sample collection and storage

Detail information are as the same as our previous study [14]. A summary of the patients' characteristics is displayed in Table 1.

On treatment of patients, our previously published treatment protocol was followed for this study [14]. The details of patients' treatment, sample collection and storage are shown in the supporting information.

Samples were grouped as follows:

- Group I-samples of RIAIS group before radiotherapy.
- Group II-samples of the control group before radiotherapy.
- Group III-samples of RIAIS group after radiotherapy.
- Group IV-samples of the control group after radiotherapy.

Specimen preparation for ^1H NMR analysis

This section was according to our previous work [14]. The details of specimen preparation for ^1H NMR analysis are shown in the supporting information. The workflow of the whole study is shown in Figure 1.

^1H NMR analysis

This section was according to our previous work [14]. Spectra primarily comprising signals from metabolites and small molecules to be obtained [15]. The details of ^1H NMR analysis are shown in the supporting information.

Multivariate statistics

This section was according to our previous work [14]. The details of multivariate statistics are shown in the supporting information.

Metabolomic identification and bioinformatics analyses

To identify the differential metabolites and pathway ascription, a corresponding metabolomic database (<http://hmdb.ca>) and Compound Discoverer 3.0 software was used as the search database. Next, we used the DAVID tool (<https://david.ncifcrf.gov>) to analyze protein function and pathway enrichment. To identify the key targets, we used Venny 2.1.0 to intersect two parts, including all proteins from differential pathways by metabolomic analysis, proteins associated with RIAISs by GeneCards database. Then, the key target was uploaded to the STRING database (<https://www.string-db.org/cgi/input.pl>), and the minimum required interaction score was set to 0.4 to obtain a protein-protein interaction (PPI) network of key targets and proteins related to the key targets. Finally, the constructed networks were merged, and the duplicated edges were removed and visualized in Cytoscape3.2.1.

Table 1. Clinical and demographic information on all 66 cervical cancer patients, 11 of whom developed RIAISs.

Characteristics	Participants	RIAISs	Controls
No.of women	66	11	11
Age (years)			
Median/mean	53/52.1	54/51.9	55/54.2
Range	30-70	31-67	37-65
FIGO stage			
I	10	2	2
II	37	6	7
III	13	2	2
IV	6	1	0
Histological type			
Squamous carcinoma	64	11	11
Adenocarcinoma	2	0	0
Therapy			
Radical hysterectomy and adjuvant radiotherapy	34	7	7
Radical radiotherapy	32	4	4

Western blot

Total pulmonary protein was extracted from 50 mg of frozen urine sample. Then the total proteins either from cells or tissues by RIPA lysis buffer were applied for the western blot analysis, which processed in accordance with our previous methods [16].

Statistical analysis

The results are shown as the mean \pm standard deviation (SD). For single comparisons, significant differences between the means were determined by one way ANOVA test. A p-value less than 0.05 was considered to indicate significant differences. All data were processed using GraphPad Prism 5.01 software.

RESULTS

¹H NMR spectrum and PCA analysis

Import ¹H NMR results into MestReNova software for phase correction and baseline adjust. Figure 2 shows the signals from different samples and groups. To establish a global overview for discrimination between different groups, we did a PCA analysis of all NMR spectral data between different groups (Figure 3), all groups have separation trends expect the urine sample.

Metabonomic analyses revealed the key pathways of RIAISs caused by radiotherapy

To explore the underlying molecular mechanisms, an ¹H NMR-based metabolomic strategy was first used to

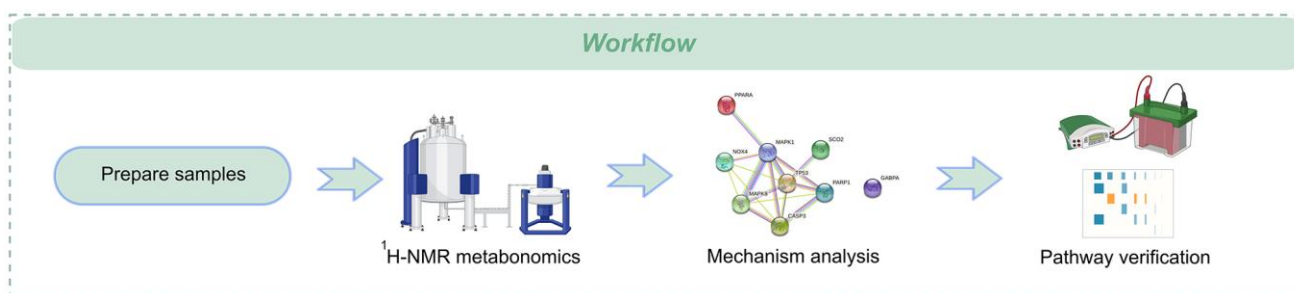


Figure 1. The workflow of the whole study.

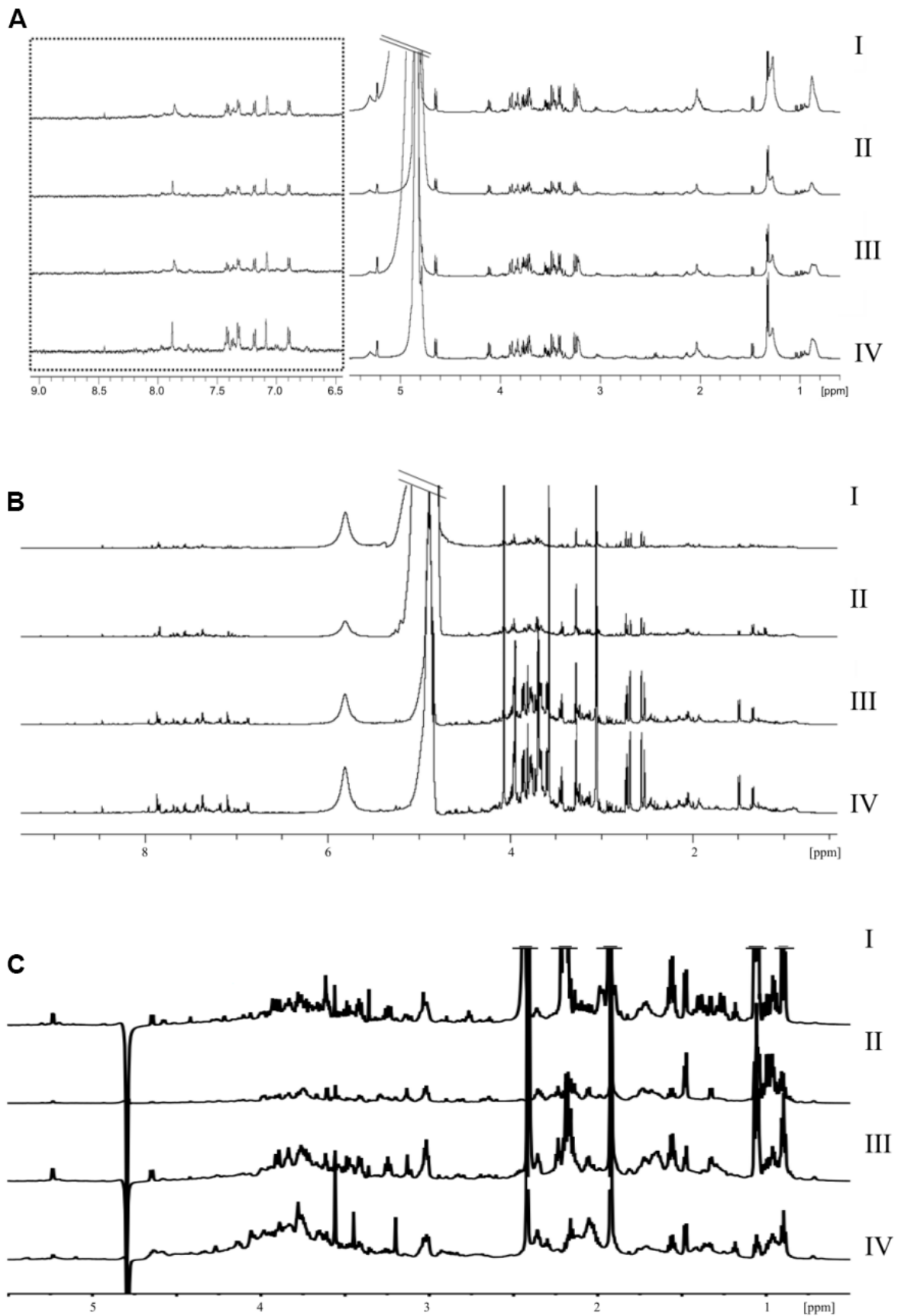


Figure 2. NMR chromatograms different samples and groups. (A) plasma (B) urine (C) feces.

identify differential metabolites in CC patients after radiotherapy. As a result, 733 metabolites were identified with a false discovery rate (FDR) ≤ 0.01 . After further data analysis by HMDB database, a total of 524 metabolites were found to be significantly different among different groups. With the criteria of p-value < 0.05 , only 151 metabolites were found. These proteins were not annotated in 20 different pathways in the KEGG database. Among these findings, the PI3K-Akt, MAPK, Thermogenesis, Choline metabolism in cancer, IL-7 metabolic pathways, p53, and other pathways were involved the main biological processes of RIAISs (Figure 4A, 4B).

To clarify the key action nodes of RIAISs caused by radiotherapy, all associated proteins involved in above differential pathways, which were identified by metabolomics (362) and RIAISs disease-associated proteins (1316) obtained from GeneCards, were used in

the integrated analysis. As shown in the Venn diagram in Figure 5A, only 9 intersection node proteins were identified: NFKB1, RAF1, TP53, AKT1, SCO2, MDM2, TLR3, PIK3CA, MAPK3 and Figure 5B shows the association of these proteins.

Network analysis of RIAISs caused by radiotherapy

To further predict the molecular mechanism of RIAISs caused by radiotherapy, a PPI network was built. As Figure 3A shows, there were a total of 58 proteins that interacted with the key proteins above and these proteins were regulated by the 9 focused proteins in the protein-protein interaction network. The 9 focused proteins could be divided into 19 signaling pathways like p53 signaling pathway, PI3K-Akt signaling pathway, Apoptosis - multiple species, Colorectal cancer and so on. Table 2 shows the fit score of rank of different pathways which exported by DAVID database.

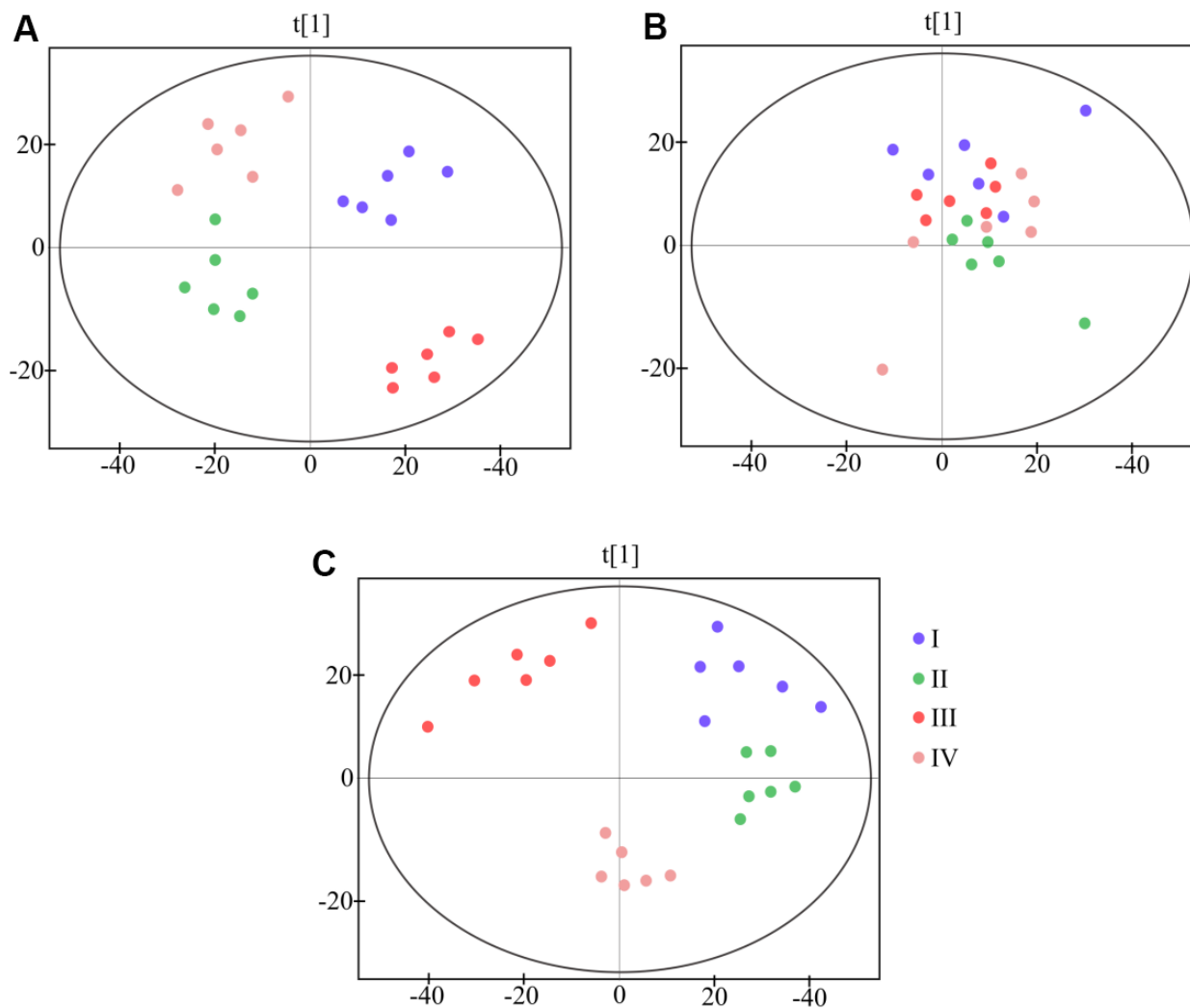


Figure 3. PCA scores plot based on ^1H NMR spectra of different sample from four groups. (A) plasma (B) urine (C) feces.

Significantly, the PI3K-Akt signaling pathway and p53 signaling pathway ranked the first and second. It prompts us that the possible molecular mechanism of RIAISs caused by radiotherapy.

According the results about the network analysis above, we choose the downstream proteins in PI3K-Akt

signaling pathway and p53 signaling pathway and tried to test and verify the phosphorylation levels of AKT and NFKB. The changes in the phosphorylation levels of AKT and NFKB in the urine samples are shown in Figures 6, 7. Western blot analysis revealed that, after radiotherapy the phosphorylation levels of AKT and NFKB were markedly increased (Tables 3, 4), it

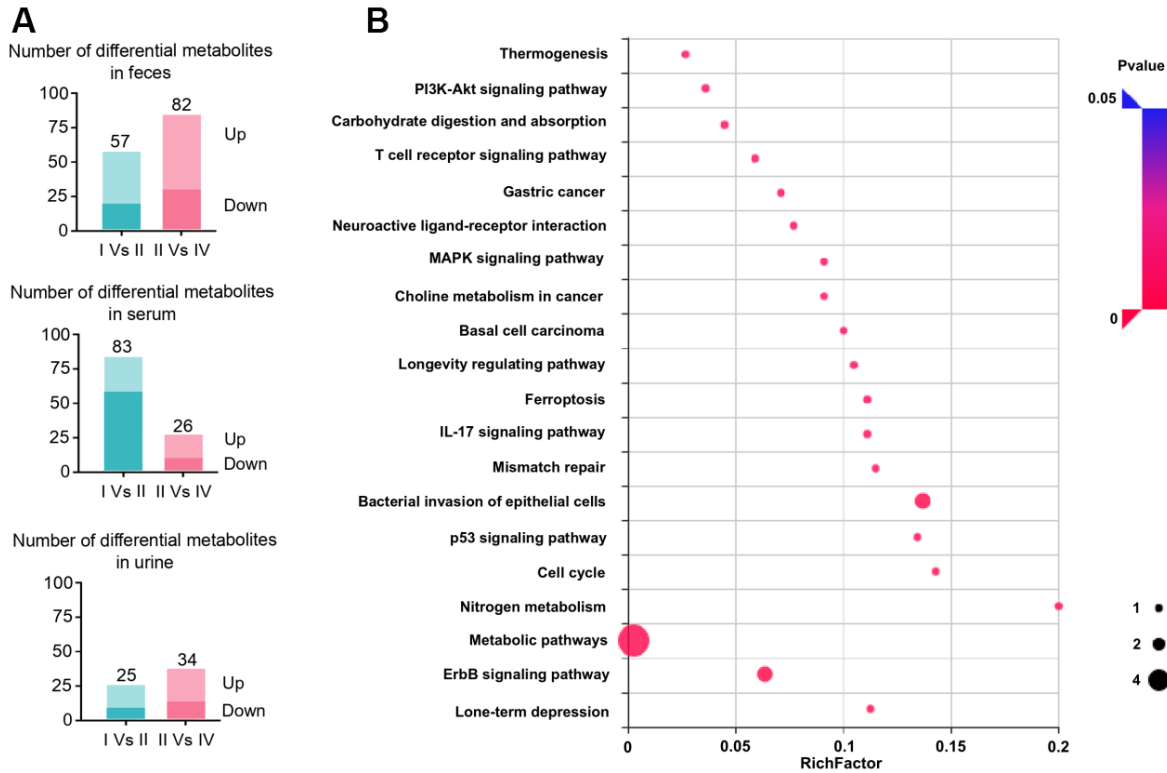


Figure 4. The integral analysis of quantitation proteomics, metabonomics and COPD associated proteins. (A) ¹H NMR-based metabolomic analysis of feces, serum, and urine sample. Quantitation of metabolites with the condition of Fold-change ≥ 1.2 or ≤ 0.83 and p-value ≤ 0.05 . (B) KEGG pathway enrichment analysis of differential metabolites.

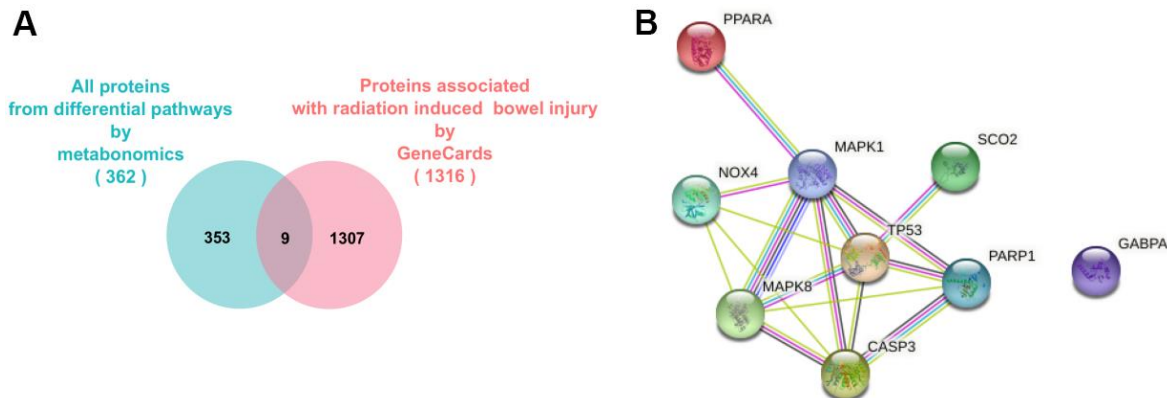


Figure 5. Network pharmacological analysis of RIAISs caused by radiotherapy. (A) Strategy of find out key target. (B) Connection network between the key nodes calculated by STRING.

Table 2. KEGG pathway enrichment analysis of key nodes.

No.	Pathway	Strength
1	p53 signaling pathway	2.15
2	PI3K-Akt signaling pathway	2.13
3	Colorectal cancer	2.06
4	Apoptosis - multiple species	1.91
5	Central carbon metabolism in cancer	1.87
6	IL-17 signaling pathway	1.82
7	MicroRNAs in cancer	1.82
8	Ras signaling pathway	1.81
9	cAMP signaling pathway	1.8
10	Th1 and Th2 cell differentiation	1.8
11	Choline metabolism in cancer	1.78
12	Sphingolipid signaling pathway	1.75
13	GnRH signaling pathway	1.72
14	Toll-like receptor signaling pathway	1.69
15	Natural killer cell mediated cytotoxicity	1.67
16	Wnt signaling pathway	1.65
17	Cellular senescence	1.61
18	Necroptosis	1.58
19	Pathways in cancer	1.58

indicated that radiotherapy activate the PI3K-Akt and p53 signaling pathway.

DISCUSSION

Radiotherapy is commonly used as the ultimate treatment for patients with locally advanced cervical cancer or poor surgical candidates [17]. For cervical cancer patients, the intestine is a major dose-limiting organ during pelvic radiotherapy because the intestine is sensitive to ionizing radiation [18]. RIAISs occurs in 50–75% of cervical cancer patients after radiotherapy, it will lead dose reductions, treatment interruptions, increased medical costs, and reduced quality of life, with symptoms including abdominal pain, diarrhea, and tenesmus. So, it is eagerly to find the molecular markers of RIAISs and supplement the details of molecular mechanism of RIAISs and it will help us improve the planning and monitoring of radiotherapy [19].

¹H NMR spectroscopy is a powerful tool for chemical structural elucidation and chemical dynamic studies and analytic instrument to get information of biomarkers and help us reveal the mechanism of disease more accurately [20]. Furthermore, it will help us understand different stratification of patients and identify specific drugs that respond better. It also provides a bridge between preclinical disease models and human patient populations, where common biomarkers and patient

populations between model systems provide important mechanistic connections and highlight potential therapeutic goals. This is the basis of precision drug therapy [21].

However, although the accuracy of instruments is developing rapidly, but single-omic results still have some limits, which mainly focus on the one-sidedness description of the entire life system. Bioinformatics now offered us more and more useful tools and databases, they could be a supplement of single-omic results and they will help us describe the vital process in a integrity view. In this study, a reasonable analysis strategy which based on metabolins and bioinformatics was used [22]. We found that radiotherapy mainly affected the PI3K-AKT and P53 pathway.

P53 is a tumor suppressor gene and is one of the primary cellular factors determining the nature of growth arrest and/or cell death after exposure to ionizing radiation. P53 pathway plays an important role in damage surveillance and as such has been dubbed the guardian of the genome. Upon DNA damage, P53 pathway was activate, downstream proteins (like NFkB) accumulation was increased and translocated to the nucleus where it binds to DNA, acting to regulate the transcription of a number of genes [23]. According to our results, after radiotherapy, the phosphorylation levels of NFkB

were markedly increased, it indicated that radiotherapy activate the p53 signaling pathway.

PI3K-AKT signaling plays a significant role in oxidative stress and inflammation [24]. In addition, it

also plays a key role in apoptosis and proliferation of tumor metastasis and invasion by regulating several pathways associated with cell growth, apoptosis, or invasion [25]. Activated AKT translocates to the cytoplasm and nucleus and activates downstream targets

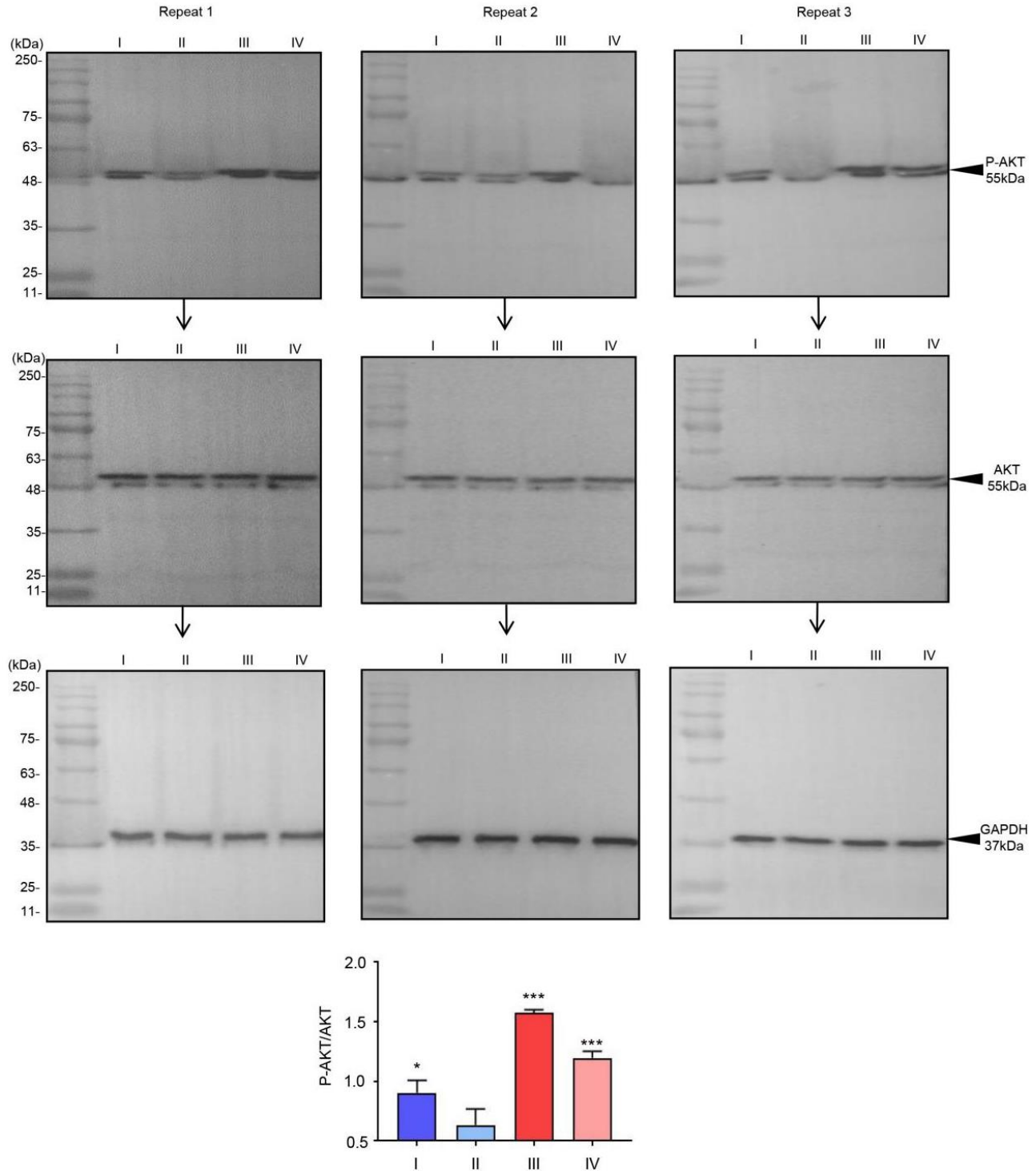


Figure 6. The phosphorylation of AKT in urine sample. Phosphorylation of AKT in urine sample regulated by radiotherapy. The relative intensity of P-AKT was compared with AKT. * $p < 0.05$, and *** $p < 0.001$ vs. II, (n = 3).

involved in survival, proliferation, cell cycle progression, growth, migration, and angiogenesis [26]. According to our results, after radiotherapy, the phosphorylation levels of AKT was markedly increased, it indicated that radiotherapy activate the PI3K-AKT signaling pathway.

In summary, this study demonstrated distinct differences in the spectra acquired between the feces of RIAIS patients before and after radiotherapy, and between RIAISs and controls. Basing on a ¹H-NMR metabonomic and bioinformatics analysis, we proposed a hypothesis about pathogenic mechanism on RIAISs

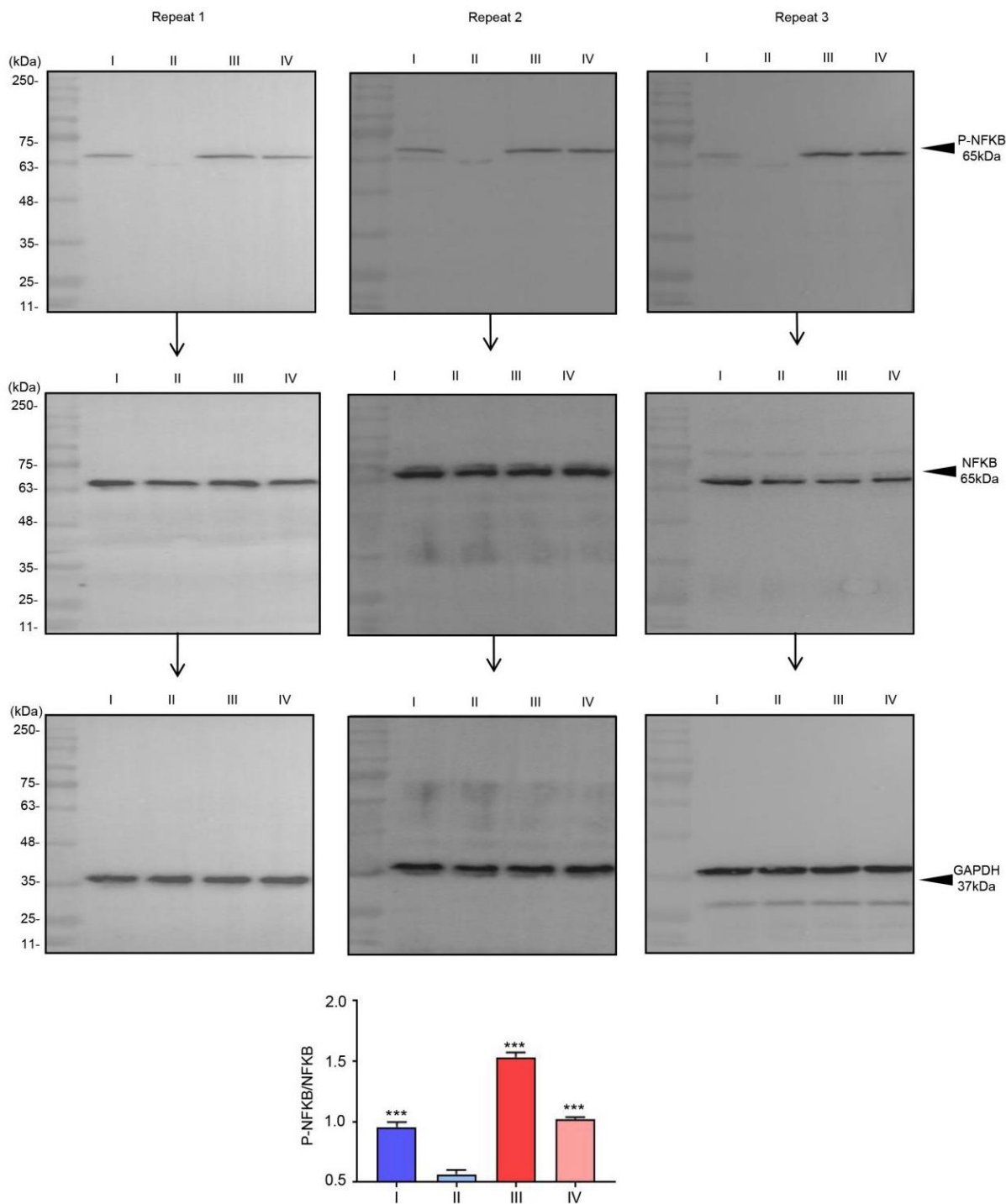


Figure 7. The phosphorylation of NFKB in urine sample. Phosphorylation of NFKB in urine sample regulated by radiotherapy. The relative intensity of P-NFKB was compared with NFKB. *** $p < 0.001$ vs. II, (n = 3).

Table 3. Raw data of western blotting with P-AKT in clinical tissue samples.

Group		I	II	III	IV
P-AKT	repeat1	1846	1362	2899	2314
	repeat2	1460	899	2825	2126
	repeat3	1669	1273	2963	2177
AKT	repeat1	3065	3076	3044	3048
	repeat2	3059	3100	3028	3077
	repeat3	3008	3088	3074	3060
GAPDH	repeat1	4290	4098	4001	4123
	repeat2	4137	4199	4241	4211
	repeat3	4087	4171	4164	4425
P-AKT/AKT	repeat1	1.00	0.74	1.58	1.26
	repeat2	0.79	0.48	1.55	1.15
	repeat3	0.92	0.68	1.60	1.18

Table 4. Raw data of western blotting with P-NFKB in clinical tissue samples.

Group		I	II	III	IV
P-NFKB	repeat1	1996	1063	2962	2027
	repeat2	1887	954	3088	2074
	repeat3	1811	960	3082	2011
NFKB	repeat1	3065	3076	3044	3048
	repeat2	3059	3100	3028	3077
	repeat3	3008	3088	3074	3060
GAPDH	repeat1	3969	4111	3949	4077
	repeat2	4109	4291	3902	4139
	repeat3	3948	4081	4012	3912
P-NFKB/NFKB	repeat1	1.00	0.53	1.49	1.02
	repeat2	0.95	0.4	1.57	1.04
	repeat3	0.92	0.48	1.54	1.01

and proofed it through western blot. Our results indicate significant dysregulation of metabolic pathways in RIAIS patients. Most importantly, we found that RIAISs were associated p53 and PI3K-AKT pathway.

AUTHOR CONTRIBUTIONS

Yanlan Chai collected specimens and drafted the manuscript. Aibibai Jielili performed ¹H-NMR assays and performed bioinformatics analysis. Juan Wang conducted western blot and signaling pathway analysis. Haiying Tang arranged references and modified the manuscript further. Zi Liu conceived of the study and participated in its design and coordination. Ping Wang provided funding and carried out final proofreading of the manuscript. All authors read and approved the final manuscript.

CONFLICTS OF INTEREST

The authors declare that they have no conflicts of interest.

ETHICAL STATEMENT AND CONSENT

This study was approved by the Ethics Committee of the First Affiliated Hospital of Xi'an Jiaotong University, Shaanxi, China. Informed consent was obtained from each participant before sample collection.

FUNDING

This study was financially supported by National Natural Science Foundation of China (Grant No. 81071838) and Chinese National Key Research and Development Project (Grant No.2018YFC1315601).

Editorial note

*This corresponding author has a verified history of publications using a personal email address for correspondence.

REFERENCES

1. Xiang M, Kidd EA. Benefit of Cisplatin With Definitive Radiotherapy in Older Women With Cervical Cancer. *J Natl Compr Canc Netw*. 2019; 17:969–75. <https://doi.org/10.6004/jccn.2019.7289> PMID:31390586
2. Gao L, Zheng H, Cai Q, Wei L. Autophagy and Tumour Radiotherapy. *Adv Exp Med Biol*. 2020; 1207:375–87. https://doi.org/10.1007/978-981-15-4272-5_25 PMID:32671760
3. Lu L, Li W, Chen L, Su Q, Wang Y, Guo Z, Lu Y, Liu B, Qin S. Radiation-induced intestinal damage: latest molecular and clinical developments. *Future Oncol*. 2019; 15:4105–18. <https://doi.org/10.2217/fon-2019-0416> PMID:31746639
4. Holyoake DL, Partridge M, Hawkins MA. Systematic review and meta-analysis of small bowel dose-volume and acute toxicity in conventionally-fractionated rectal cancer radiotherapy. *Radiother Oncol*. 2019; 138:38–44. <https://doi.org/10.1016/j.radonc.2019.05.001> PMID:31136961
5. Garin-Laflam MP, Steinbrecher KA, Rudolph JA, Mao J, Cohen MB. Activation of guanylate cyclase C signaling pathway protects intestinal epithelial cells from acute radiation-induced apoptosis. *Am J Physiol Gastrointest Liver Physiol*. 2009; 296:G740–9. <https://doi.org/10.1152/ajpgi.90268.2008> PMID:19221018
6. Tambouret R, Clement PB, Young RH. Endometrial endometrioid adenocarcinoma with a deceptive pattern of spread to the uterine cervix: a manifestation of stage IIb endometrial carcinoma liable to be misinterpreted as an independent carcinoma or a benign lesion. *Am J Surg Pathol*. 2003; 27:1080–8. <https://doi.org/10.1097/0000478-200308000-00005> PMID:12883240
7. Burzawa J, Gonzales N, Frumovitz M. Challenges in the diagnosis and management of cervical neuroendocrine carcinoma. *Expert Rev Anticancer Ther*. 2015; 15:805–10. <https://doi.org/10.1586/14737140.2015.1047767> PMID:25980782
8. Ibrahim R, Frederickson H, Parr A, Ward Y, Moncur J, Khleif SN. Expression of FasL in squamous cell carcinomas of the cervix and cervical intraepithelial neoplasia and its role in tumor escape mechanism. *Cancer*. 2006; 106:1065–77. <https://doi.org/10.1002/cncr.21697> PMID:16456813
9. Jang C, Chen L, Rabinowitz JD. Metabolomics and Isotope Tracing. *Cell*. 2018; 173:822–37. <https://doi.org/10.1016/j.cell.2018.03.055> PMID:29727671
10. Guijas C, Montenegro-Burke JR, Warth B, Spilker ME, Siuzdak G. Metabolomics activity screening for identifying metabolites that modulate phenotype. *Nat Biotechnol*. 2018; 36:316–20. <https://doi.org/10.1038/nbt.4101> PMID:29621222
11. Cui L, Lu H, Lee YH. Challenges and emergent solutions for LC-MS/MS based untargeted metabolomics in diseases. *Mass Spectrom Rev*. 2018; 37:772–92. <https://doi.org/10.1002/mas.21562> PMID:29486047
12. Emwas AH. The strengths and weaknesses of NMR spectroscopy and mass spectrometry with particular focus on metabolomics research. *Methods Mol Biol*. 2015; 1277:161–93. https://doi.org/10.1007/978-1-4939-2377-9_13 PMID:25677154
13. Zennaro L, Nicolè L, Vanzani P, Cappello F, Fassina A. ¹H-NMR spectroscopy metabonomics of reactive, ovarian carcinoma and hepatocellular carcinoma ascites. *Pleura Peritoneum*. 2020; 5:20200113. <https://doi.org/10.1515/pp-2020-0113> PMID:32566728
14. Chai Y, Wang J, Wang T, Yang Y, Su J, Shi F, Wang J, Zhou X, He B, Ma H, Liu Z. Application of ¹H NMR spectroscopy-based metabonomics to feces of cervical cancer patients with radiation-induced acute intestinal symptoms. *Radiother Oncol*. 2015; 117:294–301. <https://doi.org/10.1016/j.radonc.2015.07.037> PMID:26277430
15. Gramatyka M, Boguszewicz Ł, Ciszek M, Gabryś D, Kulik R, Sokół M. Metabolic changes in mice cardiac tissue after low-dose irradiation revealed by ¹H NMR spectroscopy. *J Radiat Res*. 2020; 61:14–26. <https://doi.org/10.1093/jrr/rrz079> PMID:31840756
16. Bonnaud S, Niaudet C, Legoux F, Corre I, Delpon G, Saulquin X, Fuks Z, Gaugler MH, Kolesnick R, Paris F. Sphingosine-1-phosphate activates the AKT pathway to protect small intestines from radiation-induced endothelial apoptosis. *Cancer Res*. 2010; 70:9905–15. <https://doi.org/10.1158/0008-5472.CAN-10-2043> PMID:21118968
17. Fouillade C, Favaudon V, Vozenin MC, Romeo PH, Bourhis J, Verrelle P, Devauchelle P, Patriarca A, Heinrich S, Mazal A, Dutreix M. [Hopes of high dose-rate radiotherapy]. *Bull Cancer*. 2017; 104:380–4. <https://doi.org/10.1016/j.bulcan.2017.01.012> PMID:28283183

18. Chaves-Pérez A, Yilmaz M, Perna C, de la Rosa S, Djouder N. URI is required to maintain intestinal architecture during ionizing radiation. *Science*. 2019; 364:eaq1165.
<https://doi.org/10.1126/science.aq1165>
PMID:[31147493](https://pubmed.ncbi.nlm.nih.gov/31147493/)
19. Nussbaum ML, Campana TJ, Weese JL. Radiation-induced intestinal injury. *Clin Plast Surg*. 1993; 20:573–80.
[https://doi.org/10.1016/S0094-1298\(20\)31199-8](https://doi.org/10.1016/S0094-1298(20)31199-8)
PMID:[8324995](https://pubmed.ncbi.nlm.nih.gov/8324995/)
20. Duffy J, Urbas A, Niemitz M, Lippa K, Marginean I. Differentiation of fentanyl analogues by low-field NMR spectroscopy. *Anal Chim Acta*. 2019; 1049:161–9.
<https://doi.org/10.1016/j.aca.2018.12.014>
PMID:[30612647](https://pubmed.ncbi.nlm.nih.gov/30612647/)
21. Emwas A, Merzaban JS, Serrai H. Theory and Applications of NMR-Based Metabolomics in Human Disease Diagnosis. *Applications of Nmr Spectroscopy*. 2015: 93–130.
22. Prasad V, Fojo T, Brada M. Precision oncology: origins, optimism, and potential. *Lancet Oncol*. 2016; 17:e81–6.
[https://doi.org/10.1016/S1470-2045\(15\)00620-8](https://doi.org/10.1016/S1470-2045(15)00620-8)
PMID:[26868357](https://pubmed.ncbi.nlm.nih.gov/26868357/)
23. Dey A, Lane DP, Verma CS. Modulating the p53 pathway. *Semin Cancer Biol*. 2010; 20:3–9.
<https://doi.org/10.1016/j.semcancer.2010.02.004>
PMID:[20193765](https://pubmed.ncbi.nlm.nih.gov/20193765/)
24. Abd El All H, Rye A, Duvillard P. p53 immunohistochemical expression of Egyptian cervical carcinoma. *Pathol Oncol Res*. 1999; 5:280–4.
<https://doi.org/10.1053/paor.1999.0134>
PMID:[10607922](https://pubmed.ncbi.nlm.nih.gov/10607922/)
25. Xu F, Na L, Li Y, Chen L. Roles of the PI3K/AKT/mTOR signalling pathways in neurodegenerative diseases and tumours. *Cell Biosci*. 2020; 10:54.
<https://doi.org/10.1186/s13578-020-00416-0>
PMID:[32266056](https://pubmed.ncbi.nlm.nih.gov/32266056/)
26. Du CX, Wang Y. Expression of P-Akt, NFkappaB and their correlation with human papillomavirus infection in cervical carcinoma. *Eur J Gynaecol Oncol*. 2012; 33:274–7.
PMID:[22873098](https://pubmed.ncbi.nlm.nih.gov/22873098/)