**Research Paper** 

### miR-214-3p derived exosomes inhibit inflammation in chondrocytes and cartilage degradation in osteoarthritis rats

#### Zhendeng Lai<sup>1,\*</sup>, Boyi Liao<sup>2,\*</sup>, Peng Fang<sup>1</sup>, Hong Qian<sup>1</sup>, Lei Zhang<sup>1</sup>, Nirong Bao<sup>1</sup>

<sup>1</sup>Department of Orthopaedics, Jinling Hospital, Nanjing University, School of Medicine, Nanjing 210002, China <sup>2</sup>Department of Orthopaedics, The People's Hospital of Wugang City, Wugang 422400, China \*Equal contribution

Correspondence to: Lei Zhang, Nirong Bao; email: <a href="mailto:k185381@163.com">k185381@163.com</a>, <a href="mailto:nirongbao1981@outlook.com">nirongbao1981@outlook.com</a>Keywords: miR-214-3p, exosomes, osteoarthritis, inflammation, cartilage tissueReceived: July 1, 2021Accepted: August 11, 2021Published:

**Copyright:** © 2021 Lai et al. This is an open access article distributed under the terms of the <u>Creative Commons Attribution</u> <u>License</u> (CC BY 3.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

#### ABSTRACT

Background: MicroRNAs (miRs) are regulators of number of cellular process. miRs enclosed within exosomes can be crucial regulators of intercellular signalling and could be an important biomarker of various age associated disorders. Role of exosomal enclosed miRs in osteoarthritis (OA) chondrocytes and synovial fibroblasts (SFBs) remains poorly studied. Here we profiled and studied the effect of synovial fluid derived exosomal miRs on inflammation, survival, proliferation of chondrocyte in correlation with cartilage degeneration.

Methods: Exosomes were isolated from synovial fluid collected from OA subjects and were analysed by transmission electron microscopy. miRs were isolated and were submitted to microarray profiling. Web based PCR analysis was done. Chondrocyte proliferation and colony formation assay was performed. Cell cycle and apoptosis study was done by flow-cytometer. Gene expression was done by qRT-PCR analysis and protein expression by western blot assay. Rat model of OA was created by operating the knee by anterior cruciate ligament and resection of medial menisci (ACLT+MMx) method. Micro-CT analysis, histological analysis, immunohistochemical staining and TUNEL assay was also performed.

Results: About 17 miRs were found to be expressed differentially in the synovial fluid collected from the control and OA subjects. Microarray analysis confirmed, expression of miR-214-3p was significantly downregulated in the synovial fluid exosome of OA subjects. miR-214-3p mimic promoted migration and proliferation of chondrocyte and suppressed apoptosis. Treatment also inhibited the levels of TNF- $\alpha$ , IL-1 $\beta$  and IL-6. SFB-miR-214-3p exosomes suppressed apoptosis and also inflammation in chondrocytes. *In vivo* study suggested that SFB-exosomal miR-214-3p from rats suppressed the formation of osteophytes, prevented degeneration of cartilage and exerted anti-inflammatory and anti-apoptotic effect in articular cartilage tissue.

Conclusion: The findings suggested that SFB-miR-214-3p exosomes can ameliorate chondrocyte inflammation and degeneration of cartilage tissues. The study confirms therapeutic potential of SFB-miR-214-3p exosomes in treating OA.

#### **INTRODUCTION**

Osteoarthritis (OA) is an inflammatory disorder characterised by cartilage degradation, synovial inflammation and remodelling of subchondral bone leading to poor life quality [1, 2]. Acute synovitis is the initial stage change associated with increased levels of pro-inflammatory mediators observed in subjects with OA [3, 4]. Hence to study and monitor the pathophysiological alterations responsible for cartilage and joint associated problems samples of synovial fluid are analysed. Synovial fibroblasts (SFBs) are responsible for secreting synovial fluid which is responsible for lubricating the articular cartilage [5, 6]. OA is characterized by increased expression of pro-inflammatory factors and cytokines such as Tumour necrosis factor (TNF), Interleukin-1β (IL-1 $\beta$ ), IL-2 and IL-7 [7–9]. These mediators are responsible for differentiation and viability of chondrocytes, also they are responsible for activation of matrix metalloproteinases (MMP) and aggrecans [9]. Use of anti-inflammatory therapies in treating the inflamed synovial joint condition is considered a favourable approach in preventing the progression of OA.

Exosomes are extracellular nano-vesicles present in the circulatory system responsible for cellular communication process responsible for regulating various biological processes. Exosomes are carriers and miRs are one of the cargos which is transported by them [10]. In addition to miRs exosomes carry various mRNAs which are transported to the cells by fusion process of cell membrane. Exosomes have recently been identified to be important mediators responsible for interaction between the cells [11]. The proteins and miRs carried by exosomes have been reported to affect the differentiation and survival of cells which affect the progression and development of OA [12, 13]. The exosomes are released by the myriad cells and tissues and SFB. Synovial fluid of osteoarthritis patients has been detected to show the presence of neutrophils derived exosomes [14]. These exosomes influence the pro-inflammatory pathways which govern degradation of extracellular matrix and angiogenesis [15, 16]. Exosomes present in SFBs are the important regulators of local inflammation of joints mediated by SFBs and chondrocytes due to their property to transport miRs and inflammatory proteins to tissues [17].

Recent reports suggest therapies for OA targeting miRs which are involved in regulating the chondrocyte functions [18–20]. In synovial fluid miRs are prone to rapid degradation, however, exosomes prevent this degradation by stabilizing them and hence improving their property of induced differentiation, survival and proliferation in cells [21]. In a report earlier, synovial fluid miR-210 have been found to be an important biomarker of OA [22]. miR-29b-3p was found to overexpressed in synovial fluid of OA patients [23]. Also, in a study earlier, miR-200-c was found to overexpressed in synovial fluid derived exosomes of OA patients [24]. Such exosomes which have prochondrogenic miRs within them could be vital substitute for damaged chondrocytes in osteoarthritis [25]. Research directed towards finding some specific which control miRs would the process of chondrogenesis can be helpful in developing therapies for countering OA.

Synovial fluid is viscous fluid present in the joints for reducing the friction between them. The fluid is rich in exosomes and some important inflammatory proteins hence it can serve as a very important determiner of the exact pathology behind the development and progression of OA [26]. By screening the miRs present in the exosomes of synovial fluid we can get a better understanding about the signalling pathways and the mechanism involved behind the progression of OA, also it would help in identifying the potential targets for treatment.

MiR-214-3p, is miRNA widely distributed in vertebrates, it is found to be associated with hepatic gluconeogenesis [27]. The miR-214-3p has also been associated with skeletal disorders, by suppressing osteogenic differentiation of myoblast cells via targeting the osteoblast specific transcription factor Osterix [28]. A report also found that miR-214-3p targets ATF4 which is an important osteogenic transcriptional factor involved in suppressing bone formation [29]. Also, in a miR-214-3p study was found to promote osteoclastogenesis via PI3K/Akt pathway involving the PTEN protein [30]. It was reported that miR-214-3p was suppressed in IL-1 $\beta$  induced OA mice model, the study confirmed potential involvement of miR-214-3p in OA in vivo [31]. However, the role of exosomal miR-214-3p in OA remains unexplored. Here, we evaluated the functional relevance of exosomal miR-214-3p for understanding its involvement as suppressor of SFBs induced inflammation of chondrocyte and degradation of cartilage.

#### MATERIALS AND METHODS

#### Sampling

About 12 patients (6 females, 6 males) suffering from OA and enrolled for total knee replacement aging between 60 to 65 years were included in the study, the synovial fluid samples were collected from the patents. Similarly normal subjects (n=12) aging between 60 to 65 years with no complaint for OA were also included, synovial fluid sample was collected from them also. Prior to study the patients and normal subjects were informed about the study and written consent was obtained from them, the study was approved by the human ethical review board of Jinling Hospital, Nanjing University, School of Medicine, China (approval no. NU04CE). The svnovial fluid samples collected were immediately processed in laboratory for isolating exosomes (Figure 1A).

#### Isolation of exosome-like vesicle

The exosomes were isolated by ultracentrifugation, briefly, the synovial fluid samples (Figure 1A) and supernatants of SFB were centrifuged in a cooling centrifuge ( $4^{\circ}$  C) at 300 g for 10 minutes, the pellet

formed at the bottom was discarded. The supernatant was again centrifuged at 2000 g for 10 minutes followed by centrifugation at 10000 g for 30 minutes again the supernatants were centrifuged for 100000 g for 60 minutes, the exosomes were obtained as pellet at bottom, which was suspended in PBS and the



Figure 1. Bioinformatic analysis of exosome encapsulated miRs. (A) Procedure for collecting synovial fluid from knee of human volunteers. (B) Western blot analysis of CD63 and TAPA-1 in exosomes like vesicles. (C) Transmission electron microscopic morphological analysis of exosomes like vesicles. (D) Heatmap showing expression of miRs in synovial fluid exosomes of OA subjects. (E) Annotations of KEGG pathway analysis showing various pathways.

suspension was analysed using nanoparticle tracking analysis (NanoSight NTA Systems, Malvern, UK) and transmission electron microscopy Talos L120C TEM (ThermoFisher, USA).

#### Isolation of exosomal miRNA and microarray study

For isolating the miRNAs from obtained exosomes, Ambion PureLink miRNA Isolation Kit (ThermoFisher USA) was used as per supplied instructions. Quantity and quality of isolated miRs was done by using Spectrophotometer (ThermoFisher, USA) and a Bioanalyzer (Agilent, USA) at absorbance maxima of 260 nm. The profiling of exosomes derived miRs was done using GeneChio miRNA 4.0 Array (Applied Biosystems, ThermoFisher, USA). The heat maps for miRs were generated using the hierarchical clustering method.

#### **RT-PCR** analysis of differential expressed miRs

A web-based PCR array analysis was done using the GeneGlobe Data Analysis Centre (Qiagen, USA). The miR PCR data of control and OA groups were normalized as per the procedure described earlier [32]. The data outputs were analysed for differentially expressed miRs from both normal and OA subjects statistically by t-test, the fold changes of miRs greater than 1.5 were included in the study. The miRs of both normal and OA group with Ct values > 35 were rejected and were considered undetected from study so as to minimise the noise of measurements. The differentially expressed miRs were then submitted for DIANA tools using miRpath v.3 as web-based application (http://snf-515788.vm.okeanos.grnet.gr/). The results were further submitted to principal component analysis.

#### Cell culture and transfection

The Sprague-Dawley rats (SD) were used the knee joints were isolated and were submitted to enzymatic digestion for isolating primary SFB and chondrocytes. The rats were supplied by the animal centre of Jinling Hospital, Nanjing University, School of Medicine, China. The synovial tissue and cartilage tissues were treated with 0.25% trypsin for 1 hour and EDTA 0.02% and then divided into small pieces of 0.5 to 1 mm<sup>3</sup>, the pieces were then treated with collagenase-II (0.2%) for 4 hours at room temperature conditions. The cells were collected and suspended in DMEM media containing fetal bovine serum (10%), the cells were then cultured at room temperature with 5 CO<sub>2</sub>. For transfecting, the cells were suspended in DMEM media and plated in 96 well plates for 24 hours until the cells reached confluence of 70 %, after this the

cells were transfected with miR-214-3p mimic or inhibitor (100 pM) as per supplied instructions. The animal study protocols were approved by the animal ethical committee of Jinling Hospital, Nanjing University, School of Medicine, China.

#### **Colony formation and proliferation studies**

Chondrocyte colony formation study was done to evaluate the role of miR-214-3p mimic or inhibitor on proliferation of isolated chondrocytes. Briefly, about 5 x  $10^2$  cells were transferred to culture plates for 24 hours and were cultured for 2 weeks in DMEM media, after 2 weeks the cells were rinsed with PBS and fixed for 20 minutes in formalin (10%) followed by crystal violet staining (0.5%) of 1 hour. The images were recorded with digital microscope for counting the total colony numbers.

For chondrocyte proliferation CCK-8 assay was performed to study the effect of miR-214-3p or SFB-miR-214-3p exosomes on proliferation of chondrocytes. The isolated rat chondrocytes ( $0.5 \times 10^4$  cells/well) were transferred to 96 well plates and treated with sodium nitroprusside followed by treatment of various concentrations of miR-214-3p or SFB-miR-214-3p exosomes for 24 to 48 hours and proliferation was evaluated. The wells were added with CCK-8 reagent 100 µl (10 %) for 2 hours, the wells were studied by microplate reader, the optical activity was observed at 450 nm.

#### Apoptosis and cell cycle analysis

For apoptosis study we performed Annexin V-FITC analysis (ThermoFisher, USA) as per supplied instructions. Briefly, the chondrocytes were centrifuged at 1000 g for 5 minutes two times in cold PBS and then suspended in binding buffer (500 µl) having 5 µl each of Annexin V-FITC and PI. The chondrocytes were then incubated for 15 minutes in dark and were then submitted for flow cytometry analysis (FACScan Beckton Dickson, USA), apoptosis rate was studied with flow cytometer.

For cell cycle analysis the isolated chondrocytes were mixed with miR-214-3p mimic or inhibitor construct along with complete media for 24 hours. The cells were then fixed in ethanol (70%) and maintained at  $20^{\circ}$  C for 24 hours. The cells were rinsed two times with PBS followed by treatment with mixture of Ribonuclease (25µg/ml) and Propidium iodide (50 µg/ml) for 30 minutes. The cells were then submitted to flow cytometry analysis (FACS scan, Beckton Dickson, USA).

## Quantitative real-time polymerase chain reaction (qRT-PCR)

The total RNA was isolated using Trizol reagent (ThermoFisher, USA) from miRs or chondrocytes treated exosomes as per the provided instructions. The RNAs were characterized quantitatively and qualitatively using a spectrophotometer and then cDNA was prepared using PrimeScript RT kit. For qRT-PCR analysis, SYBR premix (Takra Biotech, USA) was used along with qTOWER qRT-PCR system (AnalytikJena USA). GAPDH was selected as loading control, whereas the analysis was carried by  $2(-\Delta\Delta CT)$  method for studying the expression of gene.

#### Western blot analysis

The extracellular vesicles were submitted to lysis using the RIPA buffer added with protease inhibitor cocktail. The resultant was analysed for total proteins using the protein estimation kit (Sigma Aldrich, USA). The proteins were incubated with specific antibodies overnight for obtaining blots. The antibodies used were specific for IL-6 (1:500), IL-1 $\beta$  (1:500), TNF- $\alpha$  (1:500), CD81 (1:1000), CD63 (1:1000), CD9 (1:1000) and HSP70 (1:1000), all the antibodies were obtained from Abcam USA. The blots were detected using Western blot detection system (Thermo Fisher, USA).

#### Animal studies

For the study Sprague Dawley rats (male) weighing between 220-250 g were obtained from the animal centre of Jinling Hospital, Nanjing University, School of Medicine, China. The animal experiments were approved by the Jinling Hospital, Nanjing University, School of Medicine, China (approval number: NU0031AE). The animal studies were done to study the effect of SFB miR-214-3p exosomes on the cartilage tissues of rats. The animal model of OA was created by the ACLT + MMx as described earlier with minor modifications [33]. Briefly, the rats were submitted to pentobarbital anaesthesia (20 mg/kg), the right knee was operated for anterior cruciate ligament and resection of the medical menisci, the sham rats didn't undergo resection. The rats were divided into Sham operated, OA control, SFB-miR-214-3p exosomes, OA+SFBcontrol exosome group and OA+ SFB-miR-214-3p exosomal group. After 1 week of operating the rats, they were shifted to rotating cage for 15/day as described earlier [34]. After 4 weeks post-surgery, the rats injected (intra-articular route) with defined treatments according to groups once in a week, the sham operated group was injected with PBS. After 10 weeks of study, the rats were submitted to micro-CT, TUNEL assay and histopathology.

#### *In vivo* micro-CT analysis

*In vivo* micro-CT analysis was performed using micro-CT systems Bruker Germany for analysing the knee joints. The images were gathered and evaluated for the structure of joints and quantified for mean trabecular thickness (Tb. Th, mm), morphometric indices by measuring the bone volume fraction i.e the ratio of bone volume/total volume (BV/TV,%) and mean trabecular separation (Tb. Sp, mm).

#### **Histological studies**

The rat knee joints were disarticulated and the femoral condyle and tibial plateau were fixed in para formaldehyde (4%) for 24 hours. The tissues samples were decalcified by treating them with EDTA (10%) pH 7.4 for 21 days, then the tissue samples were embedded in paraffin. The tissues samples were then processed in rotary microtome to obtain tissues sections of 5 mm and submitted to haematoxylin and eosin staining (H&E).

#### **TUNEL** assay

The cell apoptosis was done by TUNEL assay with the help of cell death detection kit (ThermoFisher, USA) as the supplied instructions. Briefly, the cartilage tissue isolated from knee joint were exposed to proteinase solution  $(20 \ \mu g/ml)$  for 15 minutes, after then the apoptotic chondrocytes were processed for labelling. The apoptotic cells were quantified using a microscope.

#### Immunohistochemical analysis

The expression levels of TNF- $\alpha$  and IL-1 $\beta$  in cartilage tissues were analysed by immunohistochemical analysis. Briefly, the cartilage tissue sections were incubated for 12 hours at 4° C with primary antibodies. The sections were submitted to diaminobenzidine staining kit (Sigma Aldrich, USA) for detecting the binding of antibodies, then the sections were counterstained with haematoxylin staining. The sections for staining were evaluated by optical microscope and the images of cells were recorded and processed with Image-Pro software.

#### Statistical analysis

The results were presented as mean  $\pm$  % relative standard deviation (RSD), the comparison was done via one-way ANOVA, post hoc analysis was done by student t-test. All the statistics was performed using GraphPad prism Software. Value P<0.05 were confirmed to be significant.

#### Data availability

All data generated or analysed during this study are included in this article. Further any enquiries can be directed to the corresponding author.

#### Statement of ethics

The animal experiments were approved by the Jinling Hospital, Nanjing University, School of Medicine, China. The human study was approved by the Human Ethical Review Board of Jinling Hospital, Nanjing University, School of Medicine, China.

#### **RESULTS**

#### Human synovial fluid exosomes

The synovial fluid samples collected from OA and normal subjects were processed for isolating exosomes using the exosome isolation reagent. The exosomes were submitted to immunoblotting analysis and were confirmed to have significant expression of TAPA-1 and CD63 (Figure 1B). The exosomes showed round structure when observed under TEM (Figure 1C).

#### Expression of miRs in synovial exosomes

The synovial exosomes obtained from OA and normal subjects were analysed for miRs, it was found that among the total 17 miRs, 7 were downregulated and 10 were upregulated in OA patients compared to normal subjects. Supervised clustering approach was utilized for creating heat map which showed variations in the samples of normal and OA subjects. The results clearly suggested variable expression of about 17 miRs in the synovial exosomes of OA subjects compared to normal (Figure 1D). It was found that of the differentially expressed miRs, the expression of miR-214-3p was decreased significantly in the synovial exosomes in OA (OA-Exosomes) subjects compared to exosomes from normal subjects (Non-OA-Exosomes).

#### KEGG pathway analyses of variably expressed miRs

For evaluating the functional role of differentially expressed miRs we compared OA-Exosomes and Non-OA-Exosomes with the help of KEGG functional enrichment analysis. The results of KEGG pathway analysis suggested that the variably expressed miRs were linked to various pathways associated with signal transduction, cell migration, cell proliferation and metabolism (Figure 1E). The KEGG pathway analysis identified pathways such as MAPK signalling, Folate biosynthesis, Notch signalling, mTOR signalling pathway, Wnt signalling pathway, Focal adhesion and insulin signalling pathway (Figure 1E).

#### miR-214-3p encourages proliferation of chondrocytes and inhibits inflammation and apoptosis

As it was found that the expression of miR-214-3p was downregulated significantly in OA patients derived exosomes, we postulated that miR-214-3p may be associated with the pathogenesis of OA. The cell proliferation studies by CCK-8 assay suggested that the miR-214-3p mimics transfected chondrocytes showed improved proliferation compared to control chondrocytes, however the miR-214-3p inhibitor transfected group showed opposite results i.e decreased proliferation of chondrocytes (Figure 2A). Then, we studied the effect of miR-214-3p on the proliferation of chondrocytes by performing the colony formation assay. The outcomes suggested that, transfection of miR-214-3p mimic in chondrocytes increased the colony formation compared to control cells, whereas the chondrocytes transfected with miR-214-3p inhibitor showed decrease in extent of colony formation (Figure 2B). The results of Annexin V FITC/Propidium Iodide staining on apoptosis of chondrocytes suggested that, upon transfection of miR-214-3p mimic decreased the rate of apoptosis and the miR-214-3p inhibitor increased apoptosis compared to control (Figure 2C). It was found that transfection of miR-214-3p resulted in fewer number of chondrocytes in the G1 phase whereas the G2 phase of cell cycle had significantly increased number of chondrocytes (Figure 2D). Also, it was observed that the transfection of miR-214-3p inhibitor resulted in decreased frequency of chondrocytes in S phase of cell cycle and increased frequency in the G1 phase. It was also found that the mRNA as well as protein levels of TNF- $\alpha$ , IL-1 $\beta$  and IL-6 were decreased significantly in chondrocytes transfected with miR-214-3p mimic and increased after miR-214-3p inhibitor transfection (Figure 2E, 2F).

## SFB-miR-214-3p exosomes inhibited inflammation and apoptosis in OA chondrocyte

Morphological analysis of SFB-miR-214-3p exosomes was done and it was found that the exosomes were having approximate diameter of  $100 \pm 5$  nm which was in agreement to the features of exosomes. The miR-214-3p mimic transfected the SFB which was evidenced by high expression levels of miR-214-3p in them (Figure 3A). HSP70, CD9 and CD63 are the important protein markers expressed by exosomes, our results showed that these markers were expressed by the SFB-miR-214-3p exosomes (Figure 3B). The CCK-8 assay suggested that the SFB-miR-214-3p exosomes treated chondrocytes showed increased proliferation upon treatment with or without sodium nitroprusside compared to control chondrocytes (Figure 3C). The apoptosis rate in sodium nitroprusside treated SFB-miR-214-3p exosomes was significantly lower compared to sodium nitroprusside and control SFB-exosomes not showing the expression of miR-214-3p (OA+SFB-control exosome) (Figure 3D). On evaluating the effect of treatments on levels of TNF- $\alpha$ , IL-1 $\beta$  and IL-6 it was observed that treatment of SFB-miR-214-3p exosomes resulted in overexpression of miR-214-3p and inhibition of TNF- $\alpha$ , IL-1 $\beta$  and IL-6 at both protein as well as mRNA levels (Figure 3E, 3F) compared to SFB-control exosomes treated chondrocytes.



**Figure 2. Effect of miR-214-3p on proliferation, apoptosis and inflammatory response on chondrocytes.** (A) CCK-8 analysis showing effect of miR-214-3p mimic and inhibitor on cell proliferation for 24 and 48 hours. (B) Colony formation assay on chondrocytes exposed to miR-214-3p-mimic or inhibitor. (C) Flow cytometry analysis for studying apoptosis index after exposure to miR-214-3p mimic or inhibitor for 24 hours. (D) Cell cycle progression analysis evaluating the effect of miR-214-3p mimic or inhibitor treatment for 24 hours. (E) qRT-PCR analysis for relative mRNA levels of miR-214-3p, IL-1 $\beta$ , IL-6 and TNF- $\alpha$  after exposure to miR-214-3p mimic or inhibitor for 24 hours. (F) Western blot analysis for expression of inflammatory proteins IL-1 $\beta$ , IL-6 and TNF- $\alpha$  after exposing the chondrocytes to miR-214-3p mimic or inhibitor. The results are presented as mean (n=3) ± %RSD. \*P<0.05, \*\*P<0.01, \*\*\*P<0.001 compared to normal control.

## SFB-miR-214-3p exosomes maintained the subchondral bone structure in OA rats

For studying the effect of SFB-miR-214-3p on bone structure micro-CT analysis was done. For the *in vivo* study a rat model of OA was developed, the model was used to study any alterations in cartilages, subchondral bone such as joint space, changes in calcification and formation of osteophyte (Figure 4). It was found that, the OA rats showed significantly decreased BV/TV values and increased Tb. Sp compared to sham-operated rats. Interestingly, the treatment of SFB-miR-214-3p exosomes reversed these changes, thus confirming that SFB-miR-214-3p exosomes maintained the structure of subchondral bone of OA rats.



**Figure 3. Effect of SFB-derived exosomes like vesicles on inflammation and apoptosis in chondrocytes.** (A) qRT-PCR analysis for relative mRNA expression levels of miR-214-3p in SFB transfected with miR-214-3p mimic after 24 hours of incubation. (B) Relative protein levels by western blot analysis for expression of CD9, HSP70 and CD63 in exosomes like vesicles. (C) CCK-8 analysis for cell proliferation after treatment with SFB-miR-214-3p-exo for 24 and 48 hours. (D) Apoptosis study using flow cytometer after treatment of SFb-miR-214-3p-exo for 2 hours before treatment of SNP for 24 hours. (E) qRT-PCR analysis for relative mRNA levels of miR-214-3p, IL-1 $\beta$ , IL-6 and TNF- $\alpha$  after treatment of SFB-miR-214-3p-exo for 2 hours prior to SNP treatment of 24 hours. (F) Western blot analysis for protein levels for expression of inflammatory proteins IL-1 $\beta$ , IL-6 and TNF- $\alpha$  after treatment of SFB-miR-214-3p-exo for 2 hours. The results are presented as mean (n=3) ±%RSD.\*P<0.05, \*\*P<0.01 and \*\*\*P<0.001 compared to SNP+miR-Control-exo.

# SFB-miR-214-3p exosomes inhibited the synovial inflammation induced degeneration of cartilage and synovium in OA rats.

The histological study in OA rats was done by H&E staining (Figure 5). It was observed that against the sham operated rats the OA rats showed irregular articular morphology of cartilages, whereas OA rats treated with SFB-miR-214-3p exosomes showed enhanced thickness of cartilages and surface regularity which was associated with less severe cartilage degradation. The OA rats showed synovial thickening which was significantly decreased in OA + SFB-miR-214-3p exosomes treated rats, the findings were in agreement with the property of SFB-miR-214-3p exosomes to decrease the inflammation of synovial joints.

#### SFB-miR-214-3p exosomes suppressed the apoptosis and inflammation of chondrocytes of articular cartilage in OA rats

The effect of SFB-miR-214-3p exosomes treatment on apoptosis of chondrocytes and SFB in rat model of OA

was evaluated by TUNEL assay. Compared to sham rats, the samples of OA induced rats showed significantly high number of TUNEL positive cells in cartilage (Figure 6A) as well as in the synovial tissues (Figure 6A). However, the chondrocytes treated with SFB-miR-214-3p exosomes showed decreased number of TUNEL positive cells. The immunohistochemical staining analysis was done to evaluate the expression of TNF- $\alpha$  and IL-1 $\beta$  in cartilage tissue sections of OA rats. The findings suggested that the expression of TNF- $\alpha$ and IL-1 $\beta$  were significantly decreased in SFB-miR-214-3p exosomes treated OA rats compared to OA control rats (Figure 6B).

#### **DISCUSSION**

The present work evaluated the potential of synovial fluid derived exosomal miR in modulating the pathogenesis of OA and its effect on degeneration of knee joints of OA rats. We performed miR microarray analysis and found many miRs were expressed differentially in synovial exosomes form OA subjects. It was found that from the 17 variably expressed miRs, the major miRs downregulated were miR-432-5p, miR-138,



Figure 4. SFB-miR-214-3p exosomes maintained the subchondral bone structure in OA rats. Micro-CT analysis showing bones of rats subjected ACLT+MMX OA. Quantitative analysis showing mean trabecular thickness and mean trabecular separation. The results are presented as mean (n=3)  $\pm$  %RSD. \*P<0.05 and \*\*P<0.01 compared to sham group, #P<0.05 compared to OA rats, &P<0.05 compared to OA+miR-control-exo rats.

miR-140, miR-204, miR-211 and miR-214-3p, whereas the miRs upregulated were miR-146a, miR-132, miR-180-a, miR-181-c and miR-181-d. Among all exosomal miRs, miR-214-3p was one of the most downregulated (about 3 fold) in the synovial fluid samples of OA subjects compared to the normal subjects, also prior reports had suggested involvement of miR-214-3p in OA condition [31].

The synovial fluid which is in close contact with articular cartilage and the synovial membrane present in the knee joints, can serve as an important indicator

of any pathological condition [35, 36]. Cell derived exosomes are important indicators for studying the progression of any disease [37, 38]. Hence, in the present work we performed analysis of miRs present within the exosomes isolated from the synovial fluid of osteoarthritis and normal subjects. Morphological confirmed these particles analysis to be exosomes looking into their diameters, also it was found that these particles were present at same levels in the synovial fluid in these subjects. It was also found that the exosomes had significant levels of miRs in them.

## OA + miR-control- OA+miR-214-3pmiR-214-3p-Exos exos Sham OA exos Synovium (H & E Staining) Sham OA miR-214-3p-Exos 200 200 µn OA + miR-control-exos OA+miR-214-3p-Exos

Cartilage tissue (H & E Staining)

Figure 5. SFB-miR-214-3p exosomes inhibited the synovial inflammation induced degeneration of cartilage and synovium in OA rats. H&E staining of cartilage or synovium of OA rats treated with SFB-miR-214-3p-exo showing improved cartilage thickness and regularity.

200 µm

#### 10

00 11

We next performed KEGG pathway analysis for target genes to screen the functional characteristics of miRs which were variably expressed in OA subjects. This screening suggested that the exosomal miRs were associated with some important pathways mainly associated with signal transduction, cell migration, cell metabolism and cell proliferation. The analysis pointed some OA specific cascades such as mTOR, Wnt and MAPK. Very importantly we found that the exosomes derived from synovial fluid had property of endocytosis with chondrocytes leading to alterations of signalling process inside the chondrocytes. Hence, we spiculated that exosome derived from OA subjects may affect the metabolic process in the chondrocytes. The findings of our study confirmed that synovial fluid isolated exosomes of OA subjects affected metabolic process such as inflammatory response and apoptosis in chondrocytes, the findings were in agreement to reports published earlier which suggested that exosomes derived from IL-1 $\beta$ stimulated SFBs could promote increased expression of ADAMTS and MMP-13 in chondrocytes compared to exosomes derived from normal subjects [39]. Our findings showed that, SFB derived exosomes overexpressing miR-214-3p suppressed inflammation in chondrocytes significantly compared to exosomes from control SFBs, indicating that these SFB isolated exosomes can be important in regulating inflammatory response in the synovial space.

Literatures have confirmed that miRs are responsible for influencing the immune and inflammatory reactions targeting the associated genes responsible for apoptosis, differentiation and activation [40].

Hence, we spiculated that exosome derived miRs collected from synovial fluid of OA subjects may inhibit joint remodelling and bone degeneration. As it was documented earlier that miR-214-3p is associated with OA and also our results found that miR-214-3p was significantly downregulated in SFBs of OA subjects, we studied the ability of exosomal miR-214-3p in influencing the pathogenesis of osteoarthritis. We hypothesized that miR-214-3p may inhibit the production of inflammatory cytokine after the chondrocytes were treated with sodium nitroprusside which is majorly used as a nitric oxide donor [41]. Our findings reported that over-expression of miR-214-3p resulted in suppression of sodium nitroprusside induced over expression of TNF-a, IL-1β and IL-6 and proliferation of chondrocytes also the SFB-miR-214-3p exosomes effectively suppressed the progression of OA. Further, we evaluated the effect of SFB-miR-214-3p exosomes on ACLT + MMx induced osteoarthritis rats. Prior, to this it was reported that the intra-articular exosomal delivery can lead to regeneration of cartilage and hence stop the progression of osteoarthritis [42, 43]. In the study, we evidenced that treatment of exosomes in the articular cartilage tissues suppressed the progression of OA significantly and also enhanced the cartilage regeneration thus preventing the cartilage damage in OA rats. Interestingly, SFB-miR-214-3p exosomes treatment significantly attenuated the loss of proteoglycan, irregular surface. inflammation of synovial joints and superficial fibrillation compared to rats receiving treatment of exosomes which showed control all these





defects. The SFB-miR-214-3p exosomes treated OA rats upon micro-CT analysis suggested high bone volume fraction values compared to rats treated with control exosomes, the findings confirmed that treatment of SFB-miR-214-3p exosomes promoted healing in OA rats.

Though our findings indicated that exosomal miR-214-3p could significantly suppress the cartilage tissue degeneration, there are some limitations to our study such as more in-depth molecular analysis is needed for target gene confirmation, also the patient sample size needs to be increased for more concrete results.

In conclusion, the findings of study confirmed that exosomes derived miRs could be important regulators in progression of OA. SFB-miR-214-3p exosomes can target anti-inflammatory pathway thus suppressing the production of pro-inflammatory cytokines involved in progression of OA.

#### **AUTHOR CONTRIBUTIONS**

ZL and BL contributed equally to this work. ZL, BL, NB and LZ designed and performed experiments with PF and HQ. All the authors contributed in preparing the manuscript and read the paper before submission.

#### ACKNOWLEDGMENTS

The authors are thankful to the funding agency National Natural Science Foundation of China for providing financial support. The authors are also grateful to management and staff of Jinling Hospital, Nanjing University, School of Medicine, Nanjing, China for providing the necessary support.

#### **CONFLICTS OF INTEREST**

The authors declare that they have no conflicts of interest.

#### FUNDING

This work was supported by the National Natural Science Foundation of China [grant number 81702170, 81972044, 81772318].

#### **REFERENCES**

 Hunter DJ, Bierma-Zeinstra S. Osteoarthritis. Lancet. 2019; 393:1745–59. <u>https://doi.org/10.1016/S0140-6736(19)30417-9</u> PMID:<u>31034380</u>

- Geyer M, Schönfeld C. Novel Insights into the Pathogenesis of Osteoarthritis. Curr Rheumatol Rev. 2018; 14:98–107. <u>https://doi.org/10.2174/157339711366617080712231</u> <u>2</u> PMID:<u>28782470</u>
- Mathiessen A, Conaghan PG. Synovitis in osteoarthritis: current understanding with therapeutic implications. Arthritis Res Ther. 2017; 19:18. <u>https://doi.org/10.1186/s13075-017-1229-9</u> PMID:<u>28148295</u>
- Felson DT, Niu J, Neogi T, Goggins J, Nevitt MC, Roemer F, Torner J, Lewis CE, Guermazi A, and MOST Investigators Group. Synovitis and the risk of knee osteoarthritis: the MOST Study. Osteoarthritis Cartilage. 2016; 24:458–64. <u>https://doi.org/10.1016/j.joca.2015.09.013</u> PMID:<u>26432512</u>
- Pérez-García S, Carrión M, Villanueva-Romero R, Hermida-Gómez T, Fernández-Moreno M, Mellado M, Blanco FJ, Juarranz Y, Gomariz RP. Wnt and RUNX2 mediate cartilage breakdown by osteoarthritis synovial fibroblast-derived ADAMTS-7 and -12. J Cell Mol Med. 2019; 23:3974–83.

https://doi.org/10.1111/jcmm.14283 PMID:30903650

 Schröder A, Nazet U, Muschter D, Grässel S, Proff P, Kirschneck C. Impact of Mechanical Load on the Expression Profile of Synovial Fibroblasts from Patients with and without Osteoarthritis. Int J Mol Sci. 2019; 20:585.

https://doi.org/10.3390/ijms20030585 PMID:30704030

- Rahmati M, Mobasheri A, Mozafari M. Inflammatory mediators in osteoarthritis: A critical review of the state-of-the-art, current prospects, and future challenges. Bone. 2016; 85:81–90. <u>https://doi.org/10.1016/j.bone.2016.01.019</u> PMID:<u>26812612</u>
- Liu-Bryan R, Terkeltaub R. Emerging regulators of the inflammatory process in osteoarthritis. Nat Rev Rheumatol. 2015; 11:35–44. <u>https://doi.org/10.1038/nrrheum.2014.162</u> PMID:<u>25266449</u>
- 9. Chevalier X, Eymard F, Richette P. Biologic agents in osteoarthritis: hopes and disappointments. Nat Rev Rheumatol. 2013; 9:400–10. <u>https://doi.org/10.1038/nrrheum.2013.44</u> PMID:23545735
- Sanz-Rubio D, Martin-Burriel I, Gil A, Cubero P, Forner M, Khalyfa A, Marin JM. Stability of Circulating Exosomal miRNAs in Healthy Subjects. Sci Rep. 2018; 8:10306.

https://doi.org/10.1038/s41598-018-28748-5 PMID:<u>29985466</u>

- Behera J, Tyagi N. Exosomes: mediators of bone diseases, protection, and therapeutics potential. Oncoscience. 2018; 5:181–95. <u>https://doi.org/10.18632/oncoscience.421</u> PMID:<u>30035185</u>
- Murphy C, Withrow J, Hunter M, Liu Y, Tang YL, Fulzele S, Hamrick MW. Emerging role of extracellular vesicles in musculoskeletal diseases. Mol Aspects Med. 2018; 60:123–28. <u>https://doi.org/10.1016/j.mam.2017.09.006</u> PMID:28965750
- Malda J, Boere J, van de Lest CH, van Weeren P, Wauben MH. Extracellular vesicles — new tool for joint repair and regeneration. Nat Rev Rheumatol. 2016; 12:243–49. <u>https://doi.org/10.1038/nrrheum.2015.170</u> PMID:26729461
- Zhou QF, Cai YZ, Lin XJ. The dual character of exosomes in osteoarthritis: Antagonists and therapeutic agents. Acta Biomater. 2020; 105:15–25. <u>https://doi.org/10.1016/j.actbio.2020.01.040</u> PMID:<u>32006653</u>
- Pourakbari R, Khodadadi M, Aghebati-Maleki A, Aghebati-Maleki L, Yousefi M. The potential of exosomes in the therapy of the cartilage and bone complications; emphasis on osteoarthritis. Life Sci. 2019; 236:116861. <u>https://doi.org/10.1016/j.lfs.2019.116861</u> PMID:<u>31513815</u>
- 16. Boere J, Malda J, van de Lest CH, van Weeren PR, Wauben MH. Extracellular Vesicles in Joint Disease and Therapy. Front Immunol. 2018; 9:2575. <u>https://doi.org/10.3389/fimmu.2018.02575</u> PMID:<u>30483255</u>
- Ni Z, Kuang L, Chen H, Xie Y, Zhang B, Ouyang J, Wu J, Zhou S, Chen L, Su N, Tan Q, Luo X, Chen B, et al. The exosome-like vesicles from osteoarthritic chondrocyte enhanced mature IL-1β production of macrophages and aggravated synovitis in osteoarthritis. Cell Death Dis. 2019; 10:522. <u>https://doi.org/10.1038/s41419-019-1739-2</u> PMID:31285423
- Swingler TE, Niu L, Smith P, Paddy P, Le L, Barter MJ, Young DA, Clark IM. The function of microRNAs in cartilage and osteoarthritis. Clin Exp Rheumatol. 2019 (Suppl 120); 37:40–47. PMID:31621575
- Balaskas P, Goljanek-Whysall K, Clegg P, Fang Y, Cremers A, Emans P, Welting T, Peffers M. MicroRNA Profiling in Cartilage Ageing. Int J Genomics. 2017; 2017:2713725. <u>https://doi.org/10.1155/2017/2713725</u> PMID:<u>28890892</u>

- Nugent M. MicroRNAs: exploring new horizons in osteoarthritis. Osteoarthritis Cartilage. 2016; 24:573–80. <u>https://doi.org/10.1016/j.joca.2015.10.018</u> PMID:26576510
- Guduric-Fuchs J, O'Connor A, Camp B, O'Neill CL, Medina RJ, Simpson DA. Selective extracellular vesiclemediated export of an overlapping set of microRNAs from multiple cell types. BMC Genomics. 2012; 13:357. <u>https://doi.org/10.1186/1471-2164-13-357</u> PMID:22849433
- Xie W, Su W, Xia H, Wang Z, Su C, Su B. Synovial Fluid MicroRNA-210 as a Potential Biomarker for Early Prediction of Osteoarthritis. Biomed Res Int. 2019; 2019:7165406. <u>https://doi.org/10.1155/2019/7165406</u>

PMID:<u>31467907</u>

- Chen C, Chen H. Clinical Diagnosis Value of miR-29b-3p in Peripheral Blood Mononuclear Cells and Synovial Fluid Among Osteoarthritis Patients. Clin Lab. 2019; 65. <u>https://doi.org/10.7754/Clin.Lab.2019.190139</u> PMID:<u>31414759</u>
- 24. Withrow J, Murphy C, Liu Y, Hunter M, Fulzele S, Hamrick MW. Extracellular vesicles in the pathogenesis of rheumatoid arthritis and osteoarthritis. Arthritis Res Ther. 2016; 18:286. <u>https://doi.org/10.1186/s13075-016-1178-8</u> PMID:27906035
- 25. Mao G, Zhang Z, Hu S, Zhang Z, Chang Z, Huang Z, Liao W, Kang Y. Exosomes derived from miR-92a-3p-overexpressing human mesenchymal stem cells enhance chondrogenesis and suppress cartilage degradation via targeting WNT5A. Stem Cell Res Ther. 2018; 9:247.

https://doi.org/10.1186/s13287-018-1004-0 PMID:30257711

- Asghar S, Litherland GJ, Lockhart JC, Goodyear CS, Crilly A. Exosomes in intercellular communication and implications for osteoarthritis. Rheumatology (Oxford). 2020; 59:57–68. <u>https://doi.org/10.1093/rheumatology/kez462</u>
- PMID:<u>31628481</u>
  27. Li K, Zhang J, Yu J, Liu B, Guo Y, Deng J, Chen S, Wang C, Guo F. MicroRNA-214 suppresses gluconeogenesis by targeting activating transcriptional factor 4. J Biol Chem. 2015; 290:8185–95. https://doi.org/10.1074/jbc.M114.633990

PMID:25657009

 Shi K, Lu J, Zhao Y, Wang L, Li J, Qi B, Li H, Ma C. MicroRNA-214 suppresses osteogenic differentiation of C2C12 myoblast cells by targeting Osterix. Bone. 2013; 55:487–94. https://doi.org/10.1016/j.bone.2013.04.002 PMID:23579289

29. Wang X, Guo B, Li Q, Peng J, Yang Z, Wang A, Li D, Hou Z, Lv K, Kan G, Cao H, Wu H, Song J, et al. miR-214 targets ATF4 to inhibit bone formation. Nat Med. 2013; 19:93–100.

https://doi.org/10.1038/nm.3026 PMID:23223004

- 30. Zhao C, Sun W, Zhang P, Ling S, Li Y, Zhao D, Peng J, Wang A, Li Q, Song J, Wang C, Xu X, Xu Z, et al. miR-214 promotes osteoclastogenesis by targeting Pten/PI3k/Akt pathway. RNA Biol. 2015; 12:343–53. <u>https://doi.org/10.1080/15476286.2015.1017205</u> PMID:<u>25826666</u>
- 31. Cao Y, Tang S, Nie X, Zhou Z, Ruan G, Han W, Zhu Z, Ding C. Decreased miR-214-3p activates NF-κB pathway and aggravates osteoarthritis progression. EBioMedicine. 2021; 65:103283. <u>https://doi.org/10.1016/j.ebiom.2021.103283</u> PMID:<u>33714889</u>
- D'haene B, Mestdagh P, Hellemans J, Vandesompele J. miRNA expression profiling: from reference genes to global mean normalization. Methods Mol Biol. 2012; 822:261–72. https://doi.org/10.1007/978-1-61779-427-8 18
- PMID:22144205
  33. Hayami T, Pickarski M, Zhuo Y, Wesolowski GA, Rodan GA, Duong LT. Characterization of articular cartilage and subchondral bone changes in the rat anterior cruciate ligament transection and meniscectomized models of osteoarthritis. Bone. 2006; 38:234–43.

https://doi.org/10.1016/j.bone.2005.08.007 PMID:<u>16185945</u>

- 34. Zhou Y, Liu SQ, Yu L, He B, Wu SH, Zhao Q, Xia SQ, Mei HJ. Berberine prevents nitric oxide-induced rat chondrocyte apoptosis and cartilage degeneration in a rat osteoarthritis model via AMPK and p38 MAPK signaling. Apoptosis. 2015; 20:1187–99. <u>https://doi.org/10.1007/s10495-015-1152-y</u> PMID:<u>26184498</u>
- 35. Gao K, Zhu W, Li H, Ma D, Liu W, Yu W, Wang L, Cao Y, Jiang Y. Association between cytokines and exosomes in synovial fluid of individuals with knee osteoarthritis. Mod Rheumatol. 2020; 30:758–64. <u>https://doi.org/10.1080/14397595.2019.1651445</u> PMID:<u>31370732</u>
- Domenis R, Zanutel R, Caponnetto F, Toffoletto B, Cifù A, Pistis C, Di Benedetto P, Causero A, Pozzi M, Bassini F, Fabris M, Niazi KR, Soon-Shiong P, Curcio F. Characterization of the Proinflammatory Profile of Synovial Fluid-Derived Exosomes of Patients with Osteoarthritis. Mediators Inflamm. 2017; 2017:4814987.

https://doi.org/10.1155/2017/4814987 PMID:<u>28634420</u>

37. Tao SC, Yuan T, Zhang YL, Yin WJ, Guo SC, Zhang CQ. Exosomes derived from miR-140-5p-overexpressing human synovial mesenchymal stem cells enhance cartilage tissue regeneration and prevent osteoarthritis of the knee in a rat model. Theranostics. 2017; 7: 180–95.

https://doi.org/10.7150/thno.17133 PMID:<u>28042326</u>

 Zhou Q, Cai Y, Jiang Y, Lin X. Exosomes in osteoarthritis and cartilage injury: advanced development and potential therapeutic strategies. Int J Biol Sci. 2020; 16:1811–20. https://doi.org/10.7150/ijbs.41637

PMID:32398951

- Kato T, Miyaki S, Ishitobi H, Nakamura Y, Nakasa T, Lotz MK, Ochi M. Exosomes from IL-1β stimulated synovial fibroblasts induce osteoarthritic changes in articular chondrocytes. Arthritis Res Ther. 2014; 16:R163. <u>https://doi.org/10.1186/ar4679</u> PMID:<u>25092378</u>
- 40. Chen JQ, Papp G, Szodoray P, Zeher M. The role of microRNAs in the pathogenesis of autoimmune diseases. Autoimmun Rev. 2016; 15:1171–80. <u>https://doi.org/10.1016/j.autrev.2016.09.003</u> PMID:<u>27639156</u>
- 41. Ryu JS, Jung YH, Cho MY, Yeo JE, Choi YJ, Kim YI, Koh YG. Co-culture with human synovium-derived mesenchymal stem cells inhibits inflammatory activity and increases cell proliferation of sodium nitroprusside-stimulated chondrocytes. Biochem Biophys Res Commun. 2014; 447:715–20. https://doi.org/10.1016/j.bbrc.2014.04.077 PMID:24769201
- 42. Wu J, Kuang L, Chen C, Yang J, Zeng WN, Li T, Chen H, Huang S, Fu Z, Li J, Liu R, Ni Z, Chen L, Yang L. miR-100-5p-abundant exosomes derived from infrapatellar fat pad MSCs protect articular cartilage and ameliorate gait abnormalities via inhibition of mTOR in osteoarthritis. Biomaterials. 2019; 206:87–100. <u>https://doi.org/10.1016/j.biomaterials.2019.03.022</u> PMID:<u>30927715</u>
- 43. Wang R, Jiang W, Zhang L, Xie S, Zhang S, Yuan S, Jin Y, Zhou G. Intra-articular delivery of extracellular vesicles secreted by chondrogenic progenitor cells from MRL/MpJ superhealer mice enhances articular cartilage repair in a mouse injury model. Stem Cell Res Ther. 2020; 11:93.

https://doi.org/10.1186/s13287-020-01594-x PMID:<u>32122385</u>