

Predicting intraventricular hemorrhage growth with a machine learning-based, radiomics-clinical model

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ABSTRACT

We constructed a radiomics-clinical model to predict intraventricular hemorrhage (IVH) growth after spontaneous intracerebral hematoma. The model was developed using a training cohort (N=626) and validated with an independent testing cohort (N=270). Radiomics features and clinical predictors were selected using the least absolute shrinkage and selection operator (LASSO) method and multivariate analysis. The radiomics score (Rad-score) was calculated through linear combination of selected features multiplied by their respective LASSO coefficients. The support vector machine (SVM) method was used to construct the model. IVH growth was experienced by 13.4% and 13.7% of patients in the training and testing cohorts, respectively. The Rad-score was associated with severe IVH and poor outcome. Independent predictors of IVH growth included hypercholesterolemia (odds ratio [OR], 0.12 [95%CI, 0.02-0.90]; p=0.039), baseline Graeb score (OR, 1.26 [95%CI, 1.16-1.36]; p<0.001), time to initial CT (OR, 0.70 [95%CI, 0.58-0.86]; p<0.001), international normalized ratio (OR, 4.27 [95%CI, 1.40, 13.0]; p=0.011), and Rad-score (OR, 2.3 [95%CI, 1.6-3.3]; p<0.001). In the training cohort, the model achieved an AUC of 0.78, sensitivity of 0.83, and specificity of 0.66. In the testing cohort, AUC, sensitivity, and specificity were 0.71, 0.81, and 0.64, respectively. This radiomics-clinical model thus has the potential to predict IVH growth.

INTRODUCTION

Intraventricular hemorrhage (IVH) occurs in up to 54% of intracerebral hemorrhage (ICH) cases [1], and is considered an independent predictor of poor outcome due to the mass effect, obstructive hydrocephalus, and inflammatory meningitis [2–5]. Twelve percent of IVH cases will increase in volume by more than 2 mL.

Radiologists distinguish growth-prone hematoma based on empirical knowledge of location, radiological sign, and morphological characteristics. However, these qualitative

variables are subjective and difficult to standardize. Furthermore, atypical hematomas with fine borders can develop new IVH upon follow-up examinations. These characteristics hamper diagnosis and treatment decisions.

Radiomics is an emerging tool that allows researchers to obtain quantitative features from medical images. Radiomics features can be used to evaluate the tumor spatial heterogeneity and microenvironment and reflect tumor gene patterns [6]. It is widely used for evaluating tumor prognosis, selecting appropriate treatment, and predicting lymph node metastasis [7, 8].

Although some researchers have predicted parenchymal hemorrhage enlargement with radiomics technology [9–11], few have tried to predict IVH growth. In this study, we aimed to develop a model that incorporates clinical and radiomics features to identify patients at high risk for IVH growth in the acute phase of ICH.

RESULTS

Study population

The incidence of IVH growth in the training and testing cohorts were 84 (13.4%) and 37 (13.7%), respectively (Table 1). There was no significant difference in the incidence of IVH growth or baseline characteristics between the cohorts.

Patient characteristics

Among the 626 patients enrolled in the training cohort, those with IVH growth had a lower admission GCS (median [IQR], 12.5 [7.0-15.0] vs. 14.0 [11.0-15.0]; $p < 0.001$), larger parenchymal hemorrhage volume (median [IQR], 19.6 [11.8-30.5] mL vs. 15.9 [9.4-26.0] mL; $p < 0.01$), higher baseline Graeb score (3.0 [0.0, 7.8] vs. 0.0 [0.0, 2.0]; $p < 0.001$), and experienced a shorter time from onset to initial ICH detection (median [IQR], 2.0 [1.5-3.5] h vs. 3.0 [2.0-4.5] h; $p < 0.01$) (Table 2). Patients without a history of hypercholesterolemia (1.0 [1.2%] vs. 76.0 [14.0%]); $p < 0.01$ or hypertension (62.0 [73.8%] vs. 451.0 [83.2%]; $p < 0.05$) were more likely to experience IVH growth. Patients with IVH growth were more likely to have parenchymal hemorrhage expansion (HE) (47.0 [56.0%] vs. 80.0 [14.8%]; $p < 0.001$) and a poor outcome (81.0 [96.4%] vs. 405.0 [74.7%]; $p < 0.01$). GCS, baseline ICH volume, baseline Graeb score, time to initial CT, blood glucose, history of hypercholesterolemia, hypertension, international normalized ratio (INR), and the Rad-score were analyzed by multivariable regression.

Multivariable regression analysis indicated that a history of hypercholesterolemia (odds ratio [OR], 0.12 [95% CI, 0.02-0.90]; $p = 0.039$), baseline Graeb score (OR, 1.26 [95% CI, 1.16-1.36]; $p < 0.001$), time to initial CT (OR, 0.70 [95% CI, 0.58-0.86]; $p < 0.001$), INR (OR, 4.27 [95% CI, 1.40, 13.00]; $p = 0.011$), and Rad-score (OR, 2.30 [95% CI, 1.60-3.30]; $p < 0.001$) (Supplementary Table 2) were independently associated with IVH growth. Multicollinearity was not observed between the independent predictors and IVH growth (VIF for all < 2).

Radiomics analysis

The median ICC of 396 candidate features was 0.96 (IQR, 0.87-0.99). Twenty-eight features were excluded and 368 features with good agreement were

further analyzed by the LASSO regression model. Finally, seven features were selected to construct the radiomics signature (Supplementary Figure 1, Supplementary Table 3). The mean and median ICC of the seven selected features were 0.88 (standard deviation, 0.06) and 0.90 (IQR, 0.85-0.91), respectively. The Rad-score was calculated through the linear combination of selected features multiplied by their respective LASSO coefficients (Supplementary Material 3). The Rad-score was confirmed to be a significant predictor of IVH growth with an optimal cut-off value of -1.7259179 . Patients with a Rad-score of ≥ -1.7259179 were more likely to encounter severe IVH (Graeb score, ≥ 6 , 50.0 [22.6%] vs. 55.0 [13.6%]; $p = 0.004$) and have a poor outcome (GOS, ≤ 3 , 197.0 [89.1%] vs. 289.0 [71.4%]; $p < 0.001$) (Table 3). The results from Figure 1 indicate that the proportion of poor outcomes was lower in individuals with a lower Rad-score and progressively increased with an increasing interquartile Rad-score ($p < 0.001$). We used the Rad-score to predict poor outcome. In the training cohort, the Rad-score achieved an AUC of 0.695, a sensitivity of 0.639, and a specificity of 0.696. In the testing cohort, the AUC, sensitivity, and specificity were 0.665, 0.639, and 0.632, respectively. The Receiver-operator curves are shown in Supplementary Figure 2.

Model performance

Five features, including history of hypercholesterolemia, baseline Graeb score, time to initial CT, INR, and Rad-score, were introduced into the SVM model. In the training cohort, the model yielded an AUC of 0.78, sensitivity of 0.83, and specificity of 0.66. In the testing cohort, the AUC, sensitivity, and specificity were 0.71, 0.81, and 0.64, respectively (Figure 2, Table 4). The confusion matrices results of the model are shown in Supplementary Tables 4, 5.

Evaluation of clinical usefulness

DCA indicated that the radiomics-clinical model had a higher overall net benefit in distinguishing patients at high risk for IVH growth than the single clinical model for most of the threshold probabilities (Figure 3).

DISCUSSION

A radiomics-clinical model predicting IVH growth was established using SVM and showed good performance. The Rad-score was confirmed to be independently associated with severe IVH and poor outcomes. To the best of our knowledge, this is the first application of radiomics to predict IVH growth in a relatively large sample (a total of 896 patients enrolled).

Table 1. Baseline characteristics of patients in the training and testing cohorts (variables were presented as counts [percentages]).

Training cohort (N=626)			Testing cohort (N=270)		
Baseline IVH (%)*	IVH expansion	51.0 (24.2)	Baseline IVH (%)*	IVH expansion	20 (20.0)
	Non-IVH expansion	160.0 (75.8)		Non-IVH expansion	79 (80.0)
No baseline IVH (%)*	New IVH	33.0 (8.0)	No baseline IVH (%)*	New IVH	17 (9.9)
	No IVH	382.0 (92.0)		No IVH	154 (90.1)
IVH growth (%)*		84.0 (13.4)	IVH growth (%)*		37.0 (13.7)

IVH= intraventricular hemorrhage; *Data are the number of patients, with percentages in parentheses.

Table 2. Characteristics of patients in two cohorts and univariate analysis of variables associated with IVH growth.

	Training cohort (N=626)			Testing cohort (N=270)		
	IVH growth	Non-IVH growth	P value	IVH growth	Non-IVH growth	P value
Age (year)	61.5(53.0, 73.8)	60.0(51.0, 69.0)	0.285	65.0(55.0, 68.5)	59.0(50.0, 67.0)	0.045
Sex			0.198			0.927
Male (%)*	61.0(72.6%)	355.0(65.5%)		23.0(62.2%)	143.0(61.4%)	
Female (%)*	23.0(27.4%)	187.0(34.5%)		14.0(37.8%)	90.0(38.6%)	
Hypertension (%)*	62.0(73.8%)	451.0(83.2%)	0.037	25.0(67.6%)	192.0(82.4%)	0.035
Hypercholesterolemia (%)*	1.0(1.2%)	76.0(14%)	0.001	3.0(8.1%)	23.0(9.9%)	0.970
Diabetes (%)*	5.0(6.0%)	53.0(9.8%)	0.260	5.0(13.5%)	30.0(12.9%)	1.000
Prior hemorrhage (%)*	4.0(4.8%)	20.0(3.7%)	0.864	2.0(5.4%)	9.0(3.9%)	1.000
Alcohol-consumption (%)*	25(29.8%)	161(29.7%)	0.991	10.0(27.0%)	61.0(26.2%)	0.913
GCS	12.5(7.0, 15.0)	14.0(11.0, 15.0)	<0.001	10.0(7.0, 13.0)	13.0(10.0, 15.0)	<0.001
Time to initial CT (h)	2.0(1.5, 3.5)	3.0(2.0, 4.5)	0.001	2.5(2.0, 3.5)	3.0(2.0, 4.0)	0.474
Baseline ICH volume (mL)	19.6(11.8, 30.5)	15.9(9.4, 26.0)	0.008	21.1(12.8, 38.0)	17.0(9.0, 27.5)	0.009
Parenchymal HE (%)*	47.0(56.0%)	80.0(14.8%)	<0.001	20.0(54.1%)	35.0(15.0%)	<0.001
Blood glucose (mmol/L)	7.4(6.2, 9.0)	6.7(5.9, 8.0)	0.009	7.2(6.2, 9.2)	6.9(6.0, 8.3)	0.177
PLT count (10 ⁹ /L)	195.5(159.5, 226.8)	206.0 (169.8, 245.3)	0.051	204.0(143.0, 228.0)	201.0(163.0, 246.0)	0.219
INR	1.0(1.0, 1.1)	1.0(1.0, 1.1)	0.087	1.0(1.0, 1.1)	1.0(1.0, 1.0)	0.666
APTT (s)	34.2(31.4, 37.8)	34.0(31.1, 37.2)	0.492	32.9(31.2, 36.0)	33.3(31.2, 36.8)	0.825
Baseline Graeb score	3.0(0.0, 7.8)	0.0(0.0, 2.0)	<0.001	2.0(0.0, 5.0)	0.0(0.0, 2.0)	0.012
Follow-up Graeb score	5.0(3.0, 8.0)	0.0(0.0, 2.0)	<0.001	4.0(3.0, 6.5)	0.0(0.0, 2.5)	<0.001
GOS≤3 (%)*	81.0(96.4%)	405(74.7%)	<0.001	35.0(94.6%)	169.0(72.5%)	0.004
Rad-score	-1.5(-2.0, -1.1)	-2.1(-2.6, -1.6)	<0.001	-1.7(-2.0, -1.2)	-2.1(-2.7, -1.7)	0.001

CT= computed tomography; GCS= Glasgow Coma Scale; GOS= Glasgow Outcome Scale; ICH= intracerebral hemorrhage; HE= hemorrhage expansion; IVH= intraventricular hemorrhage; PLT= platelet; INR= international normalized ratio; APTT= activated partial thromboplastin time; Rad-score= radiomics score.

Continuous variables were presented as medians (interquartile range, [IQR]) and categorical variables were presented as counts (with percentages). P values were calculated by using χ^2 test or Wilcoxon rank-sum test. *Data are the number of patients, with percentages in parentheses.

Table 3. Relationship between Rad-score, severe IVH and poor outcome at discharge. (Variables were presented as counts [percentages]).

N (%)	Severe IVH (Graeb score, ≥6)	Poor outcome (GOS, ≤3)
Rad-score≥-1.7259179 (%)*	50.0(22.6)	197.0(89.1)
Rad-score<-1.7259179 (%)*	55.0(13.6)	289.0(71.4)
P value	0.004	<0.001

P values were calculated by using χ^2 test. *Data are the number of patients, with percentages in parentheses.

Among the seven radiomics features used to construct the Rad-score, features from group GLZSM characterized the texture homogeneity of lesions, and the Feature Haralick Correlation from group GLCM measured the degree of image gray level similarity. Previous reports indicated that hematoma heterogeneity was a sign of active bleeding and could predict hematoma development [10, 12]. We inferred that the radiomics features captured the intrahematoma heterogeneity. Nevertheless, interpreting the association between the radiomics features and the

underlying biological processes is challenging. The Rad-score incorporates multiple radiomics features and serves as a multi-factor panel that reduces the complexity of multi-feature studies [13]. For example, the Rad-score could differentiate between similar hematomas in two patients and specify their different outcomes (Figure 4).

Our study also found that increasing INR levels and baseline Graeb scores are predictors for IVH growth. A higher INR implies longer prothrombin time and worse

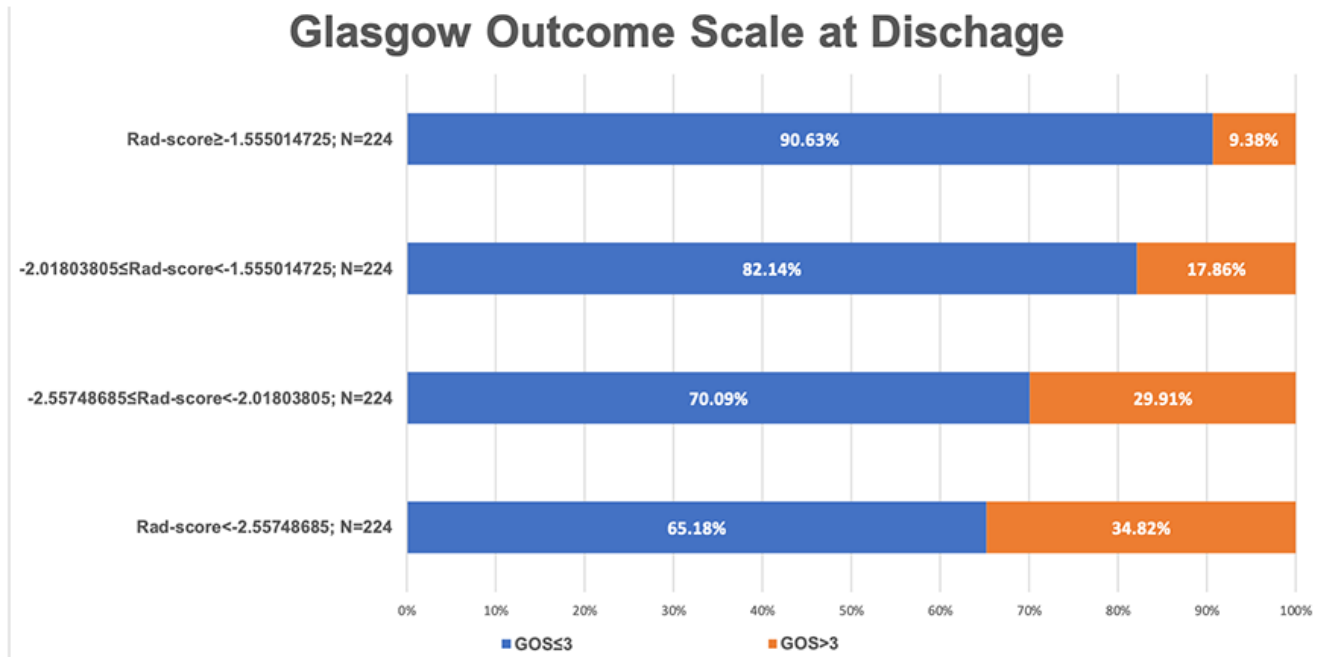


Figure 1. A bar chart demonstrating the relationship between the Rad-score and glasgow outcome scale at discharge.

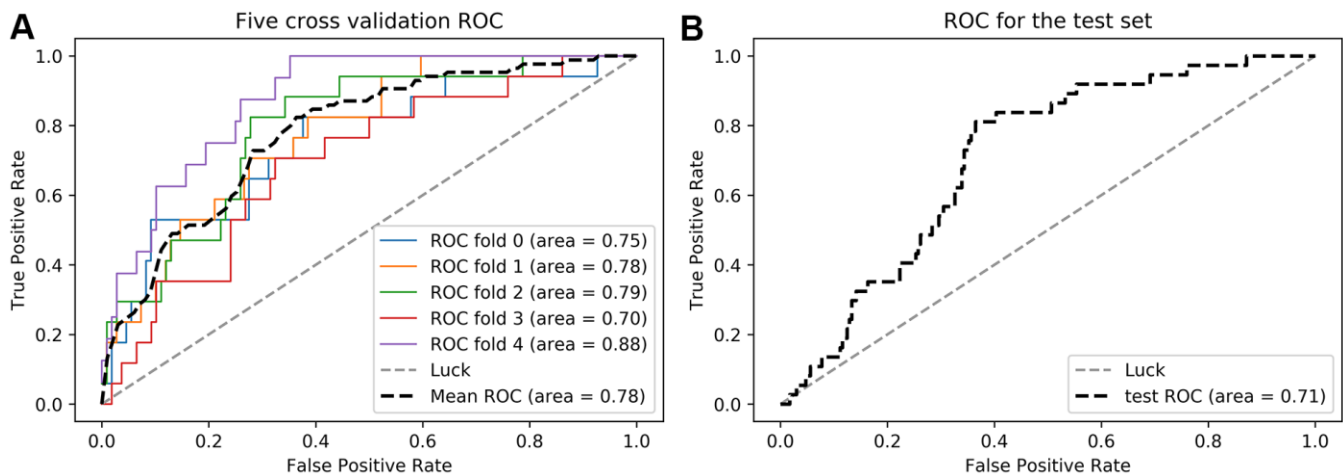


Figure 2. Receiver operator curves (ROC) of the radiomics-clinical model in the training (A) and testing (B) cohorts.

Table 4. Performance of the radiomics-clinical model.

	ACC	AUC	Sensitivity	Specificity	PPV	NPV
Training cohort	0.75	0.78	0.83	0.66	0.27	0.96
Testing cohort	0.71	0.71	0.81	0.64	0.26	0.95

ACC=area under the receiver operating curve; ACC=accuracy; PPV=positive predictive value; NPV=negative predictive value.

coagulation function, which could cause bleeding and lead to further IVH growth, as seen in both ventricular [14] and parenchymal hemorrhage growth [15–18]. The Graeb score is a grading system of 0–12, which can be used to quantify the amount of blood in each ventricle [19]. Increases in the Graeb score indicates a larger ventricular hemorrhage and can be used to predict IVH growth.

A history of hypercholesterolemia was associated with a lower risk of IVH growth in our study. Higher serum cholesterol is negatively correlated to ICH volume and HE risk in parenchymal hematoma [20, 21]. There is also a protective association between hypercholesterolemia and ICH risk (especially in non-lobar lesions) [22–24], regardless of whether the patient uses statins [20, 22]. These correlations might be explained by the role of serum cholesterol in maintaining vascular integrity and

promoting platelet aggregability [21]. Moreover, a higher level of triglycerides is related to a lower rate of deep microbleeds [25]. Thus, hypercholesterolemia may contribute to the maintenance of deep penetrating arterioles [22] and prevent IVH growth.

An increasing number of studies have focused on the contribution of IVH severity to prognosis and the tools for grading IVH extent [1, 26, 27]. A Graeb score of ≥ 5 was reported as an independent predictor of poor outcome [1]. External ventricular drain (EVD) placement or prior treatment was recommended for patients with a Graeb score of >5 [28]. Thus, we defined patients with a follow-up Graeb score of ≥ 6 as having severe IVH. Our results showed that the Rad-score could predict IVH growth, and distinguish patients who might develop severe IVH (Graeb score, ≥ 6) or have poor outcomes (GOS, ≤ 3) at the first CT scan.

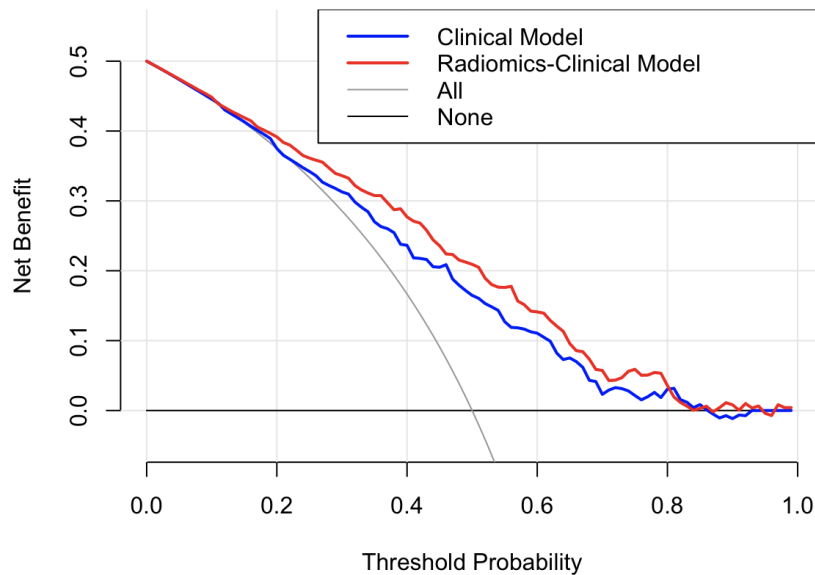


Figure 3. Decision curve analysis for the two models. The y-axis indicates the net benefit; the x-axis indicates threshold probability. The grey line represents the assumption that all patients have IVH growth. The black line represents the assumption that no patients have IVH growth. The blue line and red line represent the net benefit of the single clinical model and radiomics-clinical model, respectively. The radiomics-clinical model had a higher net benefit compared with the single clinical model across most threshold probabilities

Net benefit = $TPR_R P - \frac{R}{1-R} FPR_R (1-P)$, P represents the prevalence of the disease; R represents the threshold probability; TPR= true positive rate; FPR= false positive rate).

Currently, IVH management involves EVD, intraventricular fibrinolysis, neuro-endoscopic procedure, and lumbar drainage [29–31]. However, there is no precise clinical threshold that determines the need for EVD or thrombolysis [15]. Furthermore, there are complications regardless of the method chosen, and the net benefit of these invasive therapies remains unclear [5]. Some studies reported no significant difference in the outcome of patients who received two different treatments [32]. This emphasizes the need for early identification of patients at high risks for IVH growth and offering them targeted treatment when the rate of hematoma growth is highest [2, 5].

The radiomics-clinical model in our study can identify the high-risk patient group by a noninvasive method. Accordingly, this can help clinicians judge the patient's condition and select a treatment.

There were several limitations in our study. First, this is a retrospective study. The time for repeating the CT

scan is varied, which may underestimate the extent of IVH expansion and hydrocephalus. Second, the single-center enrollment is limited in its generalizability. Hence, a future prospective and multi-institutional study is needed. Third, it takes approximately two to five minutes to segment a complete ROI. This manual delineation of hematomas is time-consuming and has a relatively low reproducibility. Consequently, we are exploring the feasibility of an automated or semi-automated delineation of hematoma. Finally, a threshold of 2 mL may exclude patients with a volume increase of <2 mL but who still developed severe IVH. For instance, a previous study found an expansion of 1 mL in IVH volume was associated with poor outcomes [3]. Therefore, additional studies are required to further confirm the cut-off value that correlates to poor outcome.

In conclusion, we confirmed that the Rad-score at admission was associated with severe IVH and poor outcome. Our model incorporates radiomics and clinical

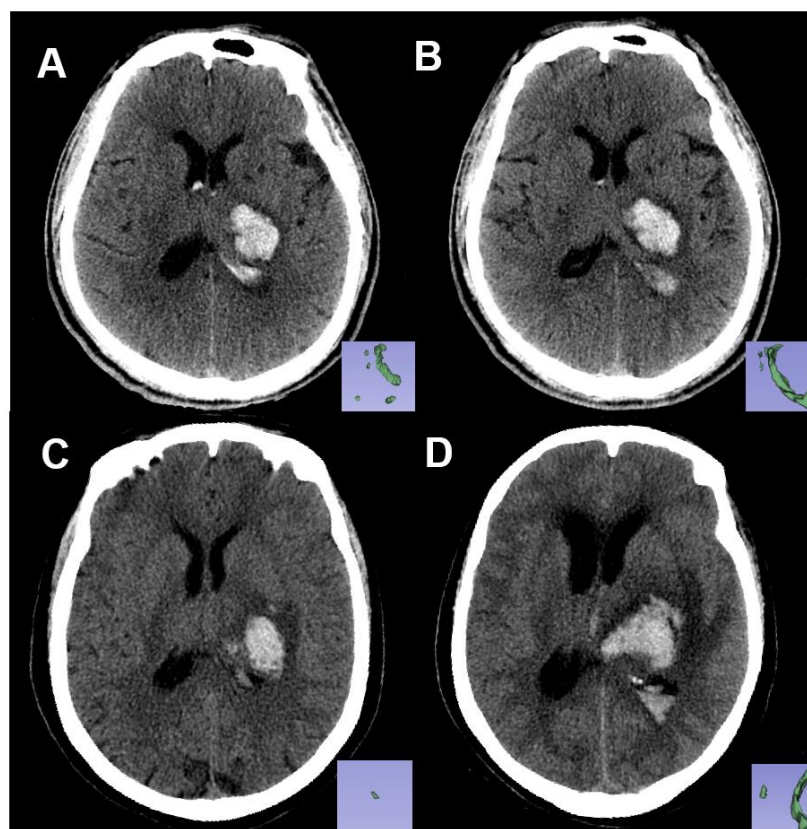


Figure 4. Non-contrast CT images of two patients with similar hematoma but different experiences. Pictures in the lower right corner are the 3-D images of the IVH. Image (A, B) are the baseline and follow-up CT images, respectively, of patient A: a 61-year-old male who had a Rad-score of -2.2119681 (<-1.7259179). Image (C, D) are the baseline and follow-up CT images, respectively, of patient B: a 67-year-old female with a Rad-score of -1.6176548 (>-1.7259179). Within 24 hours from symptom onset, the IVH volume of patient A changed from 5.62 mL to 6.84 mL, and the IVH volume of patient B changed from 0.5 mL to 12.4 mL. Patient B experienced IVH growth while patient A did not.

variables and was developed using the SVM method. The radiomics-clinical model predicted IVH growth with good performance and may help clinicians target patients who have a high IVH growth risk.

MATERIALS AND METHODS

Our study was approved by the Medical Ethics Committee of The First Affiliated Hospital of Wenzhou Medical University and written informed consent was waived.

Patients and clinical data

Patients with ICH seen between September 2013 and August 2018 in The First Affiliated Hospital of Wenzhou Medical University were retrospectively reviewed. Those who were >18 years old and received baseline and follow-up CT scans within 6 h and 72 h from the onset were included. Exclusion criteria were as follows: (1) secondary ICH caused by aneurysm, arteriovenous malformation, neoplasm, hemorrhagic infarction, or traumatic brain injury; (2) surgery or interventional therapy before follow-up CT scan; (3) primary IVH; (4) non-deep ICH location (lobar, cerebellum or brain stem hematoma); (5) use of anticoagulants or antiplatelet drugs before ICH onset; and (6) CT images with severe artifacts. Finally, a total of 896 patients were included and randomly divided into the training (N=626) and testing (N=270) cohorts.

Demographic data (age and sex), medical history (history of hypertension, diabetes, hypercholesterolemia, hemorrhage, and alcohol consumption), initial clinical data (Glasgow coma scale, time to initial CT, and baseline ICH volume), and laboratory data (blood glucose, platelet count, international normalized ratio, and activated partial thromboplastin time) were recorded after admission. The Glasgow Outcome Scale (GOS) was evaluated at discharge, and poor outcome was defined as having a GOS of ≤ 3 [33, 34].

Neuroimage acquisition and analysis

New IVH refers to patients who had no baseline IVH but developed a new IVH lesion in follow-up CT images (<72 h) (Figure 5A, 5B). IVH expansion was defined as an absolute increase from the baseline IVH volume of >2 mL between the initial and follow-up CT images (Figure 5C, 5D). We chose a threshold of 2 mL to define IVH expansion because it is correlated with poor outcome and mortality [32]. IVH growth includes new and expanding IVH. We restricted hematoma to the deep brain region for several reasons. First, deep ICH locations are more likely to have HE [35]. Second, hematomas in the deep region (thalamus and basal

ganglia) are closer to ventricle systems and have a higher risk of IVH growth and poor outcome [36, 37]. Third, in this exploratory study, we tried to exclude as many confounders as possible. Thus, we decided to exclude the non-deep hematomas and focus on the hematomas that had a greater risk for IVH growth (deep hematomas).

All CT images were acquired using a 64-channel multidetector CT scanner (LightSpeed VCT 64; GE Medical Systems, Milwaukee, WI, USA) with a scan thickness of 5 mm, reconstruction interval of 5 mm, tube voltage of 120 kV, tube current of 80 mAs, and matrix size of 512×512 . Radiological data was acquired by two radiologists (two years of experience each) who were blinded to patients' information. The baseline and follow-up IVH volume were measured using the "level tracing" function of 3D Slicer software (version 4.10.2; <http://www.slicer.org>). We also evaluated IVH severity by calculating the Graeb score [19] from the baseline and follow-up CT images. A Graeb score of ≥ 6 was defined as severe IVH based on the literature for IVH severity indicators [28, 33].

The segmentation process followed a consistent standard. Regions of interest (ROIs) were first manually segmented along the hematoma profile on each slice of non-enhanced CT (NECT) images (Figures 5, 6). Skull, peri-hemorrhagic edema, and normal brain parenchyma were manually excluded. All ROIs were first segmented by a radiologist with two years of experience. Then, to measure the inter-observer segmentation reproducibility, 100 images were randomly selected to be segmented by another radiologist with five years of experience [38, 39]. An inter-class correlation coefficient (ICC) of >0.75 was considered a good inter-observer agreement. A window width of 70 and a window level of 35 were set to clearly distinguished hematomas from brain parenchyma. All ROIs were examined by a senior radiologist with 10 years of experience.

A total of 396 radiomics features were extracted using the Artificial Intelligence Kit software (version 3.0.0.R; GE Healthcare). There were six categories: (1) Histogram, (2) Texture, (3) Form factor, (4) Grey level co-occurrence matrix (GLCM), (5) Grey level run-length matrix (GLRLM), and (6) Gray level size zone matrix (GLSZM) (more details about extracted features are shown in the Supplementary Table 1). To remove the unit data limits of each feature, the extracted features were standardized by z-score (Supplementary Material 1). The least absolute shrinkage and selection operator (LASSO) is suitable for feature selection in high-dimensional data [40]. Hence, we used the LASSO method to select the most valuable features. The

parameter (λ) is a penalty parameter that varies for each model fitting step. As the value of λ increased, radiomics features with non-zero coefficients decreased. The optimal λ was selected by a 10-fold cross-validation, and the IVH growth-related radiomics features were subsequently chosen. The radiomics score (Rad-score) was calculated through the linear combination of selected features by multiplying with their respective LASSO coefficients. We categorized the Rad-score into groups by quartiles and cut-off value

to explore the association between Rad-score, IVH severity, and clinical outcome. The workflow of ROI segmentation, feature extraction, and model construction is shown in Figure 6.

Statistical analysis

Statistical analysis was performed with SPSS (version 24.0; IBM, Armonk, NY, USA) and R (version 3.6.1; <http://www.R-project.org>). Continuous variables are

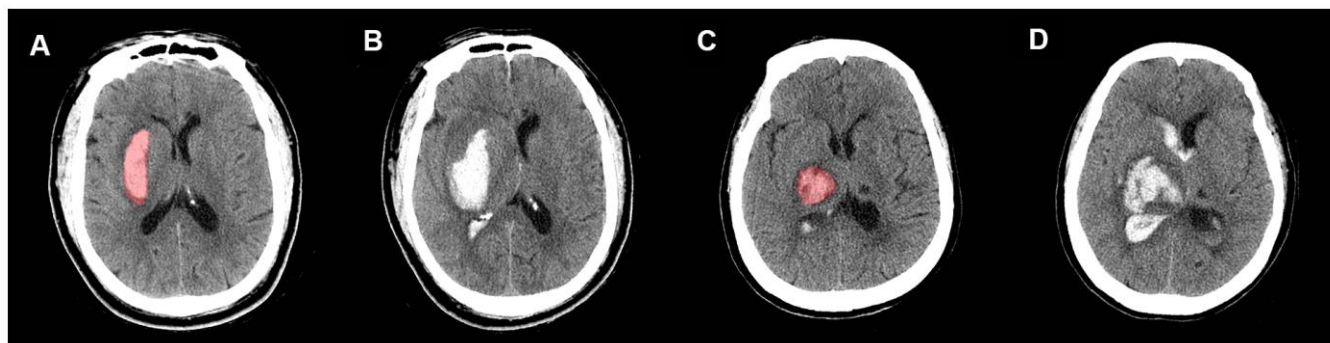


Figure 5. Representative illustration of new IVH, IVH expansion, and manual region of interest (ROI) segmentation. Images (A, B) are non-contrast CT images (axial view) of a 54-year-old male who experienced a new IVH. There was no baseline IVH (A), but the hematoma broke into ventricles on the follow-up CT (B). Images (C, D) are non-contrast CT images (axial view) of a 68-year-old female who experienced IVH expansion; (C) shows an initial IVH with a volume of 2.53 mL; follow-up CT (D) shows that the volume of IVH increased to 22.31 mL within 72 h.

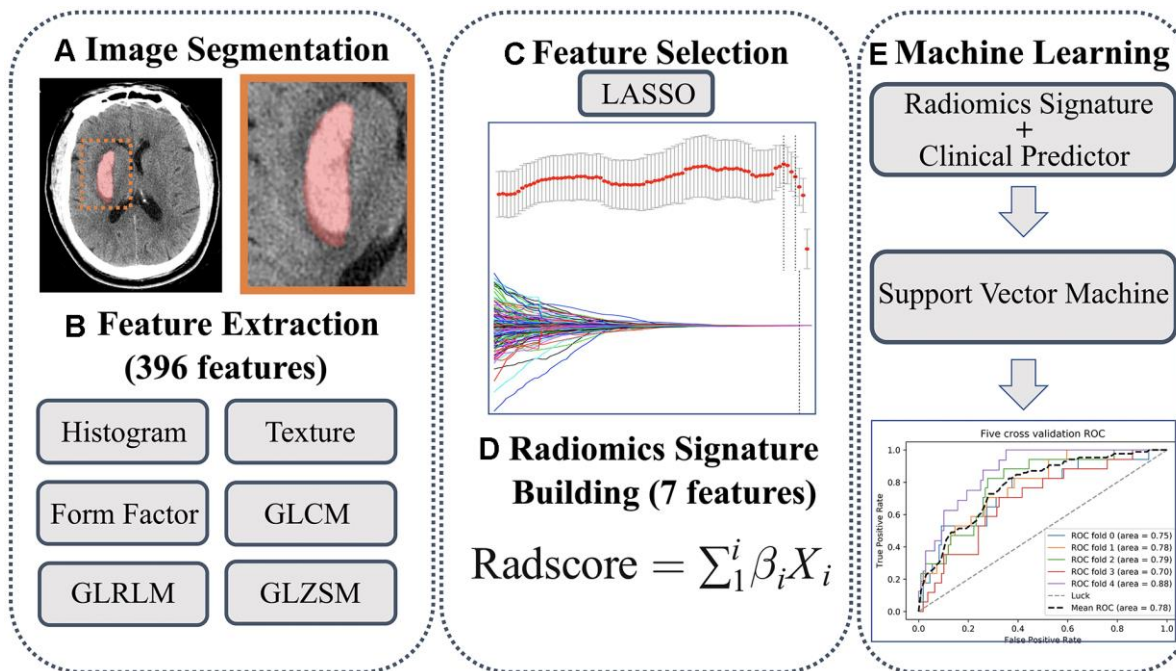


Figure 6. (A) Regions of interest were manually segmented. **(B)** A total of 396 features were extracted. **(C)** Features were selected using LASSO method. **(D)** Rad-score was calculated. **(E)** Predicting model was developed using support vector machine.

presented as medians (interquartile range, [IQR]), and categorical variables are shown as counts (with percentages). First, we performed univariate analysis to select potential clinical risk factors of IVH growth. Differences in continuous variables (age, time to initial CT, ICH volume, blood glucose, platelet count, international normalized ratio, Rad-score, and activated partial thromboplastin time) were examined using Student t-tests or Wilcoxon rank-sum test. The χ^2 test or Fisher exact test (two-tailed) were performed for categorical variables (sex, GCS, history of hypertension, diabetes, hypercholesterolemia, hemorrhage, GOS, and alcohol consumption). A multivariable logistic regression with an enter method was performed to identify factors that were independently associated with IVH growth. Variance inflation factor (VIF) was used to detect multicollinearity, and a VIF of ≥ 5 was defined as multicollinearity [41]. Receiver-operator curve (ROC) analysis was conducted to derive the Rad-score cut-off value for IVH growth prediction, and the optimal cutoff value was selected using Youden's index. A two-sided p value of <0.05 indicates a statistical difference.

Machine learning and model performance evaluation

Support vector machine (SVM) is a type of supervised machine learning method that classifies data points by maximizing the margin between classes in a high-dimensional space [42]. The independent predictors identified in multivariable analysis were introduced into the SVM model. A five-fold cross-validation was used in the training cohort to determine the optimal hyperparameter, reduce overfitting, construct a stable model, and evaluate model performance [43]. The independent testing cohort was used to simulate the prediction and further test the model performance. (additional details about SVM modeling are shown in Supplementary Material 2). The machine learning process was performed using the scikit-learn packages (0.21.3) of Python (version 3.7; <http://www.python.org>).

The model performance was evaluated by the area under the receiver operating curve (AUC). Confusion matrix-derived metrics, including accuracy (ACC), sensitivity, specificity, true positive rate (TPR), true negative rate (TNR), positive predictive value (PPV), and negative predictive value (NPV), were also calculated.

Evaluation of clinical usefulness

Decision curve analysis (DCA) measures the clinical utility of models by quantifying the net benefits at different threshold probabilities [44]. DCA was performed to compare the overall net benefits of the

radiomics-clinical model and single clinical model without the Rad-score in the cohort. The net benefit was defined as the summation of benefits minus the holistic cost [45], and calculated using the formula:

$$\text{Net benefit} = \text{TPR}_R \cdot P - \frac{R}{1-R} \cdot \text{FPR}_R \cdot (1-P), \text{ where } P \text{ is}$$

the disease prevalence; R is a threshold probability at which a patient will opt for treatment; TPR_R represents the proportion of cases with a model-calculated risk above threshold probability; and FPR_R represents the proportion of cases with a model-calculated risk lower than the threshold probability [45].

Abbreviations

AUC: area under the receiver operating curve; GOS: Glasgow Outcome Scale; ICC: inter-class correlation coefficient; ICH: intracerebral hematoma; INR: international normalized ratio; IVH: Intraventricular hemorrhage; LASSO: least absolute shrinkage and selection operator; OR: odds ratio; ROC: receiver-operator curve; SVM: support vector machine.

AUTHOR CONTRIBUTIONS

DZ wrote the manuscript. DZ and QC interpreted the data and prepared the tables and figures. QZ, WC, and XY revised the manuscript for intellectual content. YX, CZ, MZ, and CC acquired the data. YY and XY contributed to the conception and design of the study. All co-authors read and revised the article.

CONFLICTS OF INTEREST

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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REFERENCES

1. Trifan G, Arshi B, Testai FD. Intraventricular Hemorrhage Severity as a Predictor of Outcome in Intracerebral Hemorrhage. *Front Neurol*. 2019; 10:217. <https://doi.org/10.3389/fneur.2019.00217> PMID:[30915027](https://pubmed.ncbi.nlm.nih.gov/30915027/)

2. Garton T, Hua Y, Xiang J, Xi G, Keep RF. Challenges for intraventricular hemorrhage research and emerging therapeutic targets. *Expert Opin Ther Targets*. 2017; 21:1111–22.
<https://doi.org/10.1080/14728222.2017.1397628>
PMID:[29067856](https://pubmed.ncbi.nlm.nih.gov/29067856/)
3. Yogendrakumar V, Ramsay T, Fergusson D, Demchuk AM, Aviv RI, Rodriguez-Luna D, Molina CA, Silva Y, Dzialowski I, Kobayashi A, Boulanger JM, Lum C, Gubitza G, et al, and the PREDICT/Sunnybrook CTA Study Group. New and expanding ventricular hemorrhage predicts poor outcome in acute intracerebral hemorrhage. *Neurology*. 2019; 93:e879–88.
<https://doi.org/10.1212/WNL.0000000000008007>
PMID:[31371565](https://pubmed.ncbi.nlm.nih.gov/31371565/)
4. Qureshi AI, Mendelow AD, Hanley DF. Intracerebral haemorrhage. *Lancet*. 2009; 373:1632–44.
[https://doi.org/10.1016/S0140-6736\(09\)60371-8](https://doi.org/10.1016/S0140-6736(09)60371-8)
PMID:[19427958](https://pubmed.ncbi.nlm.nih.gov/19427958/)
5. Cordonnier C, Demchuk A, Ziai W, Anderson CS. Intracerebral haemorrhage: current approaches to acute management. *Lancet*. 2018; 392:1257–68.
[https://doi.org/10.1016/S0140-6736\(18\)31878-6](https://doi.org/10.1016/S0140-6736(18)31878-6)
PMID:[30319113](https://pubmed.ncbi.nlm.nih.gov/30319113/)
6. Lambin P, Leijenaar RT, Deist TM, Peerlings J, de Jong EE, van Timmeren J, Sanduleanu S, Larue RT, Even AJ, Jochems A, van Wijk Y, Woodruff H, van Soest J, et al. Radiomics: the bridge between medical imaging and personalized medicine. *Nat Rev Clin Oncol*. 2017; 14:749–62.
<https://doi.org/10.1038/nrclinonc.2017.141>
PMID:[28975929](https://pubmed.ncbi.nlm.nih.gov/28975929/)
7. Huang YQ, Liang CH, He L, Tian J, Liang CS, Chen X, Ma ZL, Liu ZY. Development and Validation of a Radiomics Nomogram for Preoperative Prediction of Lymph Node Metastasis in Colorectal Cancer. *J Clin Oncol*. 2016; 34:2157–64.
<https://doi.org/10.1200/JCO.2015.65.9128>
PMID:[27138577](https://pubmed.ncbi.nlm.nih.gov/27138577/)
8. Gillies RJ, Kinahan PE, Hricak H. Radiomics: Images Are More than Pictures, They Are Data. *Radiology*. 2016; 278:563–77.
<https://doi.org/10.1148/radiol.2015151169>
PMID:[26579733](https://pubmed.ncbi.nlm.nih.gov/26579733/)
9. Li H, Xie Y, Wang X, Chen F, Sun J, Jiang X. Radiomics features on non-contrast computed tomography predict early enlargement of spontaneous intracerebral hemorrhage. *Clin Neurol Neurosurg*. 2019; 185:105491.
<https://doi.org/10.1016/j.clineuro.2019.105491>
PMID:[31470362](https://pubmed.ncbi.nlm.nih.gov/31470362/)
10. Shen Q, Shan Y, Hu Z, Chen W, Yang B, Han J, Huang Y, Xu W, Feng Z. Quantitative parameters of CT texture analysis as potential markers for early prediction of spontaneous intracranial hemorrhage enlargement. *Eur Radiol*. 2018; 28:4389–96.
<https://doi.org/10.1007/s00330-018-5364-8>
PMID:[29713780](https://pubmed.ncbi.nlm.nih.gov/29713780/)
11. Xu W, Ding Z, Shan Y, Chen W, Feng Z, Pang P, Shen Q. A Nomogram Model of Radiomics and Satellite Sign Number as Imaging Predictor for Intracranial Hematoma Expansion. *Front Neurosci*. 2020; 14:491.
<https://doi.org/10.3389/fnins.2020.00491>
PMID:[32581674](https://pubmed.ncbi.nlm.nih.gov/32581674/)
12. Barras CD, Tress BM, Christensen S, MacGregor L, Collins M, Desmond PM, Skolnick BE, Mayer SA, Broderick JP, Diringner MN, Steiner T, Davis SM, and Recombinant Activated Factor VII Intracerebral Hemorrhage Trial Investigators. Density and shape as CT predictors of intracerebral hemorrhage growth. *Stroke*. 2009; 40:1325–31.
<https://doi.org/10.1161/STROKEAHA.108.536888>
PMID:[19286590](https://pubmed.ncbi.nlm.nih.gov/19286590/)
13. Zhao W, Xu Y, Yang Z, Sun Y, Li C, Jin L, Gao P, He W, Wang P, Shi H, Hua Y, Li M. Development and validation of a radiomics nomogram for identifying invasiveness of pulmonary adenocarcinomas appearing as subcentimeter ground-glass opacity nodules. *Eur J Radiol*. 2019; 112:161–68.
<https://doi.org/10.1016/j.ejrad.2019.01.021>
PMID:[30777206](https://pubmed.ncbi.nlm.nih.gov/30777206/)
14. Biffi A, Battey TW, Ayres AM, Cortellini L, Schwab K, Gilson AJ, Rost NS, Viswanathan A, Goldstein JN, Greenberg SM, Rosand J. Warfarin-related intraventricular hemorrhage: imaging and outcome. *Neurology*. 2011; 77:1840–46.
<https://doi.org/10.1212/WNL.0b013e3182377e12>
PMID:[22049204](https://pubmed.ncbi.nlm.nih.gov/22049204/)
15. Dey M, Jaffe J, Stadnik A, Awad IA. External ventricular drainage for intraventricular hemorrhage. *Curr Neurol Neurosci Rep*. 2012; 12:24–33.
<https://doi.org/10.1007/s11910-011-0231-x>
PMID:[22002766](https://pubmed.ncbi.nlm.nih.gov/22002766/)
16. Huttner HB, Schellinger PD, Hartmann M, Köhrmann M, Juettler E, Wikner J, Mueller S, Meyding-Lamade U, Strobl R, Mansmann U, Schwab S, Steiner T. Hematoma growth and outcome in treated neurocritical care patients with intracerebral hemorrhage related to oral anticoagulant therapy: comparison of acute treatment strategies using vitamin K, fresh frozen plasma, and prothrombin complex concentrates. *Stroke*. 2006; 37:1465–70.
<https://doi.org/10.1161/01.STR.0000221786.81354.d6>
PMID:[16675739](https://pubmed.ncbi.nlm.nih.gov/16675739/)

17. Oge DD, Topcuoglu MA, Gocmen R, Arsava EM. The dynamics of hematoma surface regularity and hematoma expansion in acute intracerebral hemorrhage. *J Clin Neurosci*. 2020; 74:160–63. <https://doi.org/10.1016/j.jocn.2020.01.081> PMID:[32089386](https://pubmed.ncbi.nlm.nih.gov/32089386/)
18. Huynh TJ, Aviv RI, Dowlatshahi D, Gladstone DJ, Laupacis A, Kiss A, Hill MD, Molina CA, Rodriguez-Luna D, Dzialowski I, Silva Y, Kobayashi A, Lum C, et al, and PREDICT/Sunnybrook CTA Investigators. Validation of the 9-Point and 24-Point Hematoma Expansion Prediction Scores and Derivation of the PREDICT A/B Scores. *Stroke*. 2015; 46:3105–10. <https://doi.org/10.1161/STROKEAHA.115.009893> PMID:[26463691](https://pubmed.ncbi.nlm.nih.gov/26463691/)
19. Graeb DA, Robertson WD, Lapointe JS, Nugent RA, Harrison PB. Computed tomographic diagnosis of intraventricular hemorrhage. Etiology and prognosis. *Radiology*. 1982; 143:91–96. <https://doi.org/10.1148/radiology.143.1.6977795> PMID:[6977795](https://pubmed.ncbi.nlm.nih.gov/6977795/)
20. Chang JJ, Katsanos AH, Khorchid Y, Dillard K, Kerro A, Burgess LG, Goyal N, Alexandrov AW, Alexandrov AV, Tsvigoulis G. Higher low-density lipoprotein cholesterol levels are associated with decreased mortality in patients with intracerebral hemorrhage. *Atherosclerosis*. 2018; 269:14–20. <https://doi.org/10.1016/j.atherosclerosis.2017.12.008> PMID:[29253643](https://pubmed.ncbi.nlm.nih.gov/29253643/)
21. Rodriguez-Luna D, Rubiera M, Ribo M, Coscojuela P, Pagola J, Piñeiro S, Ibarra B, Meler P, Maisterra O, Romero F, Alvarez-Sabin J, Molina CA. Serum low-density lipoprotein cholesterol level predicts hematoma growth and clinical outcome after acute intracerebral hemorrhage. *Stroke*. 2011; 42:2447–52. <https://doi.org/10.1161/STROKEAHA.110.609461> PMID:[21799167](https://pubmed.ncbi.nlm.nih.gov/21799167/)
22. Martini SR, Flaherty ML, Brown WM, Haverbusch M, Comeau ME, Sauerbeck LR, Kissela BM, Deka R, Kleindorfer DO, Moomaw CJ, Broderick JP, Langefeld CD, Woo D. Risk factors for intracerebral hemorrhage differ according to hemorrhage location. *Neurology*. 2012; 79:2275–82. <https://doi.org/10.1212/WNL.0b013e318276896f> PMID:[23175721](https://pubmed.ncbi.nlm.nih.gov/23175721/)
23. O'Donnell MJ, Xavier D, Liu L, Zhang H, Chin SL, Rao-Melacini P, Rangarajan S, Islam S, Pais P, McQueen MJ, Mondo C, Damasceno A, Lopez-Jaramillo P, et al, and INTERSTROKE investigators. Risk factors for ischaemic and intracerebral haemorrhagic stroke in 22 countries (the INTERSTROKE study): a case-control study. *Lancet*. 2010; 376:112–23. [https://doi.org/10.1016/S0140-6736\(10\)60834-3](https://doi.org/10.1016/S0140-6736(10)60834-3) PMID:[20561675](https://pubmed.ncbi.nlm.nih.gov/20561675/)
24. Zia E, Pessah-Rasmussen H, Khan FA, Norrving B, Janzon L, Berglund G, Engstrom G. Risk factors for primary intracerebral hemorrhage: a population-based nested case-control study. *Cerebrovasc Dis*. 2006; 21:18–25. <https://doi.org/10.1159/000089589> PMID:[16286730](https://pubmed.ncbi.nlm.nih.gov/16286730/)
25. Wieberdink RG, Poels MM, Vernooij MW, Koudstaal PJ, Hofman A, van der Lugt A, Breteler MM, Ikram MA. Serum lipid levels and the risk of intracerebral hemorrhage: the Rotterdam Study. *Arterioscler Thromb Vasc Biol*. 2011; 31:2982–89. <https://doi.org/10.1161/ATVBAHA.111.234948> PMID:[21921260](https://pubmed.ncbi.nlm.nih.gov/21921260/)
26. Stein M, Luecke M, Preuss M, Boeker DK, Joedicke A, Oertel MF. Spontaneous intracerebral hemorrhage with ventricular extension and the grading of obstructive hydrocephalus: the prediction of outcome of a special life-threatening entity. *Neurosurgery*. 2010; 67:1243–51. <https://doi.org/10.1227/NEU.0b013e3181ef25de> PMID:[20948399](https://pubmed.ncbi.nlm.nih.gov/20948399/)
27. Hwang BY, Bruce SS, Appelboom G, Piazza MA, Carpenter AM, Gigante PR, Kellner CP, Ducruet AF, Kellner MA, Deb-Sen R, Vaughan KA, Meyers PM, Connolly ES Jr. Evaluation of intraventricular hemorrhage assessment methods for predicting outcome following intracerebral hemorrhage. *J Neurosurg*. 2012; 116:185–92. <https://doi.org/10.3171/2011.9.JNS10850> PMID:[21999319](https://pubmed.ncbi.nlm.nih.gov/21999319/)
28. Herrick DB, Ullman N, Nekoovaght-Tak S, Hanley DF, Awad I, LeDroux S, Thompson CB, Ziai WC. Determinants of external ventricular drain placement and associated outcomes in patients with spontaneous intraventricular hemorrhage. *Neurocrit Care*. 2014; 21:426–34. <https://doi.org/10.1007/s12028-014-9959-x> PMID:[24522761](https://pubmed.ncbi.nlm.nih.gov/24522761/)
29. Gaberel T, Magheru C, Emery E. Management of non-traumatic intraventricular hemorrhage. *Neurosurg Rev*. 2012; 35:485–94. <https://doi.org/10.1007/s10143-012-0399-9> PMID:[22732889](https://pubmed.ncbi.nlm.nih.gov/22732889/)
30. Rabinstein AA. Intracerebral haemorrhage: no good treatment but treatment helps. *Lancet*. 2017; 389:575–76. [https://doi.org/10.1016/S0140-6736\(17\)30002-8](https://doi.org/10.1016/S0140-6736(17)30002-8) PMID:[28081951](https://pubmed.ncbi.nlm.nih.gov/28081951/)
31. Abdelmalik PA, Ziai WC. Spontaneous Intraventricular Hemorrhage: When Should Intraventricular tPA Be

- Considered? *Semin Respir Crit Care Med.* 2017; 38:745–59.
<https://doi.org/10.1055/s-0037-1607991>
 PMID:[29262432](https://pubmed.ncbi.nlm.nih.gov/29262432/)
32. Steiner T, Diringer MN, Schneider D, Mayer SA, Begtrup K, Broderick J, Skolnick BE, Davis SM. Dynamics of intraventricular hemorrhage in patients with spontaneous intracerebral hemorrhage: risk factors, clinical impact, and effect of hemostatic therapy with recombinant activated factor VII. *Neurosurgery.* 2006; 59:767–73.
<https://doi.org/10.1227/01.NEU.0000232837.34992.32> PMID:[17038942](https://pubmed.ncbi.nlm.nih.gov/17038942/)
 33. Nguyen HS, Li L, Patel M, Kurpad S, Mueller W. Radiodensity of intraventricular hemorrhage associated with aneurysmal subarachnoid hemorrhage may be a negative predictor of outcome. *J Neurosurg.* 2018; 128:1032–36.
<https://doi.org/10.3171/2016.11.JNS152839>
 PMID:[28474990](https://pubmed.ncbi.nlm.nih.gov/28474990/)
 34. Chen J, Zhang D, Li Z, Dong Y, Han K, Wang J, Hou L. Lateral Ventricular Volume Asymmetry Predicts Poor Outcome After Spontaneous Intracerebral Hemorrhage. *World Neurosurg.* 2018; 110:e958–64.
<https://doi.org/10.1016/j.wneu.2017.11.149>
 PMID:[29203311](https://pubmed.ncbi.nlm.nih.gov/29203311/)
 35. Roh D, Boehme A, Young C, Roth W, Gutierrez J, Flaherty M, Rosand J, Testai F, Woo D, Elkind MS. Hematoma expansion is more frequent in deep than lobar intracerebral hemorrhage. *Neurology.* 2020; 95:e3386–93.
<https://doi.org/10.1212/WNL.0000000000010990>
 PMID:[33219144](https://pubmed.ncbi.nlm.nih.gov/33219144/)
 36. Witsch J, Bruce E, Meyers E, Velazquez A, Schmidt JM, Suwatcharangkoon S, Agarwal S, Park S, Falo MC, Connolly ES, Claassen J. Intraventricular hemorrhage expansion in patients with spontaneous intracerebral hemorrhage. *Neurology.* 2015; 84:989–94.
<https://doi.org/10.1212/WNL.0000000000001344>
 PMID:[25663233](https://pubmed.ncbi.nlm.nih.gov/25663233/)
 37. Deng L, Zhang YD, Ji JW, Yang WS, Wei X, Shen YQ, Li R, Zhang SQ, Lv XN, Li XH, Tang ZP, Wu GF, Zhao LB, et al. Hematoma Ventricle Distance on Computed Tomography Predicts Poor Outcome in Intracerebral Hemorrhage. *Front Neurosci.* 2020; 14:589050.
<https://doi.org/10.3389/fnins.2020.589050>
 PMID:[33328859](https://pubmed.ncbi.nlm.nih.gov/33328859/)
 38. Meyer M, Ronald J, Vernuccio F, Nelson RC, Ramirez-Giraldo JC, Solomon J, Patel BN, Samei E, Marin D. Reproducibility of CT Radiomic Features within the Same Patient: Influence of Radiation Dose and CT Reconstruction Settings. *Radiology.* 2019; 293:583–91.
<https://doi.org/10.1148/radiol.2019190928>
 PMID:[31573400](https://pubmed.ncbi.nlm.nih.gov/31573400/)
 39. Choe J, Lee SM, Do KH, Lee G, Lee JG, Lee SM, Seo JB. Deep Learning-based Image Conversion of CT Reconstruction Kernels Improves Radiomics Reproducibility for Pulmonary Nodules or Masses. *Radiology.* 2019; 292:365–73.
<https://doi.org/10.1148/radiol.2019181960>
 PMID:[31210613](https://pubmed.ncbi.nlm.nih.gov/31210613/)
 40. Sauerbrei W, Royston P, Binder H. Selection of important variables and determination of functional form for continuous predictors in multivariable model building. *Stat Med.* 2007; 26:5512–28.
<https://doi.org/10.1002/sim.3148>
 PMID:[18058845](https://pubmed.ncbi.nlm.nih.gov/18058845/)
 41. Akinwande MO, Dikko HG, Samson A. Variance Inflation Factor: As a Condition for the Inclusion of Suppressor Variable(s) in Regression Analysis. *Open Journal of Statistics.* 2015; 05:754–67.
<https://doi.org/10.4236/ojs.2015.57075>
 42. Orrù G, Pettersson-Yeo W, Marquand AF, Sartori G, Mechelli A. Using Support Vector Machine to identify imaging biomarkers of neurological and psychiatric disease: a critical review. *Neurosci Biobehav Rev.* 2012; 36:1140–52.
<https://doi.org/10.1016/j.neubiorev.2012.01.004>
 PMID:[22305994](https://pubmed.ncbi.nlm.nih.gov/22305994/)
 43. Kohavi R. A study of cross-validation and bootstrap for accuracy estimation and model selection. *Proceedings of the 14th international joint conference on Artificial intelligence - Volume 2.* (Montreal, Quebec, Canada: Morgan Kaufmann Publishers Inc.), pp. 1995; 1137–43.
 44. Vickers AJ, Elkin EB. Decision curve analysis: a novel method for evaluating prediction models. *Med Decis Making.* 2006; 26:565–74.
<https://doi.org/10.1177/0272989X06295361>
 PMID:[17099194](https://pubmed.ncbi.nlm.nih.gov/17099194/)
 45. Kerr KF, Brown MD, Zhu K, Janes H. Assessing the Clinical Impact of Risk Prediction Models With Decision Curves: Guidance for Correct Interpretation and Appropriate Use. *J Clin Oncol.* 2016; 34:2534–40.
<https://doi.org/10.1200/JCO.2015.65.5654>
 PMID:[27247223](https://pubmed.ncbi.nlm.nih.gov/27247223/)

SUPPLEMENTARY MATERIALS

Supplementary Materials 1

Z-score normalization

z-score normalization to make the image intensities have the properties of a standard normal distribution by scaling values to a mean of 0 and a standard deviation of 1 using the following formula:

$$z = \frac{\chi - \mu}{\sigma}$$

where μ was the mean value of the images, and σ was the standard deviation of images.

Supplementary Materials 2

Support vector machine model construction

We introduced five features, including history of hypercholesterolemia, baseline Graeb score, time to initial CT, INR and Rad-score, to the SVM model. Radial basis function (RBF) kernel function were

applied and the optimal parameters of SVM were selected by grid-search method. Parameter C was $3.58e+01$ and parameter gamma was $6.87e-04$. The 5-fold cross-validation was employed to validate the model performance in training cohort. The model was further validated in an independent testing cohort.

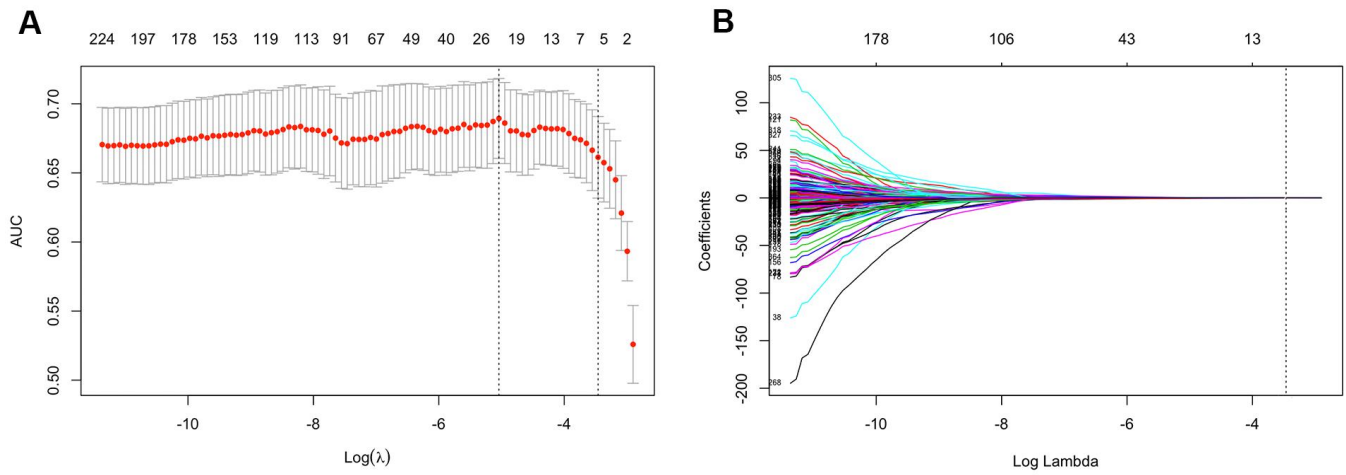
Supplementary Materials 3

Radiomics signature calculation formula

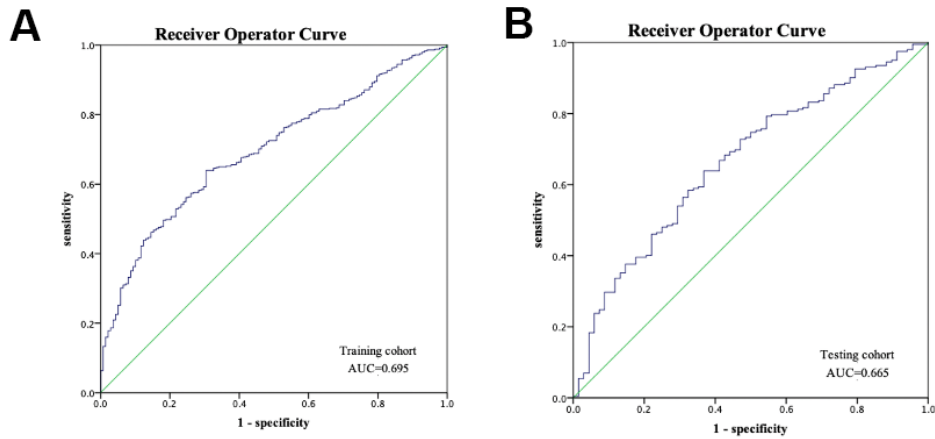
Rad-score = -
2.1378+0.135×Correlation_AllDirection_offset1_SD
-0.1723×Correlation_angle0_offset7
+0.5072×Correlation_angle0_offset4
-0.6258×HaralickCorrelation_AllDirection_offset4_SD
-0.1004×HaralickCorrelation_angle90_offset1
-0.353×ShortRunEmphasis_AllDirection_offset7_SD
+ 0.2577×ZonePercentage

Note: “SD” indicate the value reflects the standard deviation among the different directions.

Supplementary Figures



Supplementary Figure 1. Radiomics feature selection using the least absolute shrinkage and selection operator (LASSO) regression model. (A) Using 10-fold cross-validation to select tune parameter (λ). 7 features with non-zero coefficients were selected. Dotted lines on the left and right represent the minimum criterion and 1-standard error criterion (1-SE), respectively. The 1-SE criterion was applied in our study. A λ value of 0.03, with $\log(\lambda)$ of -3.46 was chosen (1-SE criteria). (B) LASSO coefficient profiles of the 396 radiomics features. The vertical line shows the optimal value of λ and 7 features with non-zero coefficients.



Supplementary Figure 2. Receiver operator curves (ROC) of Rad-score to predict poor outcome (Glasgow Outcome Scale, ≤ 3) in training cohort (A) and testing cohort (B).

Supplementary Tables

Supplementary Table 1. Radiomics features extracted from regions of interest.

Group	Number	Feature
Histogram Parameters	42	Histogram parameters are concerned with properties of individual pixels. They describe the distribution of voxel intensities within the CT image through commonly used and basic metrics. Let X denote the three-dimensional image matrix with N voxels and P the first order histogram divided by $N!$ discrete intensity levels.
Texture Parameters	54	Texture is one of the important characteristics used in identifying objects or regions of interest in an image, texture represents the appearance of the surface and how its elements are distributed. It is considered an important concept in machine vision, in a sense it assists in predicting the feeling of the surface (e.g. smoothness, coarseness ...etc.) from image. Various texture analysis approaches tend to represent views of the examined textures from different perspectives.
Form Factor Parameters	9	These group of features includes descriptors of the three-dimensional size and shape of the tumor region.
GLCM Parameters	100	The Grey level co-occurrence matrix (GLCM) $P(\mathbf{l}, \mathbf{j} \Theta, \mathbf{d})$ represents the joint probability of certain sets of pixels having certain grey-level values. It calculates how many times a pixel with grey-level i occurs jointly with another pixel having a grey value j . By varying the displacement vector \mathbf{d} between each pair of pixels. The rotation angle of an offset: $0^\circ, 45^\circ, 90^\circ, 135^\circ$ and displacement vectors (distance to the neighbor pixel: 1, 2, 3 ...), different co-occurrence distributions from the same image of reference. GLCM of an image is computed using displacement vector \mathbf{d} defined by its radius, (distance or count to the next adjacent neighbor preferably is equal to one) and rotational angles.
RLM Parameters	180	The grey level run-length matrix (RLM) $Pr(i, j \Theta)$ is defined as the numbers of runs with pixels of gray level i and run length j for a given direction θ . RLMs is generated for each sample image segment having directions ($0^\circ, 45^\circ, 90^\circ$ and 135°), then the following ten statistical features were derived: short run emphasis, long run emphasis, grey level non- uniformity, run length non-uniformity, Low Grey Level Run Emphasis, High Grey Level Run Emphasis, Short Run Low Grey Level Emphasis, Short Run High Grey Level Emphasis, Long Run Low Grey Level Emphasis and Long Run High Grey Level Emphasis.
GLZSM Parameters	11	The gray level Size Zone Matrix (SZM) is the starting point of Thibault matrices. For a texture image f with N gray levels, it is denoted $GSf(s, g)$ and provides a statistical representation by the estimation of a bivariate conditional probability density function of the image distribution values. It is calculated according to the pioneering Run Length Matrix principle: the value of the matrix $GSf(s, g)$ is equal to the number of zones of size s and of gray level g . The resulting matrix has a fixed number of lines equal to N , the number of gray levels, and a dynamic number of columns, determined by the size of the largest zone as well as the size quantization. This matrix is particularly efficient to characterize the texture homogeneity, non periodicity or speckle like texture; it had provided better characterizations than granulometry (or COM, RLM, etc.) for the classification of cell nuclei, dermis, road quality (bitumen condition) and some textures in PET images.

Supplementary Table 2. Multivariable analysis of features associated with IVH growth.

Variable	Odds ratio	95%CI	P value
Sex	-	-	0.116
Glasgow Coma Scale	-	-	0.131
Hypertension	-	-	0.076
Baseline IVH score	-	-	0.423
Baseline ICH volume	-	-	0.482
Blood glucose	-	-	0.987
PLT count	-	-	0.420
Hypercholesterolemia	0.12	0.02-0.90	0.039
INR	4.27	1.40-13.0	0.011
Baseline Graeb score	1.26	1.16-1.36	<0.001
Time to initial CT	0.70	0.58-0.86	<0.001
Rad-score	2.3	1.6-3.3	<0.001

Abbreviations: ICH=intracerebral hemorrhage; IVH=intraventricular hemorrhage; INR= international normalized ratio.

Supplementary Table 3. Details about the selected features.

Feature name	ICC	Group
Correlation_AllDirection_offset1_SD	0.91	Texture Parameters
Correlation_angle0_offset7	0.89	Texture Parameters
Correlation_angle0_offset4	0.90	Texture Parameters
HaralickCorrelation_AllDirection_offset4_SD	0.81	GLCM
HaralickCorrelation_angle90_offset1	0.90	GLCM
ShortRunEmphasis_AllDirection_offset7_SD	0.80	GLRLM
Zone Percentage	0.98	GLZSM

Abbreviations: GLCM= Grey level co-occurrence matrix, GLRLM= Grey level run-length matrix, GLZSM= Gray level size zone matrix, ICC= Inter-class correlation coefficient.

Supplementary Table 4. Confusion matrices of training cohort.

Training cohort (N=626)		Actual	
		Positive	Negative
Predicted	Positive	70	186
	Negative	14	356

Supplementary Table 5. Confusion matrices of testing cohort.

Testing cohort (N=270)		Actual	
		Positive	Negative
Predicted	Positive	30	85
	Negative	7	148