### Neurofilament light chain plasma concentration predicts neurodegeneration and clinical progression in nondemented elderly adults

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#### ABSTRACT

Previous studies demonstrated that plasma neurofilament light chain (NFL) played important predictive roles in disease progression and neurodegeneration in the preclinical phase of familial Alzheimer's disease (AD). However, whether plasma NFL has the same predictive roles in sporadic AD is still unclear. In this study, 243 cognitively normal (CN) participants from the ADNI database were divided into two subgroups (CN- and CN+) according to CSF A $\beta$  or AV45-PET. Associations of baseline plasma NFL concentrations or rate of change in plasma NFL with longitudinal data on other biomarkers were tested by multivariate linear mixed effects models (LMEMs). Results showed that plasma NFL concentration and its rate of change were already abnormally high in the preclinical phase of AD. Plasma NFL was associated with three core AD-related biomarkers in preclinical phase. Baseline plasma NFL, but not its rate of change, played predictive roles in both cognitive decline ( $\beta$  = -0.0349, p = 0.0274) and hippocampal atrophy ( $\beta$  = -0.0351, p = 0.0088), especially for preclinical AD participants. In summary, these results indicated that baseline plasma NFL, but not its rate of change, may be a valuable noninvasive tool to assess neurodegeneration and predict longitudinal disease progression in preclinical AD individuals.

#### **INTRODUCTION**

Available evidence strongly supports the notion that the initiating event in Alzheimer's disease (AD) is related to abnormality of  $\beta$ -amyloid (A $\beta$ ) peptide, which occurs when individuals are still cognitively normal [1–5].

Based on this notion, current studies have divided the clinical stages of AD into three phases: preclinical

phase, prodromal phase and dementia phase [6]. Among these phases, the most critical stage for early diagnosis and monitoring is the preclinical phase in which individuals are cognitively normal but have had pathological changes. These pathological changes can be reflected by some biomarkers, such as cerebrospinal fluid A $\beta$  (CSF A $\beta$ ), cerebrospinal fluid total tau protein (CSF t-tau), cerebrospinal fluid phosphorylated tau protein (CSF p-tau), F18-fluorodeoxyglucose positron emission tomography (FDG-PET) and magnetic resonance imaging (MRI). However, none of these biomarkers is suited to track disease progression, because of their invasiveness or high expenses. Therefore, many studies focused on blood biomarkers, such as plasma neurofilament light (NFL).

The NFL is a component of the axonal cytoskeleton and a putative marker of large-caliber axonal degeneration which is an important pathological change in neurodegeneration diseases [7]. Advancements in measurements of NFL have revealed the abnormal increase in the progression of AD, even in early stages. As for familial AD, a recent study has found important predictive roles of serous NFL in disease progression and brain neurodegeneration at the preclinical phase [8]. However, whether blood NFL has the same predictive roles in sporadic AD is still unclear. Another recent study focusing on cognitively normal (CN) individuals has found that elevated levels of CSF NFL but not CSF tTau, pTau or neurogranin (Ng) were a risk factor for mild cognitive impairment (MCI) [9]. Moreover, advancements in measurements of NFL have revealed strong correlations between CSF NFL and blood NFL, which has sparked interests in roles of blood NFL on sporadic AD [10]. Therefore, we speculated that, as for sporadic AD, plasma NFL also had some predictive roles in neurodegeneration or progression of disease in the preclinical phase. This study focused on CN individuals to test the hypothesis that the plasma NFL concentration and its rate of change are abnormally elevated in the preclinical phase of AD and they correlate with impaired cognition, neuroimaging abnormalities and CSF biomarkers of AD.

#### **RESULTS**

#### Sample characteristics

The demographics of the study population were listed in Table 1. No statistical difference was found between CNgroup (healthy control group: cognitively normal participants without significant AB-related pathological changes) and CN+ group (preclinical group: cognitively normal participants with significant Aβ-related pathological changes) in age (p = 0.0510), sex (p =0.0679) and education (p = 0.1005). The prevalence of an APOEE4 in CN+ group was higher than that in CNgroup. Plasma NFL correlated with age ( $\rho = 0.47$ , p < 0.01), but not with sex (median, 30.9 pg/ml for male vs 31.7 pg/ml for female, p = 0.44), education ( $\rho = 0.01$ , p =0.87) or APOEs4 genotype (median, 30.6 pg/ml for carriers vs 31.8 pg/ml for noncarriers, p = 0.60). As for CSF biomarkers, the levels of Aβ42, tTau and p-tau181 were much higher in CN+ group (A $\beta$ 42: p < 0.0001; t-tau: p = 0.0002; p-tau181: p < 0.0001) compared with the CN- group. However, the cognitive level (ADNI\_MEM: p = 0.1052), FDG-PET (p = 0.1525) and hippocampus volume (p = 0.0734) showed no significant differences between the two groups. It is worth noting that both plasma NFL concentration and its rate of change were higher in the CN+ group (NFL concentration: p = 0.0044; Rate of change: p < 0.0001) (Table 1, Figure 1). However, the two groups showed a high degree of overlap in the range of plasma NFL concentration as well as that of its change rate (Table 1, Figure 1).

#### Cross-sectional associations of plasma NFL with CSF biomarkers, cognition and hippocampus volume

Table 2 showed the results of a multiple linear regression analysis for cross-sectional associations. Among total CN participants, a significant association was found between CSF A $\beta$ 42 and plasma NFL concentration ( $\beta$  = -0.1421, p = 0.0320), which was not found in the two subgroups. Moreover, significant associations between CSF t-tau and plasma NFL concentration were found in both total CN participants ( $\beta$  = 0.1821, p = 0.0091) and CN+ subgroup ( $\beta$  = 0.29319, p = 0.0042). There was no significant association of plasma NFL concentration with CSF p-tau181, FDG-PET, ADNI\_MEM, ADNI\_EF, and hippocampus volume.

## Associations between rate of change in plasma NFL and rates of change in some other biomarkers

Associations between estimated annual rate of change in plasma NFL and estimated annual rates of change in some other biomarkers for CN- participants and CN+ participants were shown in Table 3. Results from multivariate linear mixed effects models (LMEMs) revealed significant associations between rate of change in CSF A $\beta$ 42 and rate of change in plasma NFL in CN+ participants ( $\beta$  = -3.2697, p = 0.0027) and total CN participants ( $\beta$  = -1.4466, p = 0.2364). Although results also showed the associations between rate of change in plasma NFL and rates of change in FDG-PET ( $\beta$  = -0.0047, p = 0.0451) and ADNI\_MEM ( $\beta$  = -0.0177, p = 0.0482) in total CN participants, these associations were not found in the two subgroups.

#### Prediction of changes in hippocampus volume and cognition by baseline plasma NFL and rate of change in plasma NFL

In CN+ group, a higher baseline plasma NFL level was significantly associated with an increased rate of change in both ADNI\_MEM ( $\beta = -0.0349$ , p = 0.0274) and hippocampus volume ( $\beta = -0.0351$ , p = 0.0088), but not

Table 1. Demographics for CN population.

Variable	CN-	CN+	Total	p value
N	130	113	243	-
Age, Mean (SD), years	72.77 (5.54)	73.33 (6.21)	72.76 (6.80)	0.0510
Female, N (%)	55 (42.31)	62 (54.87)	117 (48.15)	0.0679
Education, Mean (SD), years	16.85 (2.64)	16.35 (2.54)	16.62 (2.60)	0.1005
ΑΡΟΕε4, %	13.85	43.36	27.57	<0.0001ª
ADNI_MEM, Mean (SD)	1.13 (0.58)	1.0 (0.59)	1.07 (0.59)	0.1052
ADNI_EF, Mean (SD)	0.95 (0.79)	0.65 (0.80)	0.81 (0.81)	0.0031ª
CSF Aβ42, Mean (SD), pg/ml	237.98 (25.62)	154.732 (32.47)	199.27 (50.68)	<0.0001ª
CSF t-tau, Mean (SD), pg/ml	57.06 (22.67)	77.47 (39.93)	66.55 (33.41)	0.0002ª
CSF p-tau181, Mean (SD), pg/ml	26.95 (10.90)	41.65 (24.66)	33.79 (19.97)	<0.0001ª
Hippocampus Volume, Mean (SD), mm <sup>3</sup>	7618.49 (741.95)	7440.13 (821.38)	7535.55 (783.34)	0.0734
FDG-PET	1.32. (0.11)	1.30 (0.12)	1.32 (0.11)	0.1525
Plasma NFL, Mean (SD), pg/ml	31.61 (13.10)	36.39 (14.46)	33.84 (13.93)	$0.0044^{a}$
Rate of change Plasma NFL, year	2.34	2.94	-	$< 0.0001^{a}$
Mean (CI), pg/ml	$(1.47 \sim 3.25)$	$(1.83 \sim 4.04)$	-	-

Abbreviations: APOE, apolipoprotein E; ADNI, Alzheimer's Disease Neuroimaging Initiative; ADNI\_MEM, memory domain composite score; ADNI\_EF, executive domain composite score; CSF, cerebrospinal fluid; Aβ, amyloid-β; t-tau, total tau protein; p-tau, phosphorylated tau protein; NFL, neurofilament light chain; SD, standard deviation; CI, confidence interval; CN-, cognitively normal participants with negative Aβ; CN+, cognitively normal participants with positive Aβ <sup>a</sup> Lower-case letter indicated that the results were statistically significant.





Variable	CN-		CN+		All participants	
	β	p value	β	p value	β	p value
CSF Aβ42	-0.0215	0.827	0.02457	0.8109	-0.1421	0.0320a
CSF t-tau	-0.0325	0.741	0.29319	0.0042a	0.1821	0.0091a
CSF p-tau181	-0.0353	0.717	0.095684	0.3604	0.0887	0.2011
FDG-PET	-0.0454	0.6365	-0.09396	0.3821	-0.0811	0.2583
ADNI_MEM	0.0103	0.8375	0.024174	0.6714	0.0098	0.7946
ADNI_EF	0.0634	0.3728	-0.0243	0.7572	-0.0047	0.9311
Hippocampus Volume	-0.0797	0.3178	-0.0336	0.7360	-0.0752	0.2290

Table 2. Cross-sectional associations of plasma NFL with CSF biomarkers, cognition and hippocampus volume.

Abbreviations: ADNI, Alzheimer's Disease Neuroimaging Initiative; ADNI\_MEM, memory domain composite score; ADNI\_EF, executive domain composite score; CSF, cerebrospinal fluid; Aβ, amyloid-β; t-tau, total tau protein; p-tau, phosphorylated tau protein; FDG-PET, F18-fluorodeoxyglucose positron emission tomography; CN-, cognitively normal participants with negative Aβ; CN+, cognitively normal participants with positive Aβ

<sup>a</sup> Lower-case letter indicated that the results were statistically significant.

Cross-sectional associations between plasma NFL concentrations and other biomarkers at baseline were tested by multiple linear regression models corrected for age, sex, education and APOE genotype.

Variable	CN-		CN+		All participants	
	β	p value	β	p value	β	p value
CSF Aβ42	-1.4466	0.2364	-3.2697	$0.0027^{a}$	-2.1670	0.0059ª
CSF t-tau	0.9519	0.2780	1.1780	0.3870	1.2526	0.1100
CSF p-tau181	-0.5848	0.4607	1.7722	0.2374	0.7792	0.3500
FDG-PET	-0.0074	0.0501	-0.0016	0.6448	-0.0047	0.0451ª
ADNI_MEM	-0.0040	0.6923	-0.0317	0.0573	-0.0177	$0.0482^{a}$
ADNI_EF	-0.0157	0.2706	-0.0292	0.1804	-0.0067	0.5945
Hippocampus Volume	0.0037	0.6641	-0.0249	0.0862	-0.0089	0.2780

Table 3. Associations between rate of change in plasma NFL and rates of change in other biomarkers.

Abbreviations: ADNI, Alzheimer's Disease Neuroimaging Initiative; ADNI\_MEM, memory domain composite score; ADNI\_EF, executive domain composite score; CSF, cerebrospinal fluid; A $\beta$ , amyloid- $\beta$ ; t-tau, total tau protein; p-tau, phosphorylated tau protein; FDG-PET, F18-fluorodeoxyglucose positron emission tomography; CN-, cognitively normal participants with negative A $\beta$ ; CN+, cognitively normal participants with positive A $\beta$ 

<sup>a</sup> Lower-case letter indicated that the results were statistically significant.

The rate rate of change in plasma NFL was was extracted from LMEMs. Associations between rate of change in plasma NFL and longitudinal data of other biomarkers (CSF Aβ42, CSF t-tau, CSF p-tau181, FDG-PET, ADNI\_MEM, ADNI\_EF, Hippocampus Volume) were tested by LMEMs corrected for age, sex, education and APOE genotype.

in ADNI\_EF ( $\beta$  = -0.0135, p = 0.5070) (Table 4, Figure 2). Moreover, higher CSF A $\beta$ 42 and p-tau181 levels were significantly associated with an increased rate of change in ADNI\_MEM ( $\beta$  = -0.0429, p = 0.0120) and a higher CSF t-tau level was significantly associated with an increased rate of change in hippocampus volume ( $\beta$  = -0.0372, p = 0.0061) (Table 4). In CN- group, we did not find any association between these biomarkers (plasma NFL, CSF A $\beta$ 42, CSF t-tau, CSF p-tau181 and FDG-PET) and rate of change in cognition or hippocampus volume.

#### **DISCUSSION**

Our study is the first to focus on cognitively normal individuals to explore the value of plasma NFL in the early prediction of sporadic AD. The main findings of our study were that (1) plasma NFL concentration and rate of change in plasma NFL were abnormally high in the preclinical phase of AD (2) plasma NFL was associated with some core AD-related biomarkers in the preclinical phase of AD (3) baseline plasma NFL played predictive roles in both impaired cognition and

	Variable	CI	Ň-	CN+		
	variable -	β	p value	β	p value	
ADNI_MEM	Plasma NFL	-0.0009	0.9313	-0.0349	0.0274ª	
	Rate of change (NFL)	-0.0040	0.6923	-0.0317	0.0573	
	CSF Αβ42	0.0162	0.1216	0.0387	0.0113 <sup>a</sup>	
	CSF t-tau	-0.0130	0.1818	-0.0305	0.0513	
	CSF p-tau181	0.0111	0.2698	-0.0429	0.0120 <sup>a</sup>	
	FDG-PET	0.0112	0.2599	0.0225	0.1679	
ADNI_EF	Plasma NFL	-0.0076	0.5975	-0.0135	0.5070	
	Rate of change (NFL)	-0.0157	0.2706	-0.0292	0.1804	
	CSF Αβ42	0.0116	0.4493	0.0021	0.9140	
	CSF t-tau	0.0179	0.2305	-0.0322	0.1080	
	CSF p-tau181	-0.0093	0.5289	-0.0231	0.2810	
	FDG-PET	0.0031	0.8334	0.0171	0.4095	
Hippocampus	Plasma NFL	-0.0007	0.9311	-0.0351	$0.0088^{a}$	
Volume	Rate of change (NFL)	0.0037	0.66405	-0.0249	0.0862	
	CSF Αβ42	0.0001	0.9950	0.0209	0.1360	
	CSF t-tau	-0.0040	0.6370	-0.0372	0.0061ª	
	CSF p-tau181	0.0084	0.3380	-0.0196	0.1713	
	FDG-PET	0.01008	0.25348	0.01605	0.2432	

Table 4. Prediction of changes in hippocampus volume and cognition by biomarkers

Abbreviations: ADNI, Alzheimer's Disease Neuroimaging Initiative; ADNI\_MEM, memory domain composite score; ADNI\_EF, executive domain composite score; NFL, neurofilament light chain; CSF, cerebrospinal fluid; Aβ, amyloid-β; t-tau, total tau protein; p-tau, phosphorylated tau protein; FDG-PET, F18-fluorodeoxyglucose positron emission tomography; CN-, cognitively normal participants with negative Aβ; CN+, cognitively normal participants with positive Aβ

<sup>a</sup> Lower-case letter indicated that the results were statistically significant.

Associations between baseline biomarkers (plasma NFL concentrations, rate of change in plasma NFL, CSF Aβ42, CSF t-tau, CSF p-tau181 and FDG-PET) and longitudinal data (ADNI\_MEM, ADNI\_EF and hippocampus volume) were tested by LMEMs corrected for age, sex, education and APOE genotype.

neuroimaging abnormalities, especially for the preclinical AD individuals (4) rate of change in plasma NFL showed predictive effects on neither of impaired cognition and neuroimaging abnormalities among CN individuals.

The higher level of baseline plasma NFL concentrations in CN+ group indicated that plasma NFL concentrations have already been abnormally high in the preclinical phase of AD. This result contradicted a previous study which did not find statistical differences in plasma NFL concentration between CN+ and CN- groups [10]. It is worth noting that our study included a larger cognitively normal cohort with more stringent quality control and excluded extremes (limit: mean±3SD). Moreover, to avoid misclassifying some preclinical AD patients in

the category of the normal group, we divided participants into A $\beta$  positive or negative (A+/A-) patients according CSF AB or AV45-PET SUVR. Therefore, data quality in our study was higher than that in the previous study. Three CSF biomarkers (Aβ42, ttau, p-tau181) all significantly increased in CN+ group, whereas baseline cognitive level (especially ADNI MEM), hippocampus volume and FDG-PET showed no obvious differences compared with the CNgroup. These results were consistent with previous assumptions. A widely accepted assumption was that biomarkers have already been abnormal in the preclinical phase without significant cognitive decline. Among these biomarkers, some core biomarkers (especially  $A\beta$  and tau protein) became abnormal earlier than other markers (such as MRI and FDG-PET) [11].

However, though our results showed that plasma NFL concentration was already abnormally high in the preclinical AD phase, we could see that range of plasma NFL concentration in the two groups had considerable overlap, indicating that, unlike  $A\beta$ , plasma NFL may not be suitable as an early diagnostic marker.

Consistent with previous studies, our study found a cross-sectional association between plasma NFL and CSF t-tau. Previous studies showed that both CSF tau and plasma NFL, as indicators of neuronal injury, may reflect the degree of neurodegenerative changes [12–14]. It is worth noting that results also showed an association between plasma NFL and CSF Aβ42. An association between plasma NFL and CSF AB42 was found in total participants by the cross-sectional association test. Moreover, longitudinal results also showed associations between estimated annual rate of change in plasma NFL and estimated annual rate of change in CSF Aβ42 for total participants and CN+ participants, but not for CN- participants. These results contradicted previous studies which did not find associations between plasma NFL and CSF Aβ42 [10]. We thought the main reason for these contradictory results was that our study focused on the cognitively normal population. Previous studies indicated that Aβplaque accumulation in individuals destined to become dementia might begin two decades before the clinical symptoms [15]. Thus in the CN phase, not all individuals' CSF Aβ42 had reached the plateau stage and the changes of CSF A $\beta$ 42 were still reflected by plasma NFL. According to the above results, we could see that plasma NFL was already abnormally high in the preclinical phase of AD and was associated with some early core AD-related biomarkers (CSF A $\beta$ 42 and tau protein).

In order to explore the utility of plasma NFL as a predictor of subsequent neurodegeneration and clinical symptoms and to compare predictive roles of plasma NFL with those of some other biomarkers, we used a (retrospective) pseudo-predictive analysis. The results showed that baseline plasma NFL could predict future annualized changes of hippocampal atrophy and annualized changes of ADNI MEM especially for CN+ participants. Compared with plasma NFL, CSF ptau181 played predictive roles only in cognitive decline for CN+ participants and CSF t-tau play predictive roles only in hippocampal atrophy for CN+ participants. According to previous studies, earliest cognitive symptoms in AD were characterized by typical deficits in episodic memory (ADNI-MEM) rather than executive functioning (ADNI-EF) [11]. Previous autopsy data confirmed that cerebral atrophy was one of the neurodegenerative biomarkers which indicated dendritic pruning as well as loss of synapses and neurons [16]. Some studies implied that cerebral atrophy (especially hippocampal volume) was the most proximate pathological substrate of cognitive impairment in AD [17-19]. Overall, these results



**Figure 2. Prediction of changes in hippocampus volume and cognition by baseline plasma NFL.** Associations between baseline plasma NFL concentrations or rate of change in plasma NFL and longitudinal data were tested by LMEMs corrected for age, sex, education and APOE genotype. (A) In CN+ group, higher baseline plasma NFL levels were significantly associated with an increased rate of change in hippocampus volume ( $\beta$  = -0.0351, p = 0.0088). This association did not exist in CN- group ( $\beta$  = -0.0007, p = 0.9311). (**B**) Results did not show association between baseline plasma NFL levels and ADNI\_EF (CN-:  $\beta$  = -0.0076, p = 0.5975; CN+:  $\beta$  = -0.0135, p = 0.5070). (**C**) In CN+ group, higher baseline plasma NFL levels were significantly associated with an increased rate of change in ADNI\_MEM ( $\beta$  = -0.0349, p = 0.0274). This association did not exist in CN- group ( $\beta$  = -0.0039, p = 0.9313).

indicated that, even in the preclinical phase of AD, plasma NFL still played predictive roles not only in cerebral structural changes but also in cognitive impairment. Therefore, though plasma NFL may not be suitable as an early diagnostic marker, it may be suitable as an early neurodegenerative biomarker in preclinical AD.

Blood-based biomarkers have obvious advantages in monitoring disease progression. Several previous studies have tested some blood-based biomarkers. However, there was heterogeneity in the results. As for plasma tau protein, a study found associations between increased plasma tau protein and AD hallmarks, but these associations were mild and they varied with cohorts [20]. Though a comprehensive meta-analysis of blood markers showed total tau protein was useful in differentiating participants with established AD from healthy controls yet with some overlaps, no evidence showed that it was useful in detection of earlier disease [21]. As for plasma A $\beta$ , previous studies also failed to obtain consistent results to prove its ability to diagnose of AD [21]. Moreover, Aβ-related biomarkers may not track progression unless in very early stage of disease because A\beta-related pathology is an early event in AD and it may reach the plateau before symptom onset. Therefore, the above blood-based biomarkers seemed not to be suitable as early biomarkers in preclinical AD and our results may provide a new candidate.

A few limitations in our study should be noted. Firstly, we had some missing data in our longitudinal data, especially at later follow-up time points, which had a slight impact on the results. Secondly, the sample size in our study was comparatively small and these results should be further tested in a larger population. Thirdly, increased plasma NFL concentrations were also found in some other neurodegenerative diseases, such as frontotemporal dementia (FTD) and progressive supranuclear palsy (PSP), indicating a lack of specificity of plasma NFL [22, 23]. Though this limitation does not negate its role as a general biomarker for neurodegenerative diseases are warranted to investigate the roles of plasma NFL.

Taken together, our findings suggested that plasma NFL concentrations and rate of change in plasma NFL were already abnormally high in the preclinical AD phase. Plasma NFL was associated with some core AD-related biomarkers, cognitive decline and cerebral atrophy, especially for the preclinical AD individuals. Therefore, plasma NFL may be a valuable noninvasive tool to assess neurodegeneration and to predict preclinical AD individuals' longitudinal disease progression. However, the predictive roles of rate of change in plasma NFL were limited in the preclinical phase of sporadic AD.

#### **METHODS**

#### ADNI database

Data used in this study were obtained from the Alzheimer's Disease Neuroimaging Initiative (ADNI) database (http://adni.loni.usc.edu). The ADNI database was launched in 2003 as a public-private partnership, led by the principal investigator Michael W. Weiner, MD. The ADNI participants have been recruited from more than 50 sites across the United States and Canada. The primary objective of the ADNI has been to test whether serial MRI, PET, other biological markers, and clinical or neuropsychological assessment can be combined to measure the progression of MCI and early AD. ADNI database consists of three parts, including the ADNI 1, the ADNI Grand Opportunities (GO) and the ADNI 2. To date, these 3 protocols have recruited more than 1500 adults (age range, 55-90 years) to participate in the research, including CN older individuals, persons with early or late MCI, and patients with early AD. The follow-up duration for each study group was specified in the protocols for the ADNI 1, ADNI 2, and ADNI GO. Regional ethics committees of all institutions approved of the study. Written informed consent was obtained from all study participants.

#### Population

The study only included the CN participants. CN participants were defined as individuals with a Mini-Mental State Examination (MMSE) score from 25 to 30 inclusive, Clinical Dementia Rating (CDR) score of 0, no evidence of depression, and no memory complaints. In the ADNI database, the number of CN participants who have baseline plasma NFL samples is 482. We chose 245 participants who have available baseline CSF samples as well as neuroimaging and cognitive data. All the participants had at least one follow-up data of all biomarkers. The minimum follow-up time was one year and the maximum follow-up time was ten years. More than 75 percent of participants had at least three years of follow-up. Then we remove the extremums that were higher than three standard deviations (SD) above the mean or lower than three standard deviations below the mean (limited: mean±3SD). This step removed two extremes. Eventually, 243 CN participants were included in our study. According to the National Institute of Aging-Alzheimer's Association (NIA-AA) research framework, A\beta-related biomarkers (CSF Aβ and A $\beta$ -PET) were regarded as the core markers for the diagnosis of AD continuum [6]. In our study, AB positive or negative (A+/A-) participants were distinguished by a previously established cutoff of 192 pg/ml (CSF A $\beta$ ) or 1.11 (AV45-PET SUVR) [24]. Then CN participants were divided into two subgroups:

participants with A- (CN-) and participants with A+ (CN+). We regarded the CN+ participants as patients with preclinical AD and regarded CN- participants as controls. The final groups are presented in Table 1.

#### **Cognitive assessments**

Cognition was assessed by ADNI\_EF and ADNI\_MEM, which were two composite scales for executive functioning (ADNI\_EF) and memory (ADNI\_MEM) using data from the ADNI neuropsychological battery via item response theory methods. Compared with the separate scales, these two composite scales can reflect the cognitive level more robustly.

#### Measurement of CSF biomarkers

Aβ42, t-tau, and p-tau181 were measured by the ADNI Biomarker Core (University of Pennsylvania) using the multiplex xMAP Luminex platform (Luminex Corp, Austin, TX, USA) with the INNOBIA AlzBio3 kit (Fujirebio, Ghent, Belgium) as described previously [1].

#### Measurement of neuroimaging

MRI brain scans were acquired using 1.5-T imaging systems with a standardized protocol that included T1weighted images using a sagittal, volumetric, magnetization-prepared rapid acquisition with gradient echo sequence. Automated volume measures were FreeSurfer (http://surfer.nmr.mgh. obtained with harvard.edu/fswiki). Our study used averaged volume measurements for the right and left hippocampi. The cerebral brain metabolism was tested by mean FDG-PET counts of lateral and medial frontal regions, anterior and posterior cingulate regions, as well as lateral parietal and temporal regions. The ADNI Florbetapir summary data (AV45-PET) were calculated using the cortical summary regions of interest (ROI) divided by the whole cerebellum reference region (SUMMARYSUVR WHOLECEREBNORM).

#### Measurement of plasma NFL concentrations

Blood samples were collected, processed, aliquoted, and frozen at -80°C according to standardized procedures. Plasma NFL concentrations were measured on an ultrasensitive single-molecule array platform with a home brew kit (Simoa Homebrew Assay Development Kit; Quanterix Corporation) using the same methodology as described previously [25]. The assay used a combination of monoclonal antibodies and purified bovine NFL as a calibrator. The relative error of the blank calculated concentrations was below 20% for all calibrators, run in triplicate, resulting in lower limits of quantifications of 2.2 pg/ml and upper limits of quantification of 1620 pg/ml. All samples measured within the range spanned by the limits of quantifications, and for the low quality control samples with a concentration of 14 pg/ml, the intra-assay coefficient of variation was 11.0% and intermediate precision was 11.1%. For the high-concentration quality control samples with a concentration of 137 pg/ml, the intra-assay coefficient of variation was 8.8% and intermediate precision was 9.6%. The measurements were performed in September 2016.

#### Statistical analysis

The Mann-Whitney test and the Spearman rank correlation test were used to test the correlations between plasma NFL concentration and demographic factors including age, sex, apolipoprotein EE4 genotype (APOEɛ4), and education. The intergroup differences in demographic data and biomarkers were tested using the Wilcoxon test. The distribution of plasma NFL data was non-normal and therefore normal transformation was performed prior to subsequent analysis (using z-scale). The rate of change in plasma NFL was modeled using a multivariate linear mixed effects model (LMEM) with fixed effects of time from baseline, diagnostic status, and interaction between time from baseline and diagnostic status. The model included random slope and intercept terms for each participant with age, sex, education and APOE genotype as covariants. The rate of change in NFL for each individual was extracted from the model for subsequent analyses. Associations between plasma NFL concentration and other biomarkers at baseline were tested by multiple linear regression models corrected for age, sex, education and APOE genotype. Associations of baseline plasma NFL concentration or rate of change in plasma NFL with longitudinal data on other biomarkers were tested by LMEMs corrected for age, sex, education and APOE genotype. The Bonferroni correction was used for multiple comparisons. All tests used a significance level of P < 0.05. All statistical analyses were performed using the R programming language, version 3.5.1 (R Foundation) with LME analysis performed specifically using the nlme package, version 3.1.

#### Abbreviations

AD: Alzheimer disease; ADNI: Alzheimer's Disease Neuroimaging Initiative; APOE $\epsilon$ 4: apolipoprotein E $\epsilon$ 4 genotype; A $\beta$ :  $\beta$ -amyloid; CDR: Clinical Dementia Rating; CN: cognitively normal; CSF: cerebrospinal fluid; CSF p-tau: phosphorylated tau protein in cerebrospinal fluid; CSF t-tau: total of tau protein in cerebrospinal fluid; FDG-PET: F18-fluorodeoxyglucose positron emission tomography; FTD: frontotemporal dementia; LMEM: linear mixed-effects model; MCI: mild cognitive impairment; MRI: magnetic resonance imaging; MMSE: mini-mental state examination; NFL: Neurofilament light; Ng: neurogranin; NIA-AA: National Institute of Aging-Alzheimer's Association; PSP: progressive supranuclear palsy; ROI: regions of interest; SD: standard deviation.

#### **AUTHOR CONTRIBUTIONS**

JTY, HH conceptualized and designed the study. HH and KLC conducted, analyzed and extracted data. HH and JTY wrote the first draft of the manuscript. All authors reviewed the manuscript. All authors read and approved the final manuscript.

#### ACKNOWLEDGMENTS

Data used in preparation for this paper were obtained from the Alzheimer's Disease Neuroimaging Initiative (ADNI) database (<u>http://adni.loni.usc.edu/</u>). As such, the investigators within the ADNI contributed to the design and implementation of ADNI and/or provided data but did not participate in the analysis or in the writing of this paper. A complete listing of ADNI investigators can be found at: <u>http://adni.loni.usc.edu/wp-content/uploads/</u> how to apply/ADNI Acknowledgement List.pdf

#### **CONFLICTS OF INTEREST**

The authors declare that they have no competing interests.

#### FUNDING

This study was supported by grants from the National Key R&D Program of China (2018YFC1314700), National Natural Science Foundation of China (91849126). Shanghai Municipal Science and Technology Major Project (No.2018SHZDZX01) and ZHANGJIANG LAB, Tianqiao and Chrissy Chen Institute, and the State Key Laboratory of Neurobiology and Frontiers Center for Brain Science of Ministry of Education, Fudan University and Qingdao Applied Basic Research Project (18-2-2-43-jch). Data collection and sharing for this project was funded by the Alzheimer's Disease Neuroimaging Initiative (ADNI) (National Institutes of Health Grant U01 AG024904) and DOD ADNI (Department of Defense award number W81XWH-12-2-0012). ADNI is funded by the National Institute on Aging, the National Institute of Biomedical Imaging and Bioengineering, and through generous contributions from the following: AbbVie, Alzheimer's Association; Alzheimer's Drug Discovery Foundation; Araclon Biotech; BioClinica, Inc.; Biogen; Bristol-Myers Squibb Company; CereSpir, Inc.; Cogstate; Eisai Inc.; Elan Pharmaceuticals, Inc.; Eli Lilly and

Company; EuroImmun; F. Hoffmann-La Roche Ltd and its affiliated company Genentech, Inc.; Fujirebio; GE Ltd.; Healthcare; IXICO Janssen Alzheimer Immunotherapy Research & Development, LLC.; Johnson & Johnson Pharmaceutical Research & Development LLC.; Lumosity; Lundbeck; Merck & Co., Inc.: Meso Scale Diagnostics, LLC.: NeuroRx Research: Neurotrack Technologies; Novartis Pharmaceuticals Corporation; Pfizer Inc.; Piramal Imaging; Servier; Takeda Pharmaceutical Company; and Transition Therapeutics. The Canadian Institutes of Health Research is providing funds to support ADNI clinical sites in Canada. Private sector contributions are facilitated by the Foundation for the National Institutes of Health (www.fnih.org). The grantee organization is the Northern California Institute for Research and Education, and the study is coordinated by the Alzheimer's Therapeutic Research Institute at the University of Southern California. ADNI data are disseminated by the Laboratory for Neuro Imaging at the University of Southern California.

#### REFERENCES

 Shaw LM, Vanderstichele H, Knapik-Czajka M, Clark CM, Aisen PS, Petersen RC, Blennow K, Soares H, Simon A, Lewczuk P, Dean R, Siemers E, Potter W, et al, and Alzheimer's Disease Neuroimaging Initiative. Cerebrospinal fluid biomarker signature in Alzheimer's disease neuroimaging initiative subjects. Ann Neurol. 2009; 65:403–13.

https://doi.org/10.1002/ana.21610 PMID:19296504

 Mintun MA, Larossa GN, Sheline YI, Dence CS, Lee SY, Mach RH, Klunk WE, Mathis CA, DeKosky ST, Morris JC. [11C]PIB in a nondemented population: potential antecedent marker of Alzheimer disease. Neurology. 2006; 67:446–52. https://doi.org/10.1212/01.wnl.0000228230.26044.a4

https://doi.org/10.1212/01.wnl.0000228230.26044.a4 PMID:<u>16894106</u>

- Aizenstein HJ, Nebes RD, Saxton JA, Price JC, Mathis CA, Tsopelas ND, Ziolko SK, James JA, Snitz BE, Houck PR, Bi W, Cohen AD, Lopresti BJ, et al. Frequent amyloid deposition without significant cognitive impairment among the elderly. Arch Neurol. 2008; 65:1509–17. <u>https://doi.org/10.1001/archneur.65.11.1509</u> PMID:19001171
- Roeback JR Jr, Hla KM, Chambless LE, Fletcher RH. Effects of chromium supplementation on serum highdensity lipoprotein cholesterol levels in men taking beta-blockers. A randomized, controlled trial. Ann Intern Med. 1991; 115:917–24. <u>https://doi.org/10.7326/0003-4819-115-12-917</u> PMID:1683196

- Bouwman FH, Schoonenboom NS, Verwey NA, van Elk EJ, Kok A, Blankenstein MA, Scheltens P, van der Flier WM. CSF biomarker levels in early and late onset Alzheimer's disease. Neurobiol Aging. 2009; 30:1895–901. https://doi.org/10.1016/j.neurobiolaging.2008.02.007 PMID:18403055
- Jack CR Jr, Bennett DA, Blennow K, Carrillo MC, Dunn B, Haeberlein SB, Holtzman DM, Jagust W, Jessen F, Karlawish J, Liu E, Molinuevo JL, Montine T, et al, and Contributors. NIA-AA Research Framework: toward a biological definition of Alzheimer's disease. Alzheimers Dement. 2018; 14:535–62. <u>https://doi.org/10.1016/j.jalz.2018.02.018</u> PMID:29653606
- Zetterberg H, Hietala MA, Jonsson M, Andreasen N, Styrud E, Karlsson I, Edman A, Popa C, Rasulzada A, Wahlund LO, Mehta PD, Rosengren L, Blennow K, Wallin A. Neurochemical aftermath of amateur boxing. Arch Neurol. 2006; 63:1277–80. <u>https://doi.org/10.1001/archneur.63.9.1277</u> PMID:<u>16966505</u>
- Preische O, Schultz SA, Apel A, Kuhle J, Kaeser SA, Barro C, Gräber S, Kuder-Buletta E, LaFougere C, Laske C, Vöglein J, Levin J, Masters CL, et al, and Dominantly Inherited Alzheimer Network. Serum neurofilament dynamics predicts neurodegeneration and clinical progression in presymptomatic Alzheimer's disease. Nat Med. 2019; 25:277–83. https://doi.org/10.1038/s41591-018-0304-3

https://doi.org/10.1038/s41591-018-0 PMID:<u>30664784</u>

 Kern S, Syrjanen JA, Blennow K, Zetterberg H, Skoog I, Waern M, Hagen CE, van Harten AC, Knopman DS, Jack CR Jr, Petersen RC, Mielke MM. Association of Cerebrospinal Fluid Neurofilament Light Protein With Risk of Mild Cognitive Impairment Among Individuals Without Cognitive Impairment. JAMA Neurol. 2019; 76:187–193.

https://doi.org/10.1001/jamaneurol.2018.3459 PMID:<u>30419087</u>

- Mattsson N, Andreasson U, Zetterberg H, Blennow K. Alzheimer's Disease Neuroimaging I. Association of Plasma Neurofilament Light With Neurodegeneration in Patients With Alzheimer Disease. JAMA Neurol. 2017; 74:557–66. <u>https://doi.org/10.1001/jamaneurol.2016.6117</u> PMID:28346578
- Jack CR Jr, Knopman DS, Jagust WJ, Shaw LM, Aisen PS, Weiner MW, Petersen RC, Trojanowski JQ. Hypothetical model of dynamic biomarkers of the Alzheimer's pathological cascade. Lancet Neurol. 2010; 9:119–28. <u>https://doi.org/10.1016/S1474-4422(09)70299-6</u> PMID:<u>20083042</u>

- Petzold A. Neurofilament phosphoforms: surrogate markers for axonal injury, degeneration and loss. J Neurol Sci. 2005; 233:183–98. <u>https://doi.org/10.1016/j.jns.2005.03.015</u> PMID:15896809
- Bacioglu M, Maia LF, Preische O, Schelle J, Apel A, Kaeser SA, Schweighauser M, Eninger T, Lambert M, Pilotto A, Shimshek DR, Neumann U, Kahle PJ, et al. Neurofilament Light Chain in Blood and CSF as Marker of Disease Progression in Mouse Models and in Neurodegenerative Diseases. Neuron. 2016; 91:494–96. <u>https://doi.org/10.1016/j.neuron.2016.07.007</u> PMID:27477021
- Buerger K, Ewers M, Pirttilä T, Zinkowski R, Alafuzoff I, Teipel SJ, DeBernardis J, Kerkman D, McCulloch C, Soininen H, Hampel H. CSF phosphorylated tau protein correlates with neocortical neurofibrillary pathology in Alzheimer's disease. Brain. 2006; 129:3035–41. <u>https://doi.org/10.1093/brain/awl269</u> PMID:<u>17012293</u>
- Jack CR Jr, Lowe VJ, Weigand SD, Wiste HJ, Senjem ML, Knopman DS, Shiung MM, Gunter JL, Boeve BF, Kemp BJ, Weiner M, Petersen RC, and Alzheimer's Disease Neuroimaging Initiative. Serial PIB and MRI in normal, mild cognitive impairment and Alzheimer's disease: implications for sequence of pathological events in Alzheimer's disease. Brain. 2009; 132:1355–65. <u>https://doi.org/10.1093/brain/awp062</u> PMID:<u>19339253</u>
- Bobinski M, de Leon MJ, Wegiel J, Desanti S, Convit A, Saint Louis LA, Rusinek H, Wisniewski HM. The histological validation of post mortem magnetic resonance imaging-determined hippocampal volume in Alzheimer's disease. Neuroscience. 2000; 95:721–25. <u>https://doi.org/10.1016/S0306-4522(99)00476-5</u> PMID:<u>10670438</u>
- Mormino EC, Kluth JT, Madison CM, Rabinovici GD, Baker SL, Miller BL, Koeppe RA, Mathis CA, Weiner MW, Jagust WJ, and Alzheimer's Disease Neuroimaging Initiative. Episodic memory loss is related to hippocampal-mediated beta-amyloid deposition in elderly subjects. Brain. 2009; 132:1310–23. <u>https://doi.org/10.1093/brain/awn320</u> PMID:<u>19042931</u>
- Jack CR Jr, Dickson DW, Parisi JE, Xu YC, Cha RH, O'Brien PC, Edland SD, Smith GE, Boeve BF, Tangalos EG, Kokmen E, Petersen RC. Antemortem MRI findings correlate with hippocampal neuropathology in typical aging and dementia. Neurology. 2002; 58:750–57. <u>https://doi.org/10.1212/WNL.58.5.750</u> PMID:<u>11889239</u>
- 19. Silbert LC, Quinn JF, Moore MM, Corbridge E, Ball MJ, Murdoch G, Sexton G, Kaye JA. Changes in premorbid

brain volume predict Alzheimer's disease pathology. Neurology. 2003; 61:487–92. https://doi.org/10.1212/01.WNL.0000079053.77227.14 PMID:12939422

- Mattsson N, Zetterberg H, Janelidze S, Insel PS, Andreasson U, Stomrud E, Palmqvist S, Baker D, Tan Hehir CA, Jeromin A, Hanlon D, Song L, Shaw LM, et al, and ADNI Investigators. Plasma tau in Alzheimer disease. Neurology. 2016; 87:1827–35. <u>https://doi.org/10.1212/WNL.000000000003246</u> PMID:<u>27694257</u>
- Olsson B, Lautner R, Andreasson U, Öhrfelt A, Portelius E, Bjerke M, Hölttä M, Rosén C, Olsson C, Strobel G, Wu E, Dakin K, Petzold M, et al. CSF and blood biomarkers for the diagnosis of Alzheimer's disease: a systematic review and meta-analysis. Lancet Neurol. 2016; 15:673–84. https://doi.org/10.1016/S1474-4422(16)00070-3 PMID:27068280
- Alcolea D, Vilaplana E, Suárez-Calvet M, Illán-Gala I, Blesa R, Clarimón J, Lladó A, Sánchez-Valle R, Molinuevo JL, García-Ribas G, Compta Y, Martí MJ, Piñol-Ripoll G, et al. CSF sAPPβ, YKL-40, and neurofilament light in frontotemporal lobar degeneration. Neurology. 2017; 89:178–88.

https://doi.org/10.1212/WNL.000000000004088 PMID:28592456

- Rojas JC, Karydas A, Bang J, Tsai RM, Blennow K, Liman V, Kramer JH, Rosen H, Miller BL, Zetterberg H, Boxer AL. Plasma neurofilament light chain predicts progression in progressive supranuclear palsy. Ann Clin Transl Neurol. 2016; 3:216–25. <u>https://doi.org/10.1002/acn3.290</u> PMID:<u>27042681</u>
- Landau SM, Harvey D, Madison CM, Reiman EM, Foster NL, Aisen PS, Petersen RC, Shaw LM, Trojanowski JQ, Jack CR Jr, Weiner MW, Jagust WJ, and Alzheimer's Disease Neuroimaging Initiative. Comparing predictors of conversion and decline in mild cognitive impairment. Neurology. 2010; 75:230–38. <u>https://doi.org/10.1212/WNL.0b013e3181e8e8b8</u> PMID:20592257
- Rohrer JD, Woollacott IO, Dick KM, Brotherhood E, Gordon E, Fellows A, Toombs J, Druyeh R, Cardoso MJ, Ourselin S, Nicholas JM, Norgren N, Mead S, et al. Serum neurofilament light chain protein is a measure of disease intensity in frontotemporal dementia. Neurology. 2016; 87:1329–36.

https://doi.org/10.1212/WNL.00000000003154 PMID:<u>27581216</u>