# **Circular RNAs in leukemia**

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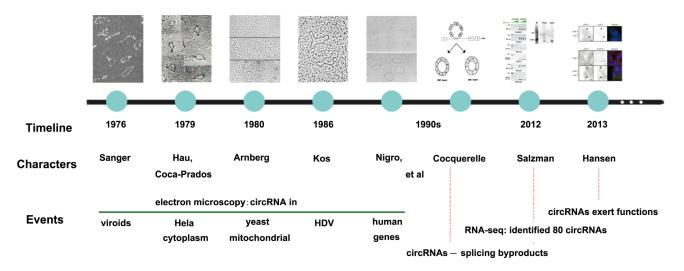
# ABSTRACT

In pace with the development of gene sequencing technology and transcriptome research, it has been found that 70 to 90% of the human genome is transcribed into RNAs, while only 2% of RNAs encode proteins. This implies that non-coding RNAs (ncRNAs) may exert vital biological functions and a full analysis of non-coding transcriptomes is needed. Over the past decade, the advance in high-throughput sequencing and transcriptome profiling has enabled the identification of circular RNAs (circRNAs) involved in many biological processes and the occurrence and development of diseases. Accumulating evidence has revealed that circRNAs may serve as new biomarkers for diagnosis as well as provide promising therapeutic approaches and novel drug screening strategies for leukemia. A comprehensive understanding of circRNAs in leukemia is a prerequisite for the development of clinical translational research. In this review, we will discuss the general information of circRNAs and focus on the current advances in understanding the association between dysregulated circRNAs and leukemia.

#### circRNAs BACKGROUND

In 1976, since Sanger [1] discovered a closed circular single-stranded RNA molecule with covalent bonds in viroids through electron microscopy, the concept of circRNAs came into being for the first time. Three years later, Hsu and Coca-Prados [2] extracted RNA from Hela cytoplasm, and the electron microscopy revealed that 1-2% of them were circRNAs. They also suggested that the majority of circRNAs may localize in cytoplasm rather than in nucleus. In 1980, Arnberg [3] observed that about 50% of the yeast mitochondrial RNA which was separated by agarose gel electrophoresis from 11S and 18S fragments contained circular molecules and was also confirmed in saccharomyces cerevisiae by ten years [4]. In 1986, Kos [5] presented HDV contained a single-stranded circular

RNA molecule. As early as 1990s, first endogenous circRNA with inverted order of exons was found in human DCC by Nigro [6], followed by human protooncogene ests-1 and mice Sry genes [7, 8]. Before 2000s, circRNAs were regarded as either splicing byproducts or intermediates escaping lariat structures from debranching [8-10]. Not until 2012, along with the mushroom growth of high-throughput sequencing and microarray technology as well as the application of bioinformatics, circRNAs had begun to be recognized as large species of RNAs with thousands of members in mammalian cells. Salzman [11] first reported about 80 circRNAs based on RNA sequencing results. Subsequently, more than 25,000 circRNAs in human fibroblasts, 1,950 circRNAs in HEK293 cells, 1,903 circRNAs in mouse brain tissue and 724 circRNAs in nematodes were found [12]. Thus, circRNAs are



**Figure 1. The development history of circRNAs.** The figure represents a summary of important events leading to the discovery of circRNAs.

thought to be important transcriptional products. Danan has found abundant circRNAs which might have certain biological functions [13]. Memczak [14] presented the opinion that coding sequences possessed previously unrecognized regulatory potential. In addition, Hansen' s [15] study, the first functional analysis of a naturally expressed circRNA, together with above-mentioned study, has aroused researchers' intense interest in circRNAs research. The main development trends of circRNAs were shown in Figure 1.

CircRNAs, a family of single-strand ncRNAs with covalently closed circular isoforms, are generated from precursor mRNAs (pre-mRNAs) [16] and can be regulated by specific RNA splicing factor which are involved in 'backsplicing' reaction [17]. Other than functioning as miRNA sponges [18-20], circRNAs could exert their bioactivities by interacting with proteins [21, 22]. For example, circ-Foxo3 was found to be associated with cell cycle progression and proliferation via forming ternary complexes with p21 and CDK2 [23]. In addition, circRNAs may play essential roles in regulating transcription and alternative splicing [24, 25]. CircRNAs that mainly distribute in cytoplasm promote transcription by interacting with the Pol II complex [26]. What's more, circRNAs can guide protein translation [27]. Abe [28] reported that circRNAs in cell-free Escherichia coli translation system could be efficiently translated by a rolling circle amplification (RCA) mechanism. Legnini [29] reported circ-znf609 could directly encode proteins and participate in the process of muscle development. More biological functions and significance of circRNAs are still under exploring.

Based on present study, widespread presence of circRNAs derived from various types of human samples is elucidated (Figure 2). It lays the foundation for unlimited potential of circRNAs. Xu [30] conducted a systematic examination of circRNAs in various human normal tissues. Further analysis suggested that circRNAs that closely related to biological functions were widely distributed and their expression was developmental stage- and tissue-specific. For instance, the 332 circRNAs enriched in bowel, colon and large intestine probably functioned in the digestive system, while the 243 circRNAs abundant in prostate and thyroid might play critical roles in male reproducing or development [31]. Some circRNAs with abnormal expression in cancer tissues were revealed to be involved in tumorigenesis, progress and metastasis and serve as possible biomarkers and new targets for treatment [32, 33]. There is evidence that plentiful circRNAs are components of blood cells [11]. Bonizzato [34] reported numbers of circRNAs in blood cells at different stages of hematopoietic differentiation, corroborating earlier suggestions that circRNAs expression were developmental stage- and tissuespecific. Moreover, abundantly presented in tumor cells, circRNAs are verified to play roles in tumor cell proliferation and metastasis [35]. CircRNAs are also detected in body fluid such as blood samples [36], cerebrospinal fluid [37], saliva [38], gastric juice [39] and even urine [40]. As a surrogate of non-invasive biomarker, circRNAs may have a chance to shine in the future. In addition, circRNAs are believed to present in human tears, milk and other body fluids. It will be quite an exciting news if circRNAs in body fluids can be verified individual-specific or disease-specific, which will alleviate patients' pain to a large extent in clinical

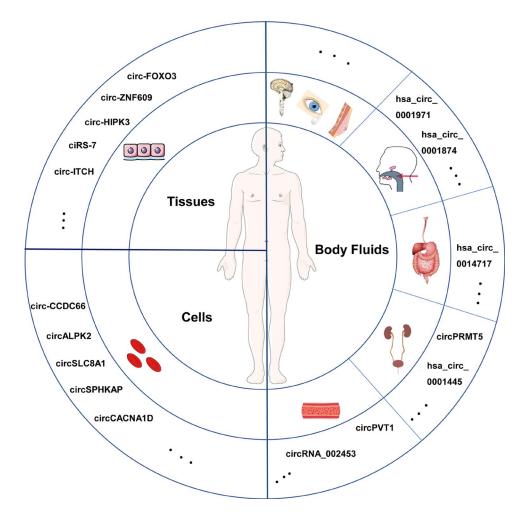


Figure 2. Different sources of circRNAs in human. CircRNAs are widespread in multiple human specimens, such as tissues, cells and diverse body fluids. Several circRNAs are listed as examples and dots in the circles represent circRNAs that have been identified and that still need to be explored.

tests, providing helpful information for diagnosis and treatment.

# **INTRODUCTION TO LEUKEMIA**

Leukemia is a life-threatening cancer with cloned hematopoietic stem cells and most often diagnosed in adults (mid 60s). The most common types of leukemia are acute mveloid leukemia (AML). acute lymphoblastic leukemia (ALL), chronic myeloid leukemia (CML) and chronic lymphocytic leukemia (CLL). With no clear preventive or control measures at present, leukemia is still a malignant disease and a threat to public health. Several studies have noted the important role of ncRNAs, such as microRNAs (miRNAs) and long non-coding RNAs (lncRNAs) in the pathogenesis of leukemia [41-43]. CircRNAs are expressed in both normal and malignant hematopoietic cells. In a certain sense, circRNAs have important implications for induced pluripotent stem cell (iPSC) modeling in hematopoietic system diseases [34]. It appears that circRNAs have more diverse expression profiles in cancer cells and may influence the protein expression levels of hematopoietic cell genes at different stages. Although overwhelming evidence has illustrated that circRNAs are of great importance to leukemia biogenesis, maintenance, and progression, a specific overview on circRNAs in leukemia is lacking. To comprehensively understand the functions and regulatory mechanisms of circRNAs in leukemia, we reviewed the remarkable and ground-breaking leukemia studies (Figure 3, Table 1).

#### Acute Myeloid Leukemia (AML)

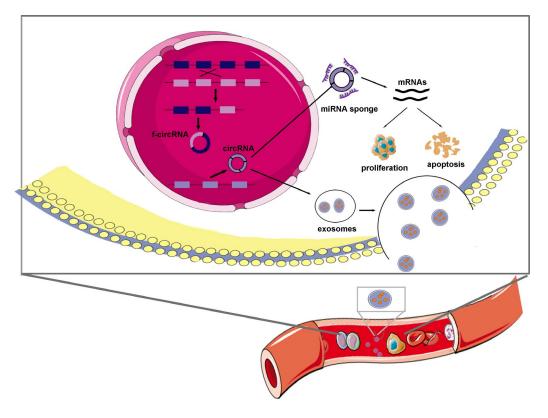
AML is the most common acute leukemia in adults. The incidence of AML increases with age and patients show significantly multifaceted biological and clinical heterogeneity. Recent studies have revealed the value of circRNAs in AML. While the expression pattern of

# Table 1. CircRNAs in leukemia.

Leukemia types	circRNA (circBase ID)	Host gene	Expression	Source	miRNA sponge (target gene/pathway)	Functions	Clinical significance	Reference
AML	f-circPR, f-circM9_1	PML/RARα, MLL/AF9	upregulated	BM	-	Facilitate cell transformation, vitality and resistance to treatment	Potential diagnostic and therapeutic implications	[44]
AML	circ-PVT1 (hsa_circ_0001821)	PVT1	upregulated	РВ	let-7 or miR-125 families*	Involved in the development of leukemia	Help to explore biological mechanisms behind MYC amplifications	[46]
AML	circNPM1 75001 (hsa_circ_0075001)	NPM1	upregulated	BM	miR-181 family/ TLR signaling pathway*	Associated with myeloid differentiation	A potential biomarker for classification and risk stratification	[51]
AML	circ-HIPK2 (hsa_circ_0001756)	HIPK2	downregulated	PB	miR-124-3p	Regulate ATRA-induced differentiation	A potential biomarker	[52]
AML	circ-ANAPC7 (has_circ_101141)	ANAPC7	upregulated	BM	miR-181 family*	Participate in the pathogenesis of AML	A promising diagnostic biomarker and novel drug target	[54]
AML	circRNA-DLEU2 (hsa_circ_0000488)	DLEU2	upregulated	BM	miR- 496/PRKACB	Promote cell proliferation and inhibited cell apoptosis	A novel biomarker and therapeutic target	[55]
AML	hsa_circ_100290	SLC30A7.	upregulated	BM	miR-203/Rab10	Promote cell proliferation and inhibited cell apoptosis	A potential diagnostic and therapeutic target	[56]

AML	hsa_circ_0004277	WDR37	downregulated	BM	miR-138-5p, miR-30c-1-3p, miR-892b, miR- 571, miR-328- 3p/SH3GL2, PPARGC1A, PIP4K2C, SH2B3, ZNF275, and ATP1B4*	Associated with risk-status and treatment	A potential diagnostic biomarker	[62]
AML	circ_0009910 (hsa_circ_100053)	MFN2	upregulated	BM	miR-20a-5p.	Promote cell proliferation, cell cycle arrest and inhibit c629ell apoptosis	A novel outcome biomarker and potential therapeutic targets	[63]
AML	circ-PAN3 (hsa_circ_0100181)	PAN3	upregulated	BM	miR-153-5p/miR- 183-5p/XIAP	Mediate the development of drug resistance	A valuable indicator for predicting clinical efficacy and potential target for reversing drug resistance	[64].
ALL	circPVT1	PVT1	upregulated	BM	let-7, miR-125*/ Bcl-2, c-Myc	Promote cell proliferation inhibit cell apoptosis	A potential therapeutic target	[70]
CML	circBA9.3	BCR-ABL1	upregulated	PB	-	Promote cell proliferation, TKI resistance and inhibit apoptosis	A potential diagnostic and therapeutic target for TKI-resistant patients	[73]
CLL	circ-CBFB (hsa_circ_0000707)	CBFB	upregulated	CLL cells	miR- 607/ <i>FZD3</i> /Wnt/β- catenin	Promote CLL cell proliferation, cell cycle progression and inhibit cellular apoptosis	An effective diagnostic and prognostic biomarker	[77]
CLL	circ_0132266 (hsa_circ_0132266)	MTO1	downregulated	CLL cells	miR-337-3p/PML	Inhibit CLL cell proliferation, and promote apoptosis	An effective diagnostic and prognostic biomarker	[78]

This table summarizes circRNAs that have been identified in human leukemia (AML, ALL, CML, CLL) and their roles in diagnosis, prognosis, disease progression, drug response evaluation. Information including host genes, expression levels, miRNA targets (direct and indirect), clinical values are shown (\*Not validated experimentally; -Not mentioned).



**Figure 3. Schematic representation of the proposed mechanism of circRNAs in leukemia.** CircRNAs and f-circRNAs could be transcribed separately from genes and fusion genes. CircRNAs that predominantly distribute in cytoplasm, play parts in leukemia mainly via sponging specific miRNAs and thus modulating mRNAs roles in cell proliferation and apoptosis. Exosomes are possible transport vectors that facilitate circRNAs circulating in blood and others and subsequently promoting leukemogenesis and progression.

circRNAs might be impaired anomalous in leukemia cells, altered circRNAs could be involved in leukemogenesis as well [44].

The cytogenetic and molecular genetic characteristics are significant prognostic factors for AML patients. In some cases, circRNAs make us better understand the pathogenic mechanism of AML. Despite the fact that some genomic amplicons are related to poor prognosis, the internal structures of amplicon formation and their potential mechanisms in AML have not yet been clarified yet. 8q24 amplifications are associated with aberrant chimeric genes and could generate oncogenic lncRNAs that play functional roles in leukemia [45]. Circ-PVT1, generated from exon 2 of PVT1 on chromosome 8, is often accompanied by 8q24 amplicons [46]. Circ-PVT1 was reported to act as a molecular sponge for tumor suppressor miRNAs, such as let-7 family [47] or miR-125 family [48]. The expression level of circ-PVT1 in AML patients with amplifications (AML-amp) was significantly higher than the patients with normal karyotype. The discovery of posttranscriptional chimeras and circRNAs involved with 8q24 amplified genes in AML-amp cases could broaden our exploration of the underlying mechanisms of in human leukemia.

Nucleophosmin (NPM1) was regarded as one of the most frequently mutated genes in AML. Bearing both proto-oncogene and tumor-suppressing properties, NPM1 could be involved in ribosomal biogenesis, apoptosis, and cell proliferation by encoding a multifunctional chaperone protein. The mutation of NPM1 might contribute to AML [49, 50]. Given the relevance of NPM1 in AML and aiming to deepen the understanding of multifaceted gene, Hirsch [51] explored the circRNA variants from NPM1 through RNA-Seq-based transcriptome analysis. In NPM1 wildtype and mutated AML patients, circNPM1 75001 was highly expressed in AML patients. Although circNPM1 75001 was independent of the NPM1 mutational status, a positive correlation was found between the expression of circNPM1 and total NPM1. Li's study also suggested an independent relationship between circRNAs and their linear host mRNAs [52]. Overall, these findings would contribute to distinguishing different AML subgroups based on circRNA expression profile and

offering new insight into exploring the biological characteristics of leukemia cells.

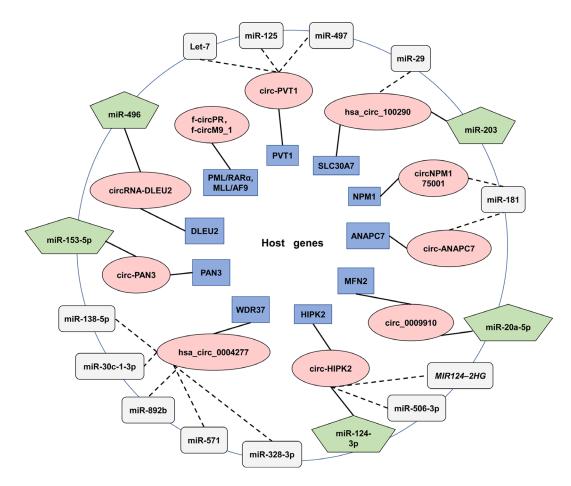
Fusion genes formed by two unrelated genes are very common in cancer genomes. Displaying adverse effects, fusion genes are important cancer markers in diseases [53]. In leukemia, chromosomal translocation often leads to gene fusion. The discovery of fusion circRNAs (f-circRNAs) and the study of their functions has created a new insight into understanding the mechanisms of leukemia and tailoring therapies for patients. Guarnerio [44] studied two types of leukemia, acute promyelocytic leukemia (APL) with translocation between PML and RARa, and AML with translocation between MLL and AF9. Based on the concept that one single gene can generate more than one circRNA, they elucidated that the corresponding fusion genes could produce f-circRNAs. Moreover, f-circRNAs can facilitate cell transformation, vitality and resistance to treatment. While f-circRNAs alone were not enough to trigger leukemia, they appeared to work with other cancer-promoting signals, such as MLL-AF9 fusion protein, to promote disease progression. It suggested that circRNAs, such as protein genes, are also affected by the rearrangement of genes, resulting in abnormal fusions. These aberrant f-circRNAs further contribute to the development of leukemia and may be an important target for antitumor drugs. However, the specific molecular mechanism of the function of f-circRNAs requires intensive study.

CircRNAs can serve as novel promising diagnosis biomarkers for AML. For example, circ-ANAPC7 which was validated upregulated frequently in AML patients was proposed to be a potential indicator for AML diagnosis [54]. circRNA-DLEU2 [55] and hsa circ 100290 [56] were also potential biomarkers identified and verified to be highly expressed in AML patients. Consistent with the commonly reported competitive endogenous RNA (ceRNA) mechanism, circRNA-DLEU2 promoted cell proliferation and reproduction through the miR-496/PRKACB axis. Hsa circ 100290 reported to function in oral squamous cell carcinomas [57] was revealed to act as an oncogene in AML occurrence and progression, promoting cell proliferation and inhibiting apoptosis at the same time. potential elucidate the mechanism To of hsa circ 100290, researchers further screened for its targeted miRNAs. As expected, hsa circ 100290 can affect cell functions via regulating miR-203/Rab10 axis. Approximately 45-50% of AML cases showed no obvious genomic alterations when assessed using conventional banding analysis [49, 58]. To diagnose and distinguish APL patients from cytogenetically normal (CN) AML patients, circRNA profiles were analyzed using acfs (accurate circRNA finder suite). A set of differentially expressed circRNAs were identified, and the results illustrated that circRNAs could facilitate the molecular stratification of AML [59]. The multiple research has identified novel genetic alterations and provided promising biomarkers for AML diagnosis and classification.

Aside from being biomarkers in leukemia, circRNAs could also instruct the treatment and predict prognosis of patients. Extramedullary infiltration (EMI), which always corresponds with relapse and refractory, is relatively common in AML [60]. Given that EMI is closely correlated with poor prognosis in AML patients, Lv [61] carried out circRNA microarrays in bone marrow mononuclear cells from EMI and non-EMI AML patients to uncover novel abnormal moleculars. 17 circRNAs were noted to act as key regulators of cell-cell crosstalk in EMI, and most of their target genes could predicted poor prognosis. Several molecular abnormalities were considered as crucial factors affecting AML treatment effect. To offer a potential diagnostic marker and treatment target in AML, Li [62] performed an analysis using a microarray platform and intensity filtering system. Through a comparative analysis of AML patients and healthy hsa circ 0004277 patients, was selected from characterized circRNA transcripts. It was found to be aberrantly decreased in CN-AML patients but significantly elevated after treatment. It is suggested that hsa circ 0004277 was strongly associated with the treatment of AML. In addition, circ 0009910 was found to highly express in AML. And higher expression levels of circ 0009910 usually predicted worse prognosis [63]. One of the obstacles in AML patients survival is the resistance to chemotherapeutic drugs. A highthroughput circRNA microarray was conducted to compare the expression profiles of circRNAs in chemosensitive and resistant AML cells. As a result, 49 circRNAs were noted to be significantly differentially expressed. Additionally, circPAN3 was confirmed to have a vital function in driving drug resistance through circPAN3-miR-153-5p/miR-183-5p-XIAP axis [64]. The majority of circRNAs were reported to alleviate the effects induced by miRNAs. Regulatory relationships between circRNAs, miRNA and their target genes were deciphered in Figure 4. To sum up, the expression levels of circRNAs are conductive to clinical diagnosis. treatment and prognosis assessment.

# Acute Lymphoblastic Leukemia (ALL)

ALL is a lymphoid malignancy affecting the B or T lineages. Its incidence peaks are between ages 2 and 5 years and the elders [65]. Emerging evidence shows that circRNAs may confer in ALL and have potential values in indicating diagnosis and therapy.



**Figure 4. CircRNAs and their correlation with miRNAs in AML.** The figure summarizes the reported circRNAs that functioning in AML. The innermost circle described host genes (Blue rectangle) and the red ovals in the middle are circRNAs that transcribed from them. The outermost circle exhibited targeted miRNAs. Solid lines connect circRNAs and their target miRNAs (Green pentagon). Dashed lines are used to connect miRNAs (Grey box) that have not been validated experimentally or proposed to interact with circRNAs in other diseases.

Based on RNA-seq results in hyperdiploid B-cell precursor ALL samples, circRNAs were found to be produced from one specific gene and differentially expressed in cells and diseases [66]. For example, circPVT1 which was reported to be involved in gastric cancer [67], AML [46] and other diseases [48, 68, 69] was also specifically highly expressed in ALL. Silencing of circPVT1 induced cell cycle arrest and apoptosis of ALL cells via targeting its neighbor gene c-Myc and increasing the expression level of antiapoptotic protein Bcl-2 [70]. In addition, it has been reported that circRNAs generated from genes that related to ALL and B-cell differentiation (JAK2, PAX5, IKZF1, ETV6 and EBF1) are prevalently present in hyperdiploid leukemia compared with normal leukocytes samples [34].

Adults and senile patients of AML are often companied with poor outcomes. Although most children with ALL can be cured, a small number of patients who relapse after treatment have poor prognosis [66]. Given the fact that the circRNA expression pattern was dysregulated in ALL patients, efforts should be made to better understand its molecular mechanisms in ALL occurrence and relapse.

#### **Chronic Myeloid Leukemia (CML)**

Chronic myeloid leukemia (CML) is a clonal myeloproliferative neoplasm originating from abnormal pluripotent stem cells. The oncogene *BCR-ABL1* which encodes a hyperactive tyrosine kinase is the major molecular factor for the pathogenesis of CML [71]. Imatinib, a specific tyrosine kinase inhibitor (TKI), has brought hope for the majority of CML patients. A small proportion of patients, however, have developed resistance to imatinib [72]. Recently, circRNAs have been implicated in the progression of CML and development of TKI resistance and opened up new opportunities for CML therapy.

To explore the internal mechanism of chemo-resistance and remove the obstacles to cure patients, f-circRNA circBA9.3 derived from the *BCR-ABL1* fusion genes was analyzed. Of note, circBA9.3 which increased the protein expression levels of ABL1 and BCR-ABL1 was associated with TKI resistance. Over-expressed in leukemia cells, circBA9.3 could facilitate cell proliferation and drug resistance. It is inspiring that circBA9.3 may serve as a promising diagnostic biomarker and therapeutic target for CML patients [73].

## Chronic Lymphocytic Leukemia (CLL)

CLL is a heterogeneous disease characterized by the monoclonal expansion of B cells. It is the most common adult leukemia in the West and the median age at diagnosis is 71 years [74, 75]. Because of its insidious onset and significant heterogeneity, the molecular details of CLL are still under investigation.

Based on the RNA-seq data, 52 circRNAs were identified and the role of one circRNA in distinguishing different B-cell malignancies was suggested [76]. Hsa circ 0000707, termed circ-CBFB, derived from CBFB on chromosome 16 and aberrantly overexpressed in CLL patients, was determined to promote cell proliferation and suppress apoptosis through miR-607/FZD3/Wnt/β-catenin pathway [77]. Given the connection between the signaling cascade and CLL progression, circ-CBFB may become a potential therapeutic target for CLL treatment. Specific miRNA in different diseases may exhibit different expression To figure out why miR-337-3p was levels. downregulated in CLL, Wu [78] has elucidated possible regulatory mechanism that circ 0132266 participated in the downregulating of miR-337-3p expression, which opens a new avenue for exploring the interaction between specific miRNA and circRNAs underlying CLL. Taken together, these data has demonstrated that circRNAs may serve as oncogenes or tumor suppressors and participate in CLL progression.

Although the molecular mechanisms of circRNAs are poorly understood in leukemia, they are indeed implicated in cellular processes and leukemia progression. Encouragingly, the exploration of aberrant circRNAs has provided us with valuable insight into the pathogenesis of hematological malignancies.

# PERSPECTIVES

Originated from human cancer xenografts, circRNAs could enter the circulation by exosomes and be easily detected in serum or plasma. Exosomes are small extracellular vesicles (EVs) (30–150 nm in size) [79] that are produced by normal or cancerous cells,

including leukemia blasts [80, 81]. They may play pleiotropic roles in regulating signaling transduction and supporting the growth and survival of leukemia cells by delivering mRNAs, ncRNAs and proteins between neighboring or distant cells [82, 83]. Exosomes can provide not only diagnostic and prognostic information for leukemia but also antileukemic treatment modalities [84, 85]. Released vesicles, such as exosomes and microvesicles, might be possible vehicles by which circRNAs are expulsed to extracellular space because enriched circRNAs have been detected in EVs [86, 87]. Researchers have also discovered hundreds of circRNAs in cellular that can be transferred to exosomes and proposed that exosomal circRNAs (exocircRNAs) traffic through a complex mechanism [88]. Furthermore, since EVs can be taken up by other cells. circRNAs could mediate intercellular excreted communication [89]. Given that exosomes can carry circRNAs and transfer them out of cells while retaining the integrity of their structures, we consider that circRNAs who are abundantly and stably present in the blood circulatory system may play nonnegligible roles in leukemogenesis, progression and drug resistance. However, little is known about exo-circRNAs in leukemia. Herein, much efforts are needed to comprehensively understand the roles of circRNAs.

A big step has been taken toward understanding the pathogenesis of leukemia since f-circRNAs have been identified and verified to promote leukemogenesis and other diseases [44, 90]. Chromosomal translocations are responsible for the onset of many types of cancers, such as leukemia. These discoveries strongly arouse our interest in seeking more f-circRNAs that have great impacts on disease development, even independent of linear transcripts and fusion proteins. Notably, many studies have shown the potential of circRNAs as promising therapeutic targets. Encouragingly, an artificial circRNA that serves as miRNA "sponge" has been designed in vitro to treat and cure Hepatitis C. It can not only adsorb miRNA but also adhere to binding proteins, fulfilling their role in preventing viral infection [91]. Once artificial circRNA is presented as a medical treatment, it will highly possible to find novel and better treatments for leukemia and many other diseases.

#### CONCLUSIONS

In this review, we summarize the latest research progress of circRNAs and highlight their biological significance and clinical values in leukemia. CircRNAs have been shown to play nonnegligible roles as gene regulators and perform crucial biological functions via ceRNA mechanism in leukemia. Other than indicating the occurrence of leukemia, circRNAs have the potential to be indicators of disease stage and progression and could be developed as medication for clinical use. With more in-depth study, we expect that circRNAs can be more appropriately and precisely used in clinical diagnosis and treatment in the future.

## **CONFLICTS OF INTEREST**

The authors declare that they have no conflict of interest.

## **FUNDING**

This study was supported by National Natural Science Foundation of China (Grant no. 81700155, 81720108002).

# **REFERENCES**

- Sanger HL, Klotz G, Riesner D, Gross HJ, Kleinschmidt AK. Viroids are single-stranded covalently closed circular RNA molecules existing as highly base-paired rod-like structures. Proc Natl Acad Sci USA. 1976; 73:3852–56. <u>https://doi.org/10.1073/pnas.73.11.3852</u> PMID:1069269
- Hsu MT, Coca-Prados M. Electron microscopic evidence for the circular form of RNA in the cytoplasm of eukaryotic cells. Nature. 1979; 280:339–40. <u>https://doi.org/10.1038/280339a0</u> PMID:<u>460409</u>
- Arnberg AC, Van Ommen GJ, Grivell LA, Van Bruggen EF, Borst P. Some yeast mitochondrial RNAs are circular. Cell. 1980; 19:313–19. <u>https://doi.org/10.1016/0092-8674(80)90505-X</u> PMID:<u>6986989</u>
- Matsumoto Y, Fishel R, Wickner RB. Circular singlestranded RNA replicon in Saccharomyces cerevisiae. Proc Natl Acad Sci USA. 1990; 87:7628–32. <u>https://doi.org/10.1073/pnas.87.19.7628</u> PMID:<u>1699230</u>
- 5. Kos A, Dijkema R, Arnberg AC, van der Meide PH, Schellekens H. The hepatitis delta (delta) virus possesses a circular RNA. Nature. 1986; 323:558–60. https://doi.org/10.1038/323558a0 PMID:2429192
- Nigro JM, Cho KR, Fearon ER, Kern SE, Ruppert JM, Oliner JD, Kinzler KW, Vogelstein B. Scrambled exons. Cell. 1991; 64:607–13. <u>https://doi.org/10.1016/0092-8674(91)90244-S</u> PMID:<u>1991322</u>
- 7. Capel B, Swain A, Nicolis S, Hacker A, Walter M, Koopman P, Goodfellow P, Lovell-Badge R. Circular

transcripts of the testis-determining gene Sry in adult mouse testis. Cell. 1993; 73:1019–30. https://doi.org/10.1016/0092-8674(93)90279-Y PMID:7684656

- Cocquerelle C, Mascrez B, Hétuin D, Bailleul B. Missplicing yields circular RNA molecules. FASEB J. 1993; 7:155–60. <u>https://doi.org/10.1096/fasebj.7.1.7678559</u> PMID:<u>7678559</u>
- 9. Qian L, Vu MN, Carter M, Wilkinson MF. A spliced intron accumulates as a lariat in the nucleus of T cells. Nucleic Acids Res. 1992; 20:5345–50. <u>https://doi.org/10.1093/nar/20.20.5345</u> PMID:<u>1437551</u>
- Pasman Z, Been MD, Garcia-Blanco MA. Exon circularization in mammalian nuclear extracts. RNA. 1996; 2:603–10. PMID:<u>8718689</u>
- Salzman J, Gawad C, Wang PL, Lacayo N, Brown PO. Circular RNAs are the predominant transcript isoform from hundreds of human genes in diverse cell types. PLoS One. 2012; 7:e30733. <u>https://doi.org/10.1371/journal.pone.0030733</u> PMID:<u>22319583</u>
- Jeck WR, Sorrentino JA, Wang K, Slevin MK, Burd CE, Liu J, Marzluff WF, Sharpless NE. Circular RNAs are abundant, conserved, and associated with ALU repeats. RNA. 2013; 19:141–57. <u>https://doi.org/10.1261/rna.035667.112</u> PMID:<u>23249747</u>
- Danan M, Schwartz S, Edelheit S, Sorek R. Transcriptome-wide discovery of circular RNAs in Archaea. Nucleic Acids Res. 2012; 40:3131–42. <u>https://doi.org/10.1093/nar/gkr1009</u> PMID:22140119
- 14. Memczak S, Jens M, Elefsinioti A, Torti F, Krueger J, Rybak A, Maier L, Mackowiak SD, Gregersen LH, Munschauer M, Loewer A, Ziebold U, Landthaler M, et al. Circular RNAs are a large class of animal RNAs with regulatory potency. Nature. 2013; 495:333–38. <u>https://doi.org/10.1038/nature11928</u> PMID:23446348
- Hansen TB, Jensen TI, Clausen BH, Bramsen JB, Finsen B, Damgaard CK, Kjems J. Natural RNA circles function as efficient microRNA sponges. Nature. 2013; 495:384–88. <u>https://doi.org/10.1038/nature11993</u> PMID:23446346
- 16. Barrett SP, Wang PL, Salzman J. Circular RNA biogenesis can proceed through an exon-containing lariat precursor. eLife. 2015; 4:e07540.

https://doi.org/10.7554/eLife.07540 PMID:26057830

- Yu CY, Li TC, Wu YY, Yeh CH, Chiang W, Chuang CY, Kuo HC. The circular RNA circBIRC6 participates in the molecular circuitry controlling human pluripotency. Nat Commun. 2017; 8:1149. <u>https://doi.org/10.1038/s41467-017-01216-w</u> PMID:<u>29074849</u>
- Han D, Li J, Wang H, Su X, Hou J, Gu Y, Qian C, Lin Y, Liu X, Huang M, Li N, Zhou W, Yu Y, Cao X. Circular RNA circMTO1 acts as the sponge of microRNA-9 to suppress hepatocellular carcinoma progression. Hepatology. 2017; 66:1151–64. https://doi.org/10.1002/hep.29270\_PMID:28520103
- Wang K, Long B, Liu F, Wang JX, Liu CY, Zhao B, Zhou LY, Sun T, Wang M, Yu T, Gong Y, Liu J, Dong YH, et al. A circular RNA protects the heart from pathological hypertrophy and heart failure by targeting miR-223. Eur Heart J. 2016; 37:2602–11. <u>https://doi.org/10.1093/eurheartj/ehv713</u> PMID:<u>26802132</u>
- 20. Zheng Q, Bao C, Guo W, Li S, Chen J, Chen B, Luo Y, Lyu D, Li Y, Shi G, Liang L, Gu J, He X, Huang S. Circular RNA profiling reveals an abundant circHIPK3 that regulates cell growth by sponging multiple miRNAs. Nat Commun. 2016; 7:11215. <u>https://doi.org/10.1038/ncomms11215</u> PMID:<u>27050392</u>
- Armakola M, Higgins MJ, Figley MD, Barmada SJ, Scarborough EA, Diaz Z, Fang X, Shorter J, Krogan NJ, Finkbeiner S, Farese RV Jr, Gitler AD. Inhibition of RNA lariat debranching enzyme suppresses TDP-43 toxicity in ALS disease models. Nat Genet. 2012; 44:1302–09. <u>https://doi.org/10.1038/ng.2434</u> PMID:<u>23104007</u>
- 22. Du WW, Zhang C, Yang W, Yong T, Awan FM, Yang BB. Identifying and Characterizing circRNA-Protein Interaction. Theranostics. 2017; 7:4183–91. https://doi.org/10.7150/thno.21299 PMID:29158818
- 23. Du WW, Yang W, Liu E, Yang Z, Dhaliwal P, Yang BB. Foxo3 circular RNA retards cell cycle progression via forming ternary complexes with p21 and CDK2. Nucleic Acids Res. 2016; 44:2846–58. <u>https://doi.org/10.1093/nar/gkw027</u> PMID:<u>26861625</u>
- 24. Li Z, Huang C, Bao C, Chen L, Lin M, Wang X, Zhong G, Yu B, Hu W, Dai L, Zhu P, Chang Z, Wu Q, et al. Exon-intron circular RNAs regulate transcription in the nucleus. Nat Struct Mol Biol. 2015; 22:256–64. https://doi.org/10.1038/nsmb.2959\_PMID:25664725

- 25. Chen LL. The biogenesis and emerging roles of circular RNAs. Nat Rev Mol Cell Biol. 2016; 17:205–11. <u>https://doi.org/10.1038/nrm.2015.32</u> PMID:26908011
- Zhang Y, Zhang XO, Chen T, Xiang JF, Yin QF, Xing YH, Zhu S, Yang L, Chen LL. Circular intronic long noncoding RNAs. Mol Cell. 2013; 51:792–806.

https://doi.org/10.1016/j.molcel.2013.08.017 PMID:24035497

- Chen CY, Sarnow P. Initiation of protein synthesis by the eukaryotic translational apparatus on circular RNAs. Science. 1995; 268:415–17. <u>https://doi.org/10.1126/science.7536344</u> PMID:<u>7536344</u>
- Abe N, Matsumoto K, Nishihara M, Nakano Y, Shibata A, Maruyama H, Shuto S, Matsuda A, Yoshida M, Ito Y, Abe H. Rolling Circle Translation of Circular RNA in Living Human Cells. Sci Rep. 2015; 5:16435. <u>https://doi.org/10.1038/srep16435</u> PMID:<u>26553571</u>
- Legnini I, Di Timoteo G, Rossi F, Morlando M, Briganti F, Sthandier O, Fatica A, Santini T, Andronache A, Wade M, Laneve P, Rajewsky N, Bozzoni I. Circ-ZNF609 Is a Circular RNA that Can Be Translated and Functions in Myogenesis. Mol Cell. 2017; 66:22–37.e9. <u>https://doi.org/10.1016/j.molcel.2017.02.017</u> PMID:<u>28344082</u>
- Xu T, Wu J, Han P, Zhao Z, Song X. Circular RNA expression profiles and features in human tissues: a study using RNA-seq data. BMC Genomics. 2017 (Suppl 6); 18:680. <u>https://doi.org/10.1186/s12864-017-4029-3</u> PMID:<u>28984197</u>
- 31. Liu YC, Chiu YJ, Li JR, Sun CH, Liu CC, Huang HD. Biclustering of transcriptome sequencing data reveals human tissue-specific circular RNAs. BMC Genomics. 2018 (Suppl 1); 19:958. <u>https://doi.org/10.1186/s12864-017-4335-9</u> PMID:<u>29363420</u>
- 32. Su Y, Xu C, Liu Y, Hu Y, Wu H. Circular RNA hsa\_circ\_0001649 inhibits hepatocellular carcinoma progression via multiple miRNAs sponge. Aging (Albany NY). 2019; 11:3362–75. <u>https://doi.org/10.18632/aging.101988</u> PMID:<u>31137016</u>
- 33. Hsiao KY, Lin YC, Gupta SK, Chang N, Yen L, Sun HS, Tsai SJ. Noncoding Effects of Circular RNA CCDC66 Promote Colon Cancer Growth and Metastasis. Cancer Res. 2017; 77:2339–50. <u>https://doi.org/10.1158/0008-5472.CAN-16-1883</u> PMID:<u>28249903</u>

- 34. Bonizzato A, Gaffo E, Te Kronnie G, Bortoluzzi S. CircRNAs in hematopoiesis and hematological malignancies. Blood Cancer J. 2016; 6:e483. <u>https://doi.org/10.1038/bcj.2016.81</u> PMID:27740630
- 35. Yang Y, Gao X, Zhang M, Yan S, Sun C, Xiao F, Huang N, Yang X, Zhao K, Zhou H, Huang S, Xie B, Zhang N. Novel Role of FBXW7 Circular RNA in Repressing Glioma Tumorigenesis. J Natl Cancer Inst. 2018; 110:304. <u>https://doi.org/10.1093/jnci/djx166</u> PMID:<u>28903484</u>
- Dong WW, Li HM, Qing XR, Huang DH, Li HG. Identification and characterization of human testis derived circular RNAs and their existence in seminal plasma. Sci Rep. 2016; 6:39080. https://doi.org/10.1038/srep39080 PMID:27958373
- Lu D, Xu AD. Mini Review: Circular RNAs as Potential Clinical Biomarkers for Disorders in the Central Nervous System. Front Genet. 2016; 7:53. <u>https://doi.org/10.3389/fgene.2016.00053</u> PMID:<u>27092176</u>
- 38. Zhao SY, Wang J, Ouyang SB, Huang ZK, Liao L. Salivary Circular RNAs Hsa\_Circ\_0001874 and Hsa\_Circ\_0001971 as Novel Biomarkers for the Diagnosis of Oral Squamous Cell Carcinoma. Cell Physiol Biochem. 2018; 47:2511–21. https://doi.org/10.1159/000491624\_PMID:29991057
- Shao Y, Li J, Lu R, Li T, Yang Y, Xiao B, Guo J. Global circular RNA expression profile of human gastric cancer and its clinical significance. Cancer Med. 2017; 6:1173–80. https://doi.org/10.1002/cam4.1055 PMID:28544609
- Chen X, Chen RX, Wei WS, Li YH, Feng ZH, Tan L, Chen JW, Yuan GJ, Chen SL, Guo SJ, Xiao KH, Liu ZW, Luo JH, et al. PRMT5 Circular RNA Promotes Metastasis of Urothelial Carcinoma of the Bladder through Sponging miR-30c to Induce Epithelial-Mesenchymal Transition. Clin Cancer Res. 2018; 24:6319–30. <u>https://doi.org/10.1158/1078-0432.CCR-18-1270</u> PMID:<u>30305293</u>
- Garzon R, Volinia S, Papaioannou D, Nicolet D, Kohlschmidt J, Yan PS, Mrózek K, Bucci D, Carroll AJ, Baer MR, Wetzler M, Carter TH, Powell BL, et al. Expression and prognostic impact of IncRNAs in acute myeloid leukemia. Proc Natl Acad Sci USA. 2014; 111:18679–84. <u>https://doi.org/10.1073/pnas.1422050112</u>

PMID:<u>25512507</u>

42. Rücker FG, Russ AC, Cocciardi S, Kett H, Schlenk RF, Botzenhardt U, Langer C, Krauter J, Fröhling S, Schlegelberger B, Ganser A, Lichter P, Zenz T, et al. Altered miRNA and gene expression in acute myeloid leukemia with complex karyotype identify networks of prognostic relevance. Leukemia. 2013; 27:353–61. https://doi.org/10.1038/leu.2012.208 PMID:22810507

- 43. Russ AC, Sander S, Lück SC, Lang KM, Bauer M, Rücker FG, Kestler HA, Schlenk RF, Döhner H, Holzmann K, Döhner K, Bullinger L. Integrative nucleophosmin mutation-associated microRNA and gene expression pattern analysis identifies novel microRNA - target gene interactions in acute myeloid leukemia. Haematologica. 2011; 96:1783–91. https://doi.org/10.3324/haematol.2011.046888 PMID:21880628
- 44. Guarnerio J, Bezzi M, Jeong JC, Paffenholz SV, Berry K, Naldini MM, Lo-Coco F, Tay Y, Beck AH, Pandolfi PP. Oncogenic Role of Fusion-circRNAs Derived from Cancer-Associated Chromosomal Translocations. Cell. 2016; 165:289–302. https://doi.org/10.1016/j.cell.2016.03.020 PMID:27040497
- 45. Burmeister T, Schwartz S, Horst HA, Rieder H, Gökbuget N, Hoelzer D, Thiel E. Molecular heterogeneity of sporadic adult Burkitt-type leukemia/lymphoma as revealed by PCR and cytogenetics: correlation with morphology, immunology and clinical features. Leukemia. 2005; 19:1391–98. <u>https://doi.org/10.1038/sj.leu.2403847</u> PMID:<u>15973450</u>
- L'Abbate A, Tolomeo D, Cifola I, Severgnini M, Turchiano A, Augello B, Squeo G, D'Addabbo P, Traversa D, Daniele G, Lonoce A, Pafundi M, Carella M, et al. MYC-containing amplicons in acute myeloid leukemia: genomic structures, evolution, and transcriptional consequences. Leukemia. 2018; 32:2152–66. <u>https://doi.org/10.1038/s41375-018-0033-0</u> PMID:29467491
- Panda AC, Grammatikakis I, Kim KM, De S, Martindale JL, Munk R, Yang X, Abdelmohsen K, Gorospe M. Identification of senescence-associated circular RNAs (SAC-RNAs) reveals senescence suppressor CircPVT1. Nucleic Acids Res. 2017; 45:4021–35. <u>https://doi.org/10.1093/nar/gkw1201</u> PMID:<u>27928058</u>
- Li X, Zhang Z, Jiang H, Li Q, Wang R, Pan H, Niu Y, Liu F, Gu H, Fan X, Gao J. Circular RNA circPVT1 Promotes Proliferation and Invasion Through Sponging miR-125b and Activating E2F2 Signaling in Non-Small Cell Lung Cancer. Cell Physiol Biochem. 2018; 51:2324–40. https://doi.org/10.1159/000495876 PMID:30537738
- 49. Döhner H, Estey E, Grimwade D, Amadori S, Appelbaum FR, Büchner T, Dombret H, Ebert BL,

Fenaux P, Larson RA, Levine RL, Lo-Coco F, Naoe T, et al. Diagnosis and management of AML in adults: 2017 ELN recommendations from an international expert panel. Blood. 2017; 129:424–47. https://doi.org/10.1182/blood-2016-08-733196 PMID:27895058

- 50. Falini B, Nicoletti I, Bolli N, Martelli MP, Liso A, Gorello P, Mandelli F, Mecucci C, Martelli MF. Translocations and mutations involving the nucleophosmin (NPM1) gene in lymphomas and leukemias. Haematologica. 2007; 92:519–32. <u>https://doi.org/10.3324/haematol.11007</u> PMID:<u>17488663</u>
- Hirsch S, Blätte TJ, Grasedieck S, Cocciardi S, Rouhi A, Jongen-Lavrencic M, Paschka P, Krönke J, Gaidzik VI, Döhner H, Schlenk RF, Kuchenbauer F, Döhner K, et al. Circular RNAs of the nucleophosmin (NPM1) gene in acute myeloid leukemia. Haematologica. 2017; 102:2039–47. https://doi.org/10.3324/haematol.2017.172866

PMID:28971903

- 52. Li S, Ma Y, Tan Y, Ma X, Zhao M, Chen B, Zhang R, Chen Z, Wang K. Profiling and functional analysis of circular RNAs in acute promyelocytic leukemia and their dynamic regulation during all-trans retinoic acid treatment. Cell Death Dis. 2018; 9:651. <u>https://doi.org/10.1038/s41419-018-0699-2</u> PMID:<u>29844435</u>
- 53. Bohlander SK. Fusion genes in leukemia: an emerging network. Cytogenet Cell Genet. 2000; 91:52–56. <u>https://doi.org/10.1159/000056818</u> PMID:<u>11173830</u>
- 54. Chen H, Liu T, Liu J, Feng Y, Wang B, Wang J, Bai J, Zhao W, Shen Y, Wang X, Yang J, Ji Y, He A, Yang Y. Circ-ANAPC7 is Upregulated in Acute Myeloid Leukemia and Appears to Target the MiR-181 Family. Cell Physiol Biochem. 2018; 47:1998–2007.

https://doi.org/10.1159/000491468 PMID:29969755

- 55. Wu DM, Wen X, Han XR, Wang S, Wang YJ, Shen M, Fan SH, Zhang ZF, Shan Q, Li MQ, Hu B, Chen GQ, Lu J, Zheng YL. Role of Circular RNA DLEU2 in Human Acute Myeloid Leukemia. Mol Cell Biol. 2018; 38:e00259-18. <u>https://doi.org/10.1128/MCB.00259-18</u> PMID:<u>30037980</u>
- 56. Fan H, Li Y, Liu C, Liu Y, Bai J, Li W. Circular RNA-100290 promotes cell proliferation and inhibits apoptosis in acute myeloid leukemia cells via sponging miR-203. Biochem Biophys Res Commun. 2018; 507:178–84. <u>https://doi.org/10.1016/j.bbrc.2018.11.002</u> PMID:<u>30424877</u>

- 57. Chen L, Zhang S, Wu J, Cui J, Zhong L, Zeng L, Ge S. circRNA\_100290 plays a role in oral cancer by functioning as a sponge of the miR-29 family. Oncogene. 2017; 36:4551–61. <u>https://doi.org/10.1038/onc.2017.89</u> PMID:<u>28368401</u>
- 58. Farag SS, Archer KJ, Mrózek K, Ruppert AS, Carroll AJ, Vardiman JW, Pettenati MJ, Baer MR, Qumsiyeh MB, Koduru PR, Ning Y, Mayer RJ, Stone RM, et al, and Cancer and Leukemia Group B 8461. Pretreatment cytogenetics add to other prognostic factors predicting complete remission and long-term outcome in patients 60 years of age or older with acute myeloid leukemia: results from Cancer and Leukemia Group B 8461. Blood. 2006; 108:63–73. https://doi.org/10.1182/blood-2005-11-4354 PMID:<u>16522815</u>
- 59. You X, Conrad TO. Acfs: accurate circRNA identification and quantification from RNA-Seq data. Sci Rep. 2016; 6:38820. https://doi.org/10.1038/srep38820 PMID:27929140
- Bakst RL, Tallman MS, Douer D, Yahalom J. How I treat extramedullary acute myeloid leukemia. Blood. 2011; 118:3785–93. <u>https://doi.org/10.1182/blood-2011-04-347229</u> PMID:<u>21795742</u>
- Lv C, Sun L, Guo Z, Li H, Kong D, Xu B, Lin L, Liu T, Guo D, Zhou J, Li Y. Circular RNA regulatory network reveals cell-cell crosstalk in acute myeloid leukemia extramedullary infiltration. J Transl Med. 2018; 16:361. <u>https://doi.org/10.1186/s12967-018-1726-x</u> PMID:<u>30558617</u>
- 62. Li W, Zhong C, Jiao J, Li P, Cui B, Ji C, Ma D. Characterization of hsa\_circ\_0004277 as a New Biomarker for Acute Myeloid Leukemia via Circular RNA Profile and Bioinformatics Analysis. Int J Mol Sci. 2017; 18:597. <u>https://doi.org/10.3390/ijms18030597</u> PMID:28282919
- Ping L, Jian-Jun C, Chu-Shu L, Guang-Hua L, Ming Z. Silencing of circ\_0009910 inhibits acute myeloid leukemia cell growth through increasing miR-20a-5p. Blood Cells Mol Dis. 2019; 75:41–47. <u>https://doi.org/10.1016/j.bcmd.2018.12.006</u> PMID:30612066
- 64. Shang J, Chen WM, Wang ZH, Wei TN, Chen ZZ, Wu WB. CircPAN3 mediates drug resistance in acute myeloid leukemia through the miR-153-5p/miR-183-5p-XIAP axis. Exp Hematol. 2019; 70:42–54.e3. https://doi.org/10.1016/j.exphem.2018.10.011 PMID:30395908
- 65. Hunger SP, Lu X, Devidas M, Camitta BM, Gaynon PS, Winick NJ, Reaman GH, Carroll WL. Improved

survival for children and adolescents with acute lymphoblastic leukemia between 1990 and 2005: a report from the children's oncology group. J Clin Oncol. 2012; 30:1663–69.

https://doi.org/10.1200/JCO.2011.37.8018 PMID:22412151

- 66. Mullighan CG, Phillips LA, Su X, Ma J, Miller CB, Shurtleff SA, Downing JR. Genomic analysis of the clonal origins of relapsed acute lymphoblastic leukemia. Science. 2008; 322:1377–80. <u>https://doi.org/10.1126/science.1164266</u> PMID:<u>19039135</u>
- 67. Chen J, Li Y, Zheng Q, Bao C, He J, Chen B, Lyu D, Zheng B, Xu Y, Long Z, Zhou Y, Zhu H, Wang Y, et al. Circular RNA profile identifies circPVT1 as a proliferative factor and prognostic marker in gastric cancer. Cancer Lett. 2017; 388:208–19. <u>https://doi.org/10.1016/j.canlet.2016.12.006</u> PMID:<u>27986464</u>
- Qin S, Zhao Y, Lim G, Lin H, Zhang X, Zhang X. Circular RNA PVT1 acts as a competing endogenous RNA for miR-497 in promoting non-small cell lung cancer progression. Biomed Pharmacother. 2019; 111:244– 50. <u>https://doi.org/10.1016/j.biopha.2018.12.007</u> PMID:<u>30590312</u>
- Kun-Peng Z, Xiao-Long M, Chun-Lin Z. Overexpressed circPVT1, a potential new circular RNA biomarker, contributes to doxorubicin and cisplatin resistance of osteosarcoma cells by regulating ABCB1. Int J Biol Sci. 2018; 14:321–30. https://doi.org/10.7150/ijbs.24360 PMID:29559849
- Hu J, Han Q, Gu Y, Ma J, McGrath M, Qiao F, Chen B, Song C, Ge Z. Circular RNA PVT1 expression and its roles in acute lymphoblastic leukemia. Epigenomics. 2018; 10:723–32. <u>https://doi.org/10.2217/epi-2017-0142</u> PMID:<u>29693417</u>
- 71. Goldman JM, Melo JV. Chronic myeloid leukemia-advances in biology and new approaches to treatment. N Engl J Med. 2003; 349:1451–64. <u>https://doi.org/10.1056/NEJMra020777</u> PMID:<u>14534339</u>
- 72. Mauro MJ. Defining and managing imatinib resistance. Hematology (Am Soc Hematol Educ Program). 2006; 2006:219–25. <u>https://doi.org/10.1182/asheducation-2006.1.219</u> PMID:<u>17124064</u>
- Pan Y, Lou J, Wang H, An N, Chen H, Zhang Q, Du X. CircBA9.3 supports the survival of leukaemic cells by up-regulating c-ABL1 or BCR-ABL1 protein levels. Blood Cells Mol Dis. 2018; 73:38–44. <u>https://doi.org/10.1016/j.bcmd.2018.09.002</u> PMID:<u>30224298</u>

- Hallek M. Chronic lymphocytic leukemia: 2017 update on diagnosis, risk stratification, and treatment. Am J Hematol. 2017; 92:946–65. <u>https://doi.org/10.1002/ajh.24826</u> PMID:<u>28782884</u>
- 75. Chiorazzi N, Rai KR, Ferrarini M. Chronic lymphocytic leukemia. N Engl J Med. 2005; 352:804–15. <u>https://doi.org/10.1056/NEJMra041720</u> PMID:<u>15728813</u>
- 76. Dahl M, Daugaard I, Andersen MS, Hansen TB, Grønbæk K, Kjems J, Kristensen LS. Enzyme-free digital counting of endogenous circular RNA molecules in B-cell malignancies. Lab Invest. 2018; 98:1657–69. <u>https://doi.org/10.1038/s41374-018-0108-6</u> PMID:<u>30087459</u>
- 77. Xia L, Wu L, Bao J, Li Q, Chen X, Xia H, Xia R. Circular RNA circ-CBFB promotes proliferation and inhibits apoptosis in chronic lymphocytic leukemia through regulating miR-607/FZD3/Wnt/β-catenin pathway. Biochem Biophys Res Commun. 2018; 503:385–90. <u>https://doi.org/10.1016/j.bbrc.2018.06.045</u> PMID:<u>29902450</u>
- 78. Wu W, Wu Z, Xia Y, Qin S, Li Y, Wu J, Liang J, Wang L, Zhu H, Fan L, Fu J, Xu W, Jin H, Li J. Downregulation of circ\_0132266 in chronic lymphocytic leukemia promoted cell viability through miR-337-3p/PML axis. Aging (Albany NY). 2019; 11:3561–73. <u>https://doi.org/10.18632/aging.101997</u> PMID:<u>31152142</u>
- 79. Pan BT, Teng K, Wu C, Adam M, Johnstone RM. Electron microscopic evidence for externalization of the transferrin receptor in vesicular form in sheep reticulocytes. J Cell Biol. 1985; 101:942–48. <u>https://doi.org/10.1083/jcb.101.3.942</u> PMID:2993317
- 80. van der Pol E, Böing AN, Harrison P, Sturk A, Nieuwland R. Classification, functions, and clinical relevance of extracellular vesicles. Pharmacol Rev. 2012; 64:676–705. <u>https://doi.org/10.1124/pr.112.005983</u> PMID:22722893
- Szczepanski MJ, Szajnik M, Welsh A, Whiteside TL, Boyiadzis M. Blast-derived microvesicles in sera from patients with acute myeloid leukemia suppress natural killer cell function via membrane-associated transforming growth factor-beta1. Haematologica. 2011; 96:1302–09. <u>https://doi.org/10.3324/haematol.2010.039743</u> PMID:21606166
- 82. Théry C, Zitvogel L, Amigorena S. Exosomes: composition, biogenesis and function. Nat Rev Immunol. 2002; 2:569–79. <u>https://doi.org/10.1038/nri855</u> PMID:<u>12154376</u>

- 83. Becker A, Thakur BK, Weiss JM, Kim HS, Peinado H, Lyden D. Extracellular Vesicles in Cancer: Cell-to-Cell Mediators of Metastasis. Cancer Cell. 2016; 30:836– 48. <u>https://doi.org/10.1016/j.ccell.2016.10.009</u> PMID:<u>27960084</u>
- Kumar B, Garcia M, Murakami JL, Chen CC. Exosomemediated microenvironment dysregulation in leukemia. Biochim Biophys Acta. 2016; 1863:464–70. <u>https://doi.org/10.1016/j.bbamcr.2015.09.017</u>
  PMID:<u>26384870</u>
- 85. Corrado C, Raimondo S, Saieva L, Flugy AM, De Leo G, Alessandro R. Exosome-mediated crosstalk between chronic myelogenous leukemia cells and human bone marrow stromal cells triggers an interleukin 8-dependent survival of leukemia cells. Cancer Lett. 2014; 348:71–76. https://doi.org/10.1016/j.canlet.2014.03.009 PMID:24657661
- 86. Li Y, Zheng Q, Bao C, Li S, Guo W, Zhao J, Chen D, Gu J, He X, Huang S. Circular RNA is enriched and stable in exosomes: a promising biomarker for cancer diagnosis. Cell Res. 2015; 25:981–84. https://doi.org/10.1038/cr.2015.82 PMID:26138677
- 87. Bao C, Lyu D, Huang S. Circular RNA expands its territory. Mol Cell Oncol. 2015; 3:e1084443. <u>https://doi.org/10.1080/23723556.2015.1084443</u> PMID:<u>27308606</u>

- Dou Y, Cha DJ, Franklin JL, Higginbotham JN, Jeppesen DK, Weaver AM, Prasad N, Levy S, Coffey RJ, Patton JG, Zhang B. Circular RNAs are downregulated in KRAS mutant colon cancer cells and can be transferred to exosomes. Sci Rep. 2016; 6:37982. <u>https://doi.org/10.1038/srep37982</u> PMID:<u>27892494</u>
- Lasda E, Parker R. Circular RNAs Co-Precipitate with Extracellular Vesicles: A Possible Mechanism for circRNA Clearance. PLoS One. 2016; 11:e0148407. <u>https://doi.org/10.1371/journal.pone.0148407</u> PMID:26848835
- 90. Tan S, Gou Q, Pu W, Guo C, Yang Y, Wu K, Liu Y, Liu L, Wei YQ, Peng Y. Circular RNA F-circEA produced from EML4-ALK fusion gene as a novel liquid biopsy biomarker for non-small cell lung cancer. Cell Res. 2018; 28:693–95. <u>https://doi.org/10.1038/s41422-018-0033-7</u> PMID:<u>29628502</u>
- 91. Jost I, Shalamova LA, Gerresheim GK, Niepmann M, Bindereif A, Rossbach O. Functional sequestration of microRNA-122 from Hepatitis C Virus by circular RNA sponges. RNA Biol. 2018; 15:1032–39. <u>https://doi.org/10.1080/15476286.2018.1435248</u> PMID:<u>29486652</u>