

Phylogenetic ubiquity of the effects of altered ubiquinone biosynthesis on survival

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The widespread use of unicellular and invertebrate model systems has revealed that the molecular mechanisms underlying cellular functions are exceedingly well conserved. The expanding research into the molecular mechanisms of aging keeps yielding this answer too [1,2]. A perfect example of this is a study by Gonidakis, Finkel and Longo appearing in the present issue of *Aging*, which shows that a disruption of the biosynthesis of ubiquinone leads to increased survival of *E. coli* during stationary phase, associated with an increased resistance to treatments that induce oxidative stress [3]. These effects are dependent upon the presence of an intact *arcA* gene, which encodes the regulatory component of the ArcA/ArcB system, a hypoxia-inducible system of transcriptional regulation.

To understand why these findings might be an example of evolutionary conservation a little background is needed. Interventions that disrupt mitochondrial function can increase lifespan in a variety of organisms including yeast [4], *Caenorhabditis elegans* [5-10], *Drosophila* [11], and mice [12-14]. One type of intervention is the disruption of mitochondrial function through the reduction of the expression of mitochondrial genes by RNAi, which increases lifespan in worms [5,10] and in flies [11], possibly via a mitochondria-specific stress response [9]. Another type of intervention is a specific alteration of mitochondrial electron transport that alters the generation of reactive oxygen species (ROS) [15,16]. For example, *C. elegans isp-1* and *nuo-6* mutants, which carry point mutations in subunits of the mitochondrial electron transport chain (ETC) display an elevated generation of mitochondrial superoxide which appears to be causal to their increased

lifespan. Indeed, antioxidants suppress the mutants' longevity and pro-oxidant treatment of the wild type phenocopies it [16]. *clk-1* (called *Mcl1* in mice) is another gene that has been studied in this context. It encodes a mitochondrial hydroxylase that is necessary for the biosynthesis of ubiquinone [6,17,18]. Ubiquinone (a.k.a. co-enzyme Q) is an electron transporter and antioxidant that is ubiquitous in the membranes of all organisms [19]. *C. elegans clk-1* mutants [20] and mouse *Mcl1*^{+/-} mutants [12,13] are long-lived and have been shown to have elevated generation of mitochondrial ROS [16,21]. In cultured vertebrate cells mitochondrial ROS have been shown to help stabilize and thus induce the protective activity of the hypoxia-inducible factor 1 α (HIF-1 α) [22-24]. It is striking therefore that HIF-1 α has been tentatively implicated in the mechanisms of longevity of both *C. elegans clk-1* [25] and mouse *Mcl1*^{+/-} mutants [26].

Of course bacteria have no mitochondria; however, as the evolutionary ancestors of mitochondria, they have a plasma membrane ETC partly homologous to that of the organelle. As in mitochondria, the bacterial ETC appears to produce significant amounts of ROS [27]. Like eukaryotic cells, they have transcription factors sensitive to hypoxia. The ArcA/ArcB two-component system is one of the key pathways up-regulated in response to anaerobic conditions. Although not genetically homologous to eukaryotic HIF-1 α , there are interesting parallels between the systems. While activated by the redox state of the bacterial quinone pool rather than by ROS [28-30], Arc activation is required for the resistance of *E. coli* to induced oxidative stress [31]. Thus, the work of Longo and co-

workers in *E. coli* suggests that there might be a truly universal link between ubiquinone, ROS generation, hypoxia-sensitive transcription factors and cellular survival.

REFERENCES

1. Blagosklonny MV and Hall MN. Growth and aging: a common molecular mechanism. *Aging* (Albany NY). 2009; 1(4): 357-362.
2. Piper MDW, Selman C, McElwee JJ, Partridge L. Separating cause from effect: how does insulin/IGF signalling control lifespan in worms, flies and mice? *Journal of Internal Medicine*. 2008; 263(2): 179-191.
3. Gonidakis S, Finkel SE, Longo VD. Lifespan extension and paraquat resistance in a ubiquinone-deficient *Escherichia coli* mutant depend on transcription factors ArcA and TdcA. *Aging*. 2011.
4. Kirchman PA, Kim S, Lai CY, Jazwinski SM. Interorganelle signaling is a determinant of longevity in *Saccharomyces cerevisiae*. *Genetics*. 1999; 152(1): 179-190.
5. Lee SS, Lee RY, Fraser AG, Kamath RS, Ahringer J, Ruvkun G. A systematic RNAi screen identifies a critical role for mitochondria in *C. elegans* longevity. *Nat Genet*. 2003; 33(1): 40-48.
6. Ewbank JJ, Barnes TM, Lakowski B, Lussier M, Bussey H, Hekimi S. Structural and functional conservation of the *Caenorhabditis elegans* timing gene *clk-1*. *Science*. 1997; 275(5302): 980-983.
7. Felkai S, Ewbank JJ, Lemieux J, Labbe JC, Brown GG, Hekimi S. CLK-1 controls respiration, behavior and aging in the nematode *Caenorhabditis elegans*. *Embo J*. 1999; 18(7): 1783-1792.
8. Feng J, Bussiere F, Hekimi S. Mitochondrial electron transport is a key determinant of life span in *Caenorhabditis elegans*. *Dev Cell*. 2001; 1(5): 633-644.
9. Yang W, Hekimi S. Two modes of mitochondrial dysfunction lead independently to lifespan extension in *Caenorhabditis elegans*. *Aging Cell*. 2010; 9(3): 433-447.
10. Dillin A, Hsu AL, Arantes-Oliveira N, Lehrer-Graiwer J, Hsin H, Fraser AG, Kamath RS, Ahringer J, Kenyon C. Rates of behavior and aging specified by mitochondrial function during development. *Science*. 2002; 298(5602): 2398-2401.
11. Copeland JM, Cho J, Lo T, Hur JH, Bahadorani S, Arabyan T, Rabie J, Soh J, Walker DW. Extension of *Drosophila* Life Span by RNAi of the Mitochondrial Respiratory Chain. *Current Biology*. 2009; 19(19): 1591-1598.
12. Lapointe J, Stepanyan Z, Bigras E, Hekimi S. Reversal of the mitochondrial phenotype and slow development of oxidative biomarkers of aging in long-lived *Mcl1*^{+/+} mice. *J Biol Chem*. 2009; 284(30): 20364-20374.
13. Liu X, Jiang N, Hughes B, Bigras E, Shoubridge E, Hekimi S. Evolutionary conservation of the *clk-1*-dependent mechanism of longevity: loss of *mcl1* increases cellular fitness and lifespan in mice. *Genes Dev*. 2005; 19(20): 2424-2434.
14. Dell'agnello C, Leo S, Agostino A, Szabadkai G, Tiveron C, Zulian A, Prella A, Roubertoux P, Rizzuto R, Zeviani M. Increased longevity and refractoriness to Ca²⁺-dependent neurodegeneration in Surf1 knockout mice. *Hum Mol Genet*. 2007; 16(4): 431-444.
15. Van Raamsdonk JM, Hekimi S. Deletion of the mitochondrial superoxide dismutase *sod-2* extends lifespan in *Caenorhabditis elegans*. *PLoS Genet*. 2009; 5(2): e1000361.
16. Yang W, Hekimi S. A mitochondrial superoxide signal triggers increased longevity in *Caenorhabditis elegans*. *PLoS Biol*. 2010; 8(12): e1000556.
17. Miyadera H, Amino H, Hiraishi A, Taka H, Murayama K, Miyoshi H, Sakamoto K, Ishii N, Hekimi S, Kita K. Altered quinone biosynthesis in the long-lived *clk-1* mutants of *Caenorhabditis elegans*. *J Biol Chem*. 2001; 276(11): 7713-7716.
18. Jonassen T, Larsen PL, Clarke CF. A dietary source of coenzyme Q is essential for growth of long-lived *Caenorhabditis elegans* *clk-1* mutants. *Proc Natl Acad Sci U S A*. 2001; 98(2): 421-426.
19. Turunen M, Olsson J, Dallner G. Metabolism and function of coenzyme Q. *Biochim Biophys Acta*. 2004; 1660(1-2): 171-199.
20. Wong A, Boutis P, Hekimi S. Mutations in the *clk-1* gene of *Caenorhabditis elegans* affect developmental and behavioral timing. *Genetics*. 1995; 139(3): 1247-1259.
21. Lapointe J, Hekimi S. Early mitochondrial dysfunction in long-lived *Mcl1*^{+/+} mice. *J Biol Chem*. 2008; 283(38): 26217-26227.
22. Bell EL, Klimova TA, Eisenbart J, Moraes CT, Murphy MP, Budinger GR, Chandel NS. The Qo site of the mitochondrial complex III is required for the transduction of hypoxic signaling via reactive oxygen species production. *J Cell Biol*. 2007; 177(6): 1029-1036.
23. Brunelle JK, Bell EL, Quesada NM, Vercauteren K, Tiranti V, Zeviani M, Scarpulla RC, Chandel NS. Oxygen sensing requires mitochondrial ROS but not oxidative phosphorylation. *Cell Metabolism*. 2005; 1(6): 409-414.
24. Guzy RD, Hoyos B, Robin E, Chen H, Liu L, Mansfield KD, Simon MC, Hammerling U, Schumacker PT. Mitochondrial complex III is required for hypoxia-induced ROS production and cellular oxygen sensing. *Cell Metab*. 2005; 1(6): 401-408.
25. Lee SJ, Hwang AB, Kenyon C. Inhibition of Respiration Extends *C. elegans* Life Span via Reactive Oxygen Species that Increase HIF-1 Activity. *Curr Biol*. 2010; 20(23): 2131-2136.
26. Wang D, Malo D, Hekimi S. Elevated mitochondrial reactive oxygen species generation affects the immune response via hypoxia-inducible factor-1 α in long-lived *Mcl1*^{+/+} mouse mutants. *J Immunol*. 2010; 184(2): 582-590.
27. Gonzalez-Flecha B, Demple B. Metabolic sources of hydrogen peroxide in aerobically growing *Escherichia coli*. *J Biol Chem*. 1995; 270(23): 13681-13687.
28. Bekker M, Alexeeva S, Laan W, Sawers G, Teixeira de Mattos J, Hellingwerf K. The ArcBA two-component system of *Escherichia coli* is regulated by the redox state of both the ubiquinone and the menaquinone pool. *J Bacteriol*. 2010; 192(3): 746-754.
29. Georgellis D, Kwon O, Lin EC. Quinones as the redox signal for the arc two-component system of bacteria. *Science*. 2001; 292(5525): f 2314-2316.
30. Malpica R, Franco B, Rodriguez C, Kwon O, Georgellis D. Identification of a quinone-sensitive redox switch in the ArcB sensor kinase. *Proceedings of the National Academy of Sciences of the United States of America*. 2004; 101(36): 13318-13323.
31. Loui C, Chang A, Lu S. Role of the ArcAB two-component system in the resistance of *Escherichia coli* to reactive oxygen stress. *BMC Microbiology*. 2009; 9(1): 183.