

Impact papers on aging in 2009

Mikhail V. Blagosklonny¹, Judy Campisi², David A. Sinclair³, Andrzej Bartke⁴, Maria A. Blasco⁵, William M. Bonner⁶, Vilhelm A. Bohr⁷, Robert M. Brosh Jr⁷, Anne Brunet⁸, Ronald A. DePinho⁹, Lawrence A. Donehower¹⁰, Caleb E. Finch¹¹, Toren Finkel¹², Myriam Gorospe⁷, Andrei V. Gudkov¹, Michael N. Hall¹³, Siegfried Hekimi¹⁴, Stephen L. Helfand¹⁵, Jan Karlseder¹⁶, Cynthia Kenyon¹⁷, Guido Kroemer¹⁸, Valter Longo¹¹, Andre Nussenzweig⁶, Heinz D. Osiewacz¹⁹, Daniel S. Peeper²⁰, Thomas A. Rando⁸, K Lenhard Rudolph²¹, Paolo Sassone-Corsi²², Manuel Serrano⁵, Norman E. Sharpless²³, Vladimir P. Skulachev²⁴, Jonathan L. Tilly²⁵, John Tower¹¹, Eric Verdin¹⁷, Jan Vijg²⁶

¹ Roswell Park Cancer Institute, Buffalo, NY, USA, ² Buck Institute for Age Research, Novato, CA, USA

³ Harvard Medical School, Boston, MA, USA, ⁴ Southern Illinois University, Springfield, IL, USA

⁵ Spanish National Cancer Center, Madrid, Spain, ⁶ National Cancer Institute, NIH, Bethesda, MD, USA

⁷ National Institute on Aging, NIH, Baltimore, MD, USA, ⁸ Stanford University, Stanford, CA, USA

⁹ Dana-Farber Cancer Institute, Boston, MA, USA, ¹⁰ Baylor College of Medicine, Houston, TX, USA

¹¹ University of Southern California, Los Angeles, CA USA, ¹² NHLBI, NIH, Bethesda, MD, USA

¹³ University of Basel, Basel, Switzerland, ¹⁴ McGill University, Montreal, Canada

¹⁵ Brown University, Providence, RI, USA, ¹⁶ The Salk Institute, La Jolla, CA, USA

¹⁷ University of California, San Francisco, CA, USA, ¹⁸ INSERM, U848, Villejuif, France

¹⁹ Goethe University Frankfurt, Frankfurt, Germany, ²⁰ The Netherlands Cancer Institute, Amsterdam, Netherlands

²¹ Medical School Hannover, Hannover, Germany, ²² University of California, Irvine, CA, USA

²³ University of North Carolina, Chapel Hill, NC, USA, ²⁴ Moscow State University, Moscow, Russia

²⁵ Massachusetts General Hospital, Boston, MA, USA, ²⁶ Albert Einstein College of Medicine, Bronx, USA

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Correspondence: Blagosklonny, MD/PhD, MD, PhD, Professor, Department of Cell Stress Biology, Roswell Park Cancer Institute, BLSC, L3-312, Elm and Carlton Streets, Buffalo, NY 14263, USA

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E-mail: Blagosklonny@oncotarget.com

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Abstract

The editorial board of *Aging* reviews research papers published in 2009, which they believe have or will have a significant impact on aging research. Among many others, the topics include genes that accelerate aging or in contrast promote longevity in model organisms, DNA damage responses and telomeres, molecular mechanisms of life span extension by calorie restriction and pharmacologic interventions into aging. The emerging message in 2009 is that aging is not random but determined by a genetically-regulated longevity network and can be decelerated both genetically and pharmacologically.

Telomeres

The 2009 Nobel Prize in Physiology or Medicine was awarded to Elizabeth Blackburn, Carol Greider and Jack Szostak for their contributions to our understanding of how the ends of eukaryotic chromosomes, telomeres, are maintained by a specialized reverse transcriptase, telomerase. This award is the closest Nobel Prize to date related to aging. Of course, the major significance of the work relates to basic cell biology and cancer, rather than aging research. In fact, whereas telomere shortening explains the Hayflick limit (replicative senescence) in human cells, it cannot explain the difference in longevity between mice and men. But there may be other links between telomeres and aging. In 2009, several publications by Epel, Blackburn and co-workers provide a new link between telomere length and age-related diseases. As published in the first issue of *Aging*, the rate of telomere shortening in peripheral leukocytes predicts mortality from cardiovascular disease in elderly men [1]. Even more intriguingly, pessimism correlates with short leukocyte telomeres and elevated interleukin (IL)-6 in post-menopausal women [2]. The cause-and-effect relationship between telomere length and these physiological endpoints is unknown, but several non-mutually exclusive explanations can be proposed. Rapid telomere shortening may indicate a cellular hyper-activation, hyper-proliferation and/or hyper-secretory phenotypes often associated with cellular senescence, stem cell exhaustion and diseases of aging.

In agreement with these possibilities, telomere length was shown to regulate the expression of interferon-stimulated gene 15 (ISG15). Short-telomeres up-regulated ISG15 independent of DNA damage signaling. This finding demonstrated for the first time that an endogenous human gene can be regulated by telomere length prior to the onset of telomere dysfunction and DNA damage signals. It was suggested that the upregulation of ISG15 by telomere shortening may contribute to the chronic inflammation associated with human aging [3]. Pertinent to this idea, also in 2009, the secretion of inflammatory cytokines such as IL-6 and IL-8 by senescent cells, whether made senescent by dysfunctional telomeres or DNA damage, was shown to be suppressed by two micro-RNAs (miR-146a and 146b) [4]. It was proposed that these micro-RNAs modulate inflammatory responses by affecting signal transduction pathways that contribute to a larger senescence associated secretory phenotype. It will be of interest to know whether miR-146a/b also suppresses ISG15 expression, and if this effect is influenced by telomere status.

It was also demonstrated that dysfunction of a telomere-binding protein is sufficient to produce severe telomeric

damage in the absence of telomere shortening, resulting in premature tissue degeneration and development of neoplastic lesions [5]. New insight has been gained in the understanding of how telomeres are maintained and how the processes of DNA repair occur in telomeres. For example, it appears that the guardians of the genome, the RecQ helicases, actively participate in this repair process [6].

Damaged telomeres were also found to be the major factor contributing to the wide variability in the amount of DNA damage signaling in human tumor cell lines, findings that may help clarify the relative contributions of non-telomeric DNA double-strand breaks and damaged telomeres to levels of genomic instability [7].

DNA damage response and aging

In 2009, it was demonstrated that the persistent (but not transient) DNA damage response (DDR) associated with senescent cells is essential for their ability to express and secrete inflammatory cytokines [8]. Cell surface-bound IL-1alpha is essential for the senescence-associated secretion of IL-6 and IL-8, 2 proinflammatory cytokines, reinforcing the senescence phenotype [9].

Both the initiation and maintenance of cytokine secretion required the DDR proteins ATM, NBS1 and CHK2, but not p53. ATM was also essential for IL-6 secretion during oncogene-induced senescence and by damaged cells that bypass senescence. It was proposed that this activity of the DDR allows senescent cells to communicate their compromised state to the surrounding tissue [8]. In addition, a DDR may occur in senescent cells even in the absence of detectable DNA damage [10]. This pseudo-DDR is a marker of cellular hyperactivation and is inhibited by rapamycin [10], a clinically approved drug that decelerates cellular senescence [11]. Thus, persistent DDR signaling, regardless of DNA damage, may be a part of the senescent phenotype.

It was shown that longevity extension mutations in the yeast SCH9, the yeast homolog of the conserved pro-aging gene S6K (Ribosomal Protein S6 Kinase), caused a major reduction in age-dependent DNA damage by lowering the activity of error-prone DNA repair genes [12].

Also, age-dependent deterioration of nuclear pore complexes causes an increase in nuclear permeability and the leaking of cytoplasmic proteins into the nucleus in postmitotic cells [13].

The ability to respond to stress decreases with age. Stress-responding factors which regulate transcription can influence longevity. In 2009, Westerheide et al demonstrated that stress-induced regulation of heat shock factor 1 (HSF-1) by the deacetylase SIRT1 (sirtuin 1) may play a role in the regulation of life span [14]. Defining the targets of sirtuins may help to understand the importance of transcriptional regulation in age-related diseases.

An intriguing possibility is that the response of the cells to certain types of DNA damage (e.g. DNA breaks) results in epigenetic changes that alter gene expression [15]. These changes do occur in mammals and it will be interesting to test whether these epigenetic changes in response to DNA damage are associated with, or can actually cause aging.

Mitochondria, oxidative stress and aging

On the other hand, the free radical theory, which posits that aging is caused by an *accumulation* of oxidative damage, was critically questioned in 2009. First, overexpression of major antioxidant enzymes, which decrease free radicals, did not extend the lifespan of mice [16]. Second, deletion of mitochondrial superoxide dismutase (Sod-2) extended life span in *Caenorhabditis elegans* [17]. Third, life span extension by dietary restriction was not linked to protection against somatic DNA damage in *Drosophila melanogaster* [18]. Fourth, Sod-2 haploinsufficiency did not accelerate murine aging, even in mice with dysfunctional telomeres [19]. In addition it was demonstrated that the reduced energy metabolism and the increased oxidative stress in the mitochondria of young Mcl1^{+/-} mice results in an almost complete protection from the age-dependent loss of mitochondrial function. Moreover, this altered mitochondrial condition is linked to a significant attenuation of the rate of development of oxidative biomarkers of aging. Thus, this study indicates that mitochondrial oxidative stress is not causal to aging [20]. It was reported that RNAi of five genes encoding components of mitochondrial respiratory complexes I, III, IV, and V leads to increased life span in flies. Long-lived flies with reduced expression of electron transport chain (ETC) genes do not consistently show reduced assembly of respiratory complexes or reduced ATP levels. In addition, extended longevity is not consistently correlated with increased resistance to the free-radical generator paraquat [21].

These results are in agreement with previous papers showing that antioxidants overexpression causes minor effects in life span extension in yeast, flies, and mice

compared to those caused by mutations in signal transduction genes. It is likely that increase protection against superoxide must be accompanied by a number of other changes to be effective in life span extension. For instance, LON, a AAA protease located in the mitochondrial matrix, increases stress tolerance, mitochondrial oxygen consumption, while decreasing oxidative damage of proteins in the fungal aging model *Podospora anserina* [22]. In the same model organism, deletion of a gene encoding a O-methyltransferase, which decrease levels of reactive oxygen species, leads to a decreased lifespan [23].

Calorie restriction (CR)

Caloric restriction (CR) without malnutrition delays aging and extends life span in diverse species; however, its effect in primates had not been clearly established. In 2009, a 20-year longitudinal study of adult-onset CR in rhesus monkeys demonstrated that moderate CR lowered the incidence of aging-related deaths. At the time point reported, 50% of control animals had survived, compared with 80% of CR animals. CR delayed the onset of several age-associated pathologies such as diabetes, cancer, cardiovascular disease and brain atrophy [24]. The CR trial in primates raised hope that CR might be effective in humans.

In 2009, numerous studies continued to establish links between caloric restriction (CR) and longevity signaling pathways, including Sir2 (sirtuin) and p53 in *D. melanogaster* [25] and the E3 ubiquitin ligase WWP-1 in *C. elegans* [26] as well as upstream and downstream components of the TOR (Target of Rapamycin) pathway: RHEB-1 in *C. elegans* [27], Tor1 and Sch9 (a homolog of the mammalian kinases Akt and S6K) in yeast [28], and 4E-BP (Eukaryotic Translation Initiation Factor 4E Binding Protein) in *Drosophila* [29]. It was shown that glucose shortens the life span of *C. elegans* by downregulating DAF-16/FOXO activity and aquaporin gene expression [30]. In addition, the HIF (hypoxia inducible factor) pathway was implicated in aging and longevity in *C. elegans* [31, 32]. The different results of two studies have been in general reconciled [33]. In 2009, it has also been shown that in *C. elegans* CR is mediated by a network of independent, but overlapping pathways [34], suggesting a 'CR network'. Notably, neuronal SIRT1 regulated endocrine and behavioral responses to CR [35].

It has been shown that disruption of growth hormone receptor (GHR) prevents calorie restriction from improving insulin action and longevity [36]. In normal mice, CR increased insulin sensitivity in liver and muscle. In GHRKO mice, intrinsic insulin-sensitivity

could be attributed to a reduction of inhibitory serine phosphorylation of IRS-1 (Insulin receptor substrate 1) in muscle. CR failed to further increase insulin signaling (insulin sensitivity) in GHRKO mice as compared to normal mice, likely explaining the absence of CR effects on longevity in these long-lived mice [36].

Finally, it was tested whether reallocation of nutrients from reproduction to somatic maintenance could explain the life extending effect of CR. If this were the case, long life under dietary restriction and high fecundity (reproduction) under full feeding would be mutually exclusive. Adding methionine alone to the dietary restriction condition was necessary and sufficient to increase fecundity as much as did full feeding, but without reducing lifespan. Reallocation of nutrients therefore does not explain the responses to dietary restriction. In contrast, reduced activity of the insulin/insulin-like growth factor signaling protected against the shortening of lifespan with full feeding [37].

Pharmacologic intervention

The ultimate goal of biomedical research is the development of therapeutic drugs. As shown previously, activation of mTOR (mammalian Target of Rapamycin) is required for acquiring senescent phenotype in p21-arrested human cells, whereas deactivation of mTOR converts senescence into quiescence. In 2009, it was further demonstrated that the inhibitor of mTOR rapamycin decelerated cellular senescence of p21-arrested human and mouse cells [11]. Similarly, inhibitors of PI-3K and MEK, LY-294002 and U0126, deactivated mTOR and suppressed cellular senescence (converting it into quiescence) [38], defining direct and indirect mTOR inhibitors as aging-suppressants or geropressants.

The most striking event of the year was the demonstration that rapamycin, administered to middle-aged (600 day old) mice, significantly extended their life span [39]. The effect was seen at three independent test sites in genetically heterogeneous mice, chosen to avoid genotype-specific effects on disease susceptibility [39]. Rapamycin also prolonged the life of 22-month old mice [40]. [Note: publications by Bjedov et al (Cell Metab 2010 Jan) and by Moskalev and Shaposhnikov (coming in print 2010) that rapamycin extends life span in *Drosophila* will be reviewed next year].

It was shown that clioquinol, a metal chelator that has beneficial effects in several models of neurodegenerative diseases, inhibits the activity of the mitochondrial enzyme CLK-1 in mammalian cells.

Clioquinol-treated nematodes and mice presented a variety of phenotypes produced by mutational reduction of CLK-1. Given that reduction of CLK-1 slows down aging in these organisms, these results suggest that clioquinol (by inhibiting CLK-1) may slow down the aging process [41].

Finally, as a follow-up of the work on the anti-aging effects of mitochondria-targeted antioxidant SkQ1 [42], it was demonstrated that Sk inhibits age-dependent involution of the thymus in normal and senescence-prone rats [43].

Stem cells and aging

In 2009, several lines of evidence suggested that overactivation of signaling pathways might cause exhaustion of stem cells and that vice versa 'longevity genes' could prevent stem cell exhaustion. Thus, mTOR mediated Wnt-induced epidermal stem cell exhaustion and aging phenotypes in skin [44]. Further, hyper-activation of mTORC1 caused hyperproliferation and subsequent exhaustion of hematopoietic stem cells. Pharmacological approaches showed that PTEN, TSC1 and PML regulate hematopoietic stem cell (HSC) maintenance through mTORC1 [45]. In addition, FOXO transcription factors were found to be necessary for adult neural stem cell homeostasis [46, 47]. Importantly, stem cell aging could be suppressed pharmacologically [40, 44]. The PI3K-AKT-FoxO pathway is integral to lifespan regulation in lower organisms plays a prominent role in neural stem/progenitor cell (NSC) proliferation and renewal. FoxO-deficient mice show initial increased brain size and proliferation of neural progenitor cells during early postnatal life, followed by precocious significant decline in the NSC pool and accompanying neurogenesis in adult brains [46].

In addition, functions of aging organs can be rejuvenated by young supporting stem cells. As published in the first issue of *Aging*, once-monthly infusions of bone marrow (BM)-derived cells from young adult female mice sustained the fertility of aging females long past their time of normal reproductive failure [48]. The fertility-promoting effects were observed regardless of whether the infusions were initiated in young adult or middle-aged females, and were specific for bone marrow harvested from female donors. This "rejuvenation" did not depend on the development of mature eggs from germline cells in the donor marrow, but from host germline cells sustained by the infusions [48, 49]. In fact, very recent studies showed that aged mouse ovaries lacking oocytes retain a rare population of germline stem cells that, when

transplanted into a young host ovarian environment, are able to generate immature oocytes contained within follicles [49]. Thus, reproductive failure with age may be due, at least in part, to deterioration of somatic microenvironments (niches) that support stem cell function.

Nuclear reprogramming and senescence

Much interest has also been devoted in the past year to nuclear reprogramming of differentiated cells into induced pluripotent stem (iPS) cells by using defined factors. Understanding which factors facilitate the reprogramming process is thought to give clues to the process of carcinogenesis. Inversely, nuclear reprogramming could be also envisioned as a “rejuvenation process”. In this regard, p53 and p16^{INK4a} tumor suppressor proteins were shown to be important in limiting reprogramming [50-55]. Activation of p53 was suggested to be more important in murine cells, whereas activation of p16^{INK4a} appeared the predominant barrier in human cells [50].

Of particular importance to the field of regenerative medicine, which will need patient-specific stem cells derived from older patients, is reprogramming efficiency in fibroblasts from aged humans versus young humans. There is an age-associated decline in reprogramming efficiency, which is largely reversed by inactivation of the p16^{INK4a} tumor suppressor gene, whose expression is increased markedly with aging in several human and murine tissues [50, 55]. Along these lines, it was shown that the increased expression of p16^{INK4a} with aging could be measured on human peripheral blood samples, and that an individual’s p16^{INK4a} expression was a good biomarker of their “molecular age” [56]. The same group also provided further understanding of the observed linkage of SNPs near the *CDKN2a/b* locus (which encodes the p16^{INK4a}, p15^{INK4b} and ARF tumor suppressors) with human atherosclerotic disease [57]. Expression of *CDKN2a/b* transcripts is decreased in individuals harboring the risk alleles, suggesting that atherosclerotic disease may result from aberrant, unrestrained proliferation. In this regard, studies on mice overexposing the *CDKN2a/b* locus were found to have delayed aging and extended longevity [58].

Genetics of aging

In 2009, numerous publications extended our knowledge on the role of sirtuins [35], TOR signaling [59, 60], and the stress response factors HSF-1 and DAF-16 [61] in aging. Of particular importance, it was shown that deletion of the gene encoding Ribosomal

Protein S6 Kinase 1 (S6K1) and disruption of PKA extend the life span of mice [62, 63], whereas the gene encoding Eukaryotic Translation Initiation Factor 4E Binding Protein (4E-BP) was shown to be essential for life span extension by CR in *Drosophila* [29]. Moreover, 4E-BP was shown to act downstream of TOR to modulate cardiac aging in *Drosophila* [64]. Finally, SIRT6 was shown to play a critical role in DNA double-strand break repair [65].

In 2009, Kenyon and co-workers further uncovered mechanisms of their previous observations made in 1999 (Hsin and Kenyon, *Nature*, 1999, 399:362-6) that in *C elegans* and *Drosophila* the aging of the soma is influenced by the germline: namely, when germline-stem cells are removed, aging slows and lifespan is increased. In 2009, it was published that a predicted transcription elongation factor, TCER-1, plays a key role in this process [66]. When the germ cells are removed, the levels of TCER-1 rise in somatic tissues. This increase is sufficient to trigger key downstream events, as overexpression of *tcer-1* extends the lifespan of normal animals that have an intact reproductive system. Intriguingly, TCER-1 specifically links the activity of a broadly deployed transcription factor, DAF-16/FOXO, to longevity signals from reproductive tissues [66]. In mice, *Foxo1* integrates insulin signaling with mitochondrial function, and inhibition of *Foxo1* can improve hepatic metabolism during insulin resistance and the metabolic syndrome [67].

A prior work by Willcox et al (*PNAS* 2008, 105: 13987) showed that genetic variation within the FOXO3A gene was strongly associated with human longevity. Long-lived men also presented several additional phenotypes linked to healthy aging, including lower prevalence of cancer and cardiovascular disease, and high physical and cognitive function. Long-lived men also exhibited greater insulin sensitivity associated with homozygosity for the FOXO3A GG genotype. In 2009, confirming the Willcox observation, the flurry of papers showed the association between SNPs in the *FoxO3A* gene and extreme longevity in Japanese, German, American, Italian, and Chinese populations [68-71].

There were intriguing publications on the complex role of p53 in longevity. In *Drosophila melanogaster*, p53 exerted developmental stage-specific and sex-specific effects on adult life span, indicative of sexual antagonistic pleiotropy [72, 73]. Further, an association between single nucleotide polymorphisms (SNPs) in p53 pathway genes and human fertility suggested that p53 regulates the efficiency of human reproduction. These results provide a plausible explanation for

selective pressure to retain some alleles in the p53 pathway, and suggest that such alleles are a good example of antagonistic pleiotropy [74].

Interestingly, SNPs in the p21 gene correlated with longevity in an Italian population [75]. Several papers have highlighted an important role of p53 in tissue fitness through its impact in preventing mobilization of stem cells harboring persistent DNA damage (ie, dysfunctional telomeres) [76, 77]. However, the phenotypic outcome was tissue and context specific. In mouse epidermis deletion of p53 rescued organ maintenance and body fitness of newborn mice with dysfunctional telomeres [76]. In contrast, p53 deletion in the intestinal epithelium accelerated tissue destruction and shortened the lifespan of aging telomere dysfunctional mice [77]. The latter phenotype was associated with aberrant survival chromosomal instable stem cell clones leading to abnormal differentiation and p53-independent apoptosis. The limitation of the survival of chromosomal instable stem cells is likely to represent a key step in the known role of p53 as a tumor suppressor. Also it was shown that the p53 family member, TAp63, is essential for maintenance of epidermal and dermal precursors and that, in its absence, these precursors senesce and skin ages prematurely [78].

Model systems continue to be instrumental in understanding the genetics of longevity. The *WRN* gene defective in the premature aging disorder Werner syndrome encodes a protein with both helicase and exonuclease activities [79]. To dissect its genetic functions, human *WRN* was tested for its ability to rescue *sgs1*-related phenotypes. *WRN* was shown to genetically interact with topoisomerase 3 and restore the slow growth phenotype of *sgs1 top3*. *WRN* helicase but not exonuclease activity was genetically required for restoration of *top3* growth phenotype, demonstrating separation of function of *WRN* catalytic activities. In a *top3* mutant background, DNA unwinding by *WRN* helicase may be deleterious to cell growth and genome homeostasis [80].

In 2009, a few studies delved into the genetics of the insulin-producing pancreatic beta-cell aging in humans and mice [81-83]. A loss of beta-cell replication with aging is a contributor to age-related increase in the incidence of type II diabetes. Prior work had shown that p16^{INK4a} tumor suppressor causes an age-dependent decline in beta-cell replication. In 2009, it was reported that loss of Polycomb (PcG) repression of p16^{INK4a} mediated by the EZH2 histone methyltransferase occurred with aging in humans and mice [82]. In mice, somatic deletion of EZH2 led to loss of beta-cell

replication and diabetes, and these effects could be rescued by concomitant deletion of p16^{INK4a} and Arf.

This work linked alterations of chromatin architecture with aging to expression of anti-proliferative molecules. Bhushan and colleagues also reported a similar regulation of p16^{INK4a} expression with aging by the Bmi-1 PcG protein, which functions in concert with EZH2 to repress p16^{INK4a} expression [81]. Lastly, it was shown that p38MAPK activates p16^{INK4a} with aging in beta-cells, suggesting a possible pharmacologic approach to regulating aging of this tissue [83].

Autophagy

In 2009, the simple dogma that autophagy is always associated with or causes senescence was challenged. Although autophagy remains a crucial anti-aging mechanism, the relationship is likely to be complex. Thus, autophagy was shown to be activated during cellular senescence, and activation correlated with negative feedback in the PI3K-mTOR pathway. A subset of autophagy-related genes was up-regulated during senescence: overexpression of one gene, ULK3, induced autophagy and senescence. Furthermore, inhibition of autophagy delayed the senescence phenotype, including senescence-associated secretion. These data suggest that autophagy, and its consequent protein turnover, may mediate acquisition of the senescence phenotype [84]. Inhibition of autophagy in adult *Drosophila* [85] or *C. elegans* [86] was found not to affect longevity, however autophagy was required for the increased life span caused by several pharmacologic and genetic manipulations in yeast, *Drosophila* and *C. elegans* [87-90], suggesting that autophagy may be limiting for life span under some conditions but not others. Interestingly, resveratrol-mediated inhibition of mammalian S6 kinase by resveratrol suppressed autophagy [91]. In 2009, several reports further demonstrated that the TOR signaling pathway targets the Atg1/Atg13 protein kinase complex to control autophagy [92-94]. Furthermore, TOR-mediated autophagy regulates cell death in *Drosophila* neurodegenerative disease [95].

The natural polyanion spermidine can extend the chronological and replicative life span in yeast and increase the median and maximal longevity of fruit flies and nematodes (*C. elegans*). Spermidine was found to act as a potent inducer of autophagy in all species tested, including yeast, *Drosophila*, *C. elegans* [96]. The antiaging effect of spermidine was abolished by the deletion or depletion of essential autophagy genes in yeast, *Drosophila* and *C. elegans* [96]. In mice, a dietary supplementation with polyanions (including

spermidine) also increases healthspan and lifespan [97], although the dependency of this phenomenon on autophagy has not been addressed yet. Spermidine likewise induces autophagy and longevity through its capacity to inhibit histone acetylases in yeast cells [96].

Sirtuin-1 and that of its *C. elegans* orthologue induce autophagy in human and nematode cells. Sirtuin-1 is also required for the induction of autophagy by its allosteric activator resveratrol (both in human cells and nematodes), culture in nutrient-free media (in human cells) and caloric restriction (in nematodes). In *C. elegans*, it was found that activation of Sirtuin-1 extended longevity in an autophagy-dependent fashion. Thus, the knockdown of the essential autophagy gene Beclin1/ATG6 abolished life span extension by Sirtuin-1 activation [87]. These results underscore the contribution autophagy to the regulation of longevity by pharmacological agents [98].

Post-transcriptional gene regulation and aging

In fact, 2009 saw an escalation in interest in microRNAs and other non-coding RNAs implicated in aging and replicative senescence. A prominent example of this regulation came studies of the mitogen-activated protein kinase (MAPK) signaling component MKK4 (MAPK kinase kinase 4). MKK4 levels were elevated in aging tissues and in senescent cells thanks to reductions in the abundance of four microRNAs (miR-15b, miR-24, miR-25, and miR-141) that interacted with the 5'- and 3'-untranslated regions of the MKK4 mRNA and repressed its translation [99].

The other major class of post-transcriptional regulatory factors, RNA-binding proteins (RBPs), were also the focus of important age-related studies in 2009. Several RBPs that affect the turnover and translation of proteins implicated in proliferation, survival, inflammation, neurodegeneration, and cancer (HuR, AUF1, TIA-1, TTP) displayed elevated abundance in a broad array of human tissues and in all ages, suggesting that their influence extends throughout the human life span [100]. The RBP TTP (tristetraprolin) attracted especial attention because it triggered replicative senescence [101]; in keeping with the tumor-suppressive influence of replicative senescence, TTP was found to be eliminated in certain cancers [102].

Circadian clock

There is growing evidence for a link between circadian rhythm, signal-transduction genes, metabolism, cancer and aging [103, 104]. The circadian clock gene *period* extended the health span of aging in *Drosophila*

melanogaster [105]. Further, circadian control of the NAD⁺ salvage pathway by CLOCK-SIRT1 was demonstrated [106]. Intriguingly, light was found to activate MAPK (mitogen activated pathway kinase) in zebrafish cells, and this light-dependent activation controlled DNA repair [107]. In rats, circadian disruption induced by light-at-night accelerates aging and promotes tumorigenesis in rats [108]. In mice, it was reported that N-acetyl-L-cysteine (NAC), an antioxidant, ameliorated symptoms of premature aging associated with the deficiency of the circadian protein BMAL1 [109].

Cancer and aging

CR is known to slow aging and delay cancer. In 2009, it was reported that fasting abrogates side effects caused by chemotherapy in cancer patients. Importantly, for those patients in whom cancer progression could be assessed, fasting did not prevent chemotherapy-induced reduction of tumor volume or tumor markers [110]. The link between aging and cancer via p53 was shown to be complex in 2009. Thus, the ability of p53 to act as a defense against tumor progression was shown to be age-dependent [111]. Further, Levine and co-workers previously showed that p53 activity declines with age, and a recent study showed that p53 transcriptional activity is reduced in senescent cells [112]. Interestingly, SIRT1 knockout mice, which do not live longer when calorically restricted, were found to have normal rates of skin cancer but the ability of resveratrol, a SIRT1 activator, to protect the mice was greatly reduced [113], indicating that the anti-tumor activity of resveratrol is mediated at least in part by SIRT1.

Reduced incidence and delayed occurrence of fatal neoplastic diseases in growth hormone receptor/binding protein knockout mice. These changes of fatal neoplasms are similar to the effects observed with calorie restriction and therefore could possibly be a major contributing factor to the extended life span observed in the GHR/BP KO mice. [114]

Overall, 2009 was an exciting year for increasing our understanding of aging and its relationship to age-related disease, and developing promising strategies and candidates for pharmacological interventions into the aging process. Several approaches in combination with drugs and diet may slow aging, although not making it negligible [115].

ACKNOWLEDGMENTS AND ANNOUNCEMENTS

We apologize to the authors whose important publications were not discussed due to space limitations

or were simply overlooked. Here we referenced only papers published in the 2009 calendar year. That was not an easy task given that most of publications are the continuation of or based on previous research. We expect to make this a tradition to publish ‘the year overview’ every year. The next year, the task will be easier, given that “Aging in 2010” will be the continuation of “Aging in 2009”.

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REFERENCES

1. Epel ES, Merkin SS, Cawthon R, Blackburn EH, Adler NE, M.J. P, Seeman TE. The rate of leukocyte telomere shortening predicts mortality from cardiovascular disease in elderly men. *Aging* 2009, 1: 81-88.
2. O'Donovan A, Lin J, Dhabhar FS, Wolkowitz O, Tillie JM, Blackburn E, Epel E. Pessimism correlates with leukocyte telomere shortness and elevated interleukin-6 in post-menopausal women. *Brain Behav Immun* 2009, 23: 446-449.
3. Lou Z, Wei J, Riethman H, Baur JA, Voglauer R, Shay JW, Wright WE. Telomere length regulates ISG15 expression in human cells. *Aging* 2009, 1: 608-621.
4. Bhaumik D, Scott GK, Schokrpur S, Patil CK, Orjalo AV, Rodier F, Lithgow GJ, Campisi J. MicroRNAs miR-146a/b negatively modulate the senescence-associated inflammatory mediators IL-6 and IL-8. *Aging* 2009, 1: 402-411.
5. Martinez P, Thanasoula M, Munoz P, Liao C, Tejera A, McNees C, Flores JM, Fernandez-Capetillo O, Tarsounas M, Blasco MA. Increased telomere fragility and fusions resulting from TRF1 deficiency lead to degenerative pathologies and increased cancer in mice. *Genes Dev* 2009, 23: 2060-2075.
6. Ghosh A, Rossi ML, Aulds J, Croteau D, Bohr VA. Telomeric D-loops containing 8-oxo-2'-deoxyguanosine are preferred substrates for Werner and Bloom syndrome helicases and are bound by POT1. *J Biol Chem* 2009, 284: 31074-31084.
7. Nakamura AJ, Redon CE, Bonner WM, Sedelnikova OA. Telomere-dependent and telomere-independent origins of endogenous DNA damage in tumor cells. *Aging* 2009, 1: 212-218.
8. Rodier F, Coppé JP, Patil CK, Hoeijmakers WA, Muñoz DP, Raza SR, Freund A, Campeau E, Davalos AR, Campisi J. Persistent DNA damage signalling triggers senescence-associated inflammatory cytokine secretion. *Nat Cell Biol.* 2009, 11: 973-979.
9. Orjalo AV, Bhaumik D, Gengler BK, Scott GK, Campisi J. Cell surface-bound IL-1alpha is an upstream regulator of the senescence-associated IL-6/IL-8 cytokine network. *Proc Natl Acad Sci U S A* 2009, 106: 17031-17036.
10. Pospelova TV, Demidenk ZN, Bukreeva EI, Pospelov VA, Gudkov AV, Blagosklonny MV. Pseudo-DNA damage response in senescent cells. *Cell Cycle* 2009, 8: 4112-4118.
11. Demidenko ZN, Zubova SG, Bukreeva EI, Pospelov VA, Pospelova TV, Blagosklonny MV. Rapamycin decelerates cellular senescence. *Cell Cycle* 2009, 8: 1888-1895.
12. Madia F, Wei M, Yuan V, Hu J, Gattazzo C, Pham P, Goodman MF, Longo VD. Oncogene homologue Sch9 promotes age-dependent mutations by a superoxide and Rev1/Polzeta-dependent mechanism. *J Cell Biol* 2009, 186: 509-523.
13. D'Angelo MA, Raices M, Panowski SH, Hetzer MW. Age-dependent deterioration of nuclear pore complexes causes a loss of nuclear integrity in postmitotic cells. *Cell* 2009, 136: 284-295.
14. Westerheide SD, Anckar J, Stevens SMJ, Sistonen L, Morimoto RI. Stress-inducible regulation of heat shock factor 1 by the deacetylase SIRT1. *Science* 2009, 323: 1063-1066.
15. Sinclair DA, Oberdoerffer P. The ageing epigenome: damaged beyond repair? *Ageing Res Rev* 2009, 8: 189-198.
16. Pérez VI, Van Remmen H, Bokov A, Epstein CJ, Vijg J, Richardson A. The overexpression of major antioxidant enzymes does not extend the lifespan of mice. *Aging Cell* 2009, 8: 73-75.
17. Van Raamsdonk JM, Hekimi S. Deletion of the mitochondrial superoxide dismutase sod-2 extends lifespan in *Caenorhabditis elegans*. *PLoS Genet* 2009, 5: e1000361.
18. Edman U, Garcia AM, Busuttill RA, Sorensen D, Lundell M, Kapahi P, Vijg J. Lifespan extension by dietary restriction is not linked to protection against somatic DNA damage in *Drosophila melanogaster*. *Aging Cell* 2009, 8: 331-338.
19. Guachalla LM, Ju Z, Koziel R, von Figura G, Song Z, Fusser M, Epe B, Jansen-Dirr P, Rudolph KL. Sod2 haploinsufficiency does not accelerate aging of telomere dysfunctional mice. *Aging* 2009, 1: 303-315.
20. Lapointe J, Stepanyan Z, Bigras E, Hekimi S. Reversal of the mitochondrial phenotype and slow development of oxidative biomarkers of aging in long-lived Mcl1+/- mice. *J Biol Chem* 2009, 284: 20364-20374.
21. Copeland JM, Cho J, Lo T, Jr., Hur JH, Bahadorani S, Arabyan T, Rabie J, Soh J, Walker DW. Extension of *Drosophila* life span by RNAi of the mitochondrial respiratory chain. *Curr Biol* 2009, 19: 1591-1598.
22. Luce K, Osiewacz HD. Increasing organismal healthspan by enhancing mitochondrial protein quality control. *Nat Cell Biol* 2009, 11: 852-858.
23. Kunstmann B, Osiewacz HD. The S-adenosylmethionine dependent O-methyltransferase PaMTH1: a longevity assurance factor protecting *Podospora anserina* against oxidative stress. *Aging* 2009, 1: 328-334.
24. Colman RJ, Anderson RM, Johnson SC, Kastman EK, Kosmatka KJ, Beasley TM, Allison DB, Cruzen C, Simmons HA, Kemnitz JW, Weindruch R. Caloric restriction delays disease onset and mortality in rhesus monkeys. *Science* 2009, 325: 201-204.
25. Bauer JH, Morris SN, Chang C, Flatt T, Wood JG, Helfand SL. dSir2 and Dmp53 interact to mediate aspects of CR-dependent life span extension in *D. melanogaster*. *Aging* 2009, 1: 38-48.
26. Carrano AC, Liu Z, Dillin A, Hunter T. A conserved ubiquitination pathway determines longevity in response to diet restriction. *Nature* 2009, 460: 396-399.
27. Honjoh S, Yamamoto T, Uno M, Nishida E. Signalling through RHEB-1 mediates intermittent fasting-induced longevity in *C. elegans*. *Nature* 2009, 457: 726-730.
28. Wei M, Fabrizio P, Madia F, Hu J, Ge H, Li LM, Longo VD. Tor1/Sch9-regulated carbon source substitution is as effective as calorie restriction in life span extension. *PLoS Genet.* 2009, 5: e1000467.
29. Zid BM, Rogers AN, Katewa SD, Vargas MA, Kolipinski MC, Lu TA, Benzer S, Kapahi P. 4E-BP extends lifespan upon dietary

restriction by enhancing mitochondrial activity in *Drosophila*. *Cell* 2009, 139: 149-160.

30. Lee SJ, Murphy CT, Kenyon C. Glucose shortens the life span of *C. elegans* by downregulating DAF-16/FOXO activity and aquaporin gene expression. *Cell Metab* 2009, 10: 379-391.

31. Chen D, Thomas EL, Kapahi P. HIF-1 modulates dietary restriction-mediated lifespan extension via IRE-1 in *Caenorhabditis elegans*. *PLoS Genet* 2009, 5: e1000486.

32. Mehta R, Steinkraus KA, Sutphin GL, Ramos FJ, Shamieh LS, Huh A, Davis C, Chandler-Brown D, Kaerberlein M. Proteasomal regulation of the hypoxic response modulates aging in *C. elegans*. *Science* 2009, 324: 1196-1198.

33. Kaerberlein M, Kapahi P. The hypoxic response and aging. *Cell Cycle* 2009, 8: 2324.

34. Greer EL, Brunet A. Different dietary restriction regimens extend lifespan by both independent and overlapping genetic pathways in *C. elegans*. *Aging Cell* 2009, 8: 113-127.

35. Cohen DE, Supinski AM, Bonkowski MS, Donmez G, Guarente LP. Neuronal SIRT1 regulates endocrine and behavioral responses to calorie restriction. *Genes Dev* 2009, 23: 2812-2817.

36. Bonkowski MS, Dominici FP, Arum O, Rocha JS, Al Regaiey KA, Westbrook R, Spong A, Panici J, Masternak MM, Kopchick JJ, Bartke A. Disruption of growth hormone receptor prevents calorie restriction from improving insulin action and longevity. *PLoS One* 2009, 4: e4567.

37. Grandison RC, Piper MD, Partridge L. Amino-acid imbalance explains extension of lifespan by dietary restriction in *Drosophila*. *Nature* 2009, 462: 1061-1064.

38. Demidenko ZN, Shtutman M, Blagosklonny MV. Pharmacologic inhibition of MEK and PI-3K converges on the mTOR/S6 pathway to decelerate cellular senescence. *Cell Cycle* 2009, 8: 1896-1900.

39. Harrison DE, Strong R, Sharp ZD, Nelson JF, Astle CM, Flurkey K, Nadon NL, Wilkinson JE, Frenkel K, Carter CS, Pahor M, Javors MA, Fernandez E, Miller RA. Rapamycin fed late in life extends lifespan in genetically heterogeneous mice. *Nature* 2009, 460: 392-396.

40. Chen C, Liu Y, Zheng P. mTOR regulation and therapeutic rejuvenation of aging hematopoietic stem cells. *Sci Signal* 2009, 2: ra75.

41. Wang Y, Branicky R, Stepanyan Z, Carroll M, Guimond MP, Hiji A, Hayes S, McBride K, Hekimi S. The anti-neurodegeneration drug cloquinol inhibits the aging-associated protein CLK-1. *J Biol Chem* 2009, 284: 314-323.

42. Skulachev VP, Anisimov VN, Antonenko YN, Bakeeva LE, Chernyak BV, Elichev VP, Filenko OF, Kalinina NI, Kapelko VI, Kolosova NG, Kopnin BP, Korshunova GA, Lichinitser MR, Obukhova LA, Pasyukova EG, Pisarenko OI, Roginsky VA, Ruuge EK, Senin II, Severina II, Skulachev MV, Spivak IM, Tashlitsky VN, Tkachuk VA, Vyssokikh MY, Yaguzhinsky LS, Zorov DB. An attempt to prevent senescence: a mitochondrial approach. *Biochim Biophys Acta* 2009, 1787: 437-461.

43. Obukhova LA, Skulachev VP, Kolosova NG. Mitochondria-targeted antioxidant SkQ1 inhibits age-dependent involution of the thymus in normal and senescence-prone rats. *Aging* 2009, 1: 389-401.

44. Castilho RM, Squarize CH, Chodosh LA, Williams BO, Gutkind JS. mTOR mediates Wnt-induced epidermal stem cell exhaustion and aging. *Cell Stem Cell* 2009, 5: 279-289.

45. Gan B, DePinho RA. mTORC1 signaling governs hematopoietic stem cell quiescence. *Cell Cycle* 2009, 8: 1003-1006.

46. Paik JH, Ding Z, Narurkar R, Ramkissoon S, Muller F, Kamoun WS, Chae SS, Zheng H, Ying H, Mahoney J, Hiller D, Jiang S, Protopopov A, Wong WH, Chin L, Ligon KL, DePinho RA. FoxOs cooperatively regulate diverse pathways governing neural stem cell homeostasis. *Cell Stem Cell* 2009, 5: 540-553.

47. Renault VM, Rafalski VA, Morgan AA, Salih DA, Brett JO, Webb AE, Villeda SA, Thekkat PU, Guillerey C, Denko NC, Palmer TD, Butte AJ, Brunet A. FoxO3 regulates neural stem cell homeostasis. *Cell Stem Cell* 2009, 5: 527-539.

48. Selesniemi K, Lee HJ, Niikura T, Tilly JL. Young adult donor bone marrow infusions into female mice postpone age-related reproductive failure and improve offspring survival. *Aging* 2009, 1: 49-57.

49. Niikura Y, Niikura T, Tilly JL. Aged mouse ovaries possess rare premeiotic germ cells that can generate oocytes following transplantation into a young host environment. *Aging* 2009, 1: 971-978.

50. Li H, Collado M, Villasante A, Strati K, Ortega S, Canamero M, Blasco MA, Serrano M. The Ink4/Arf locus is a barrier for iPS cell reprogramming. *Nature* 2009, 460: 1136-1139.

51. Kawamura T, Suzuki J, Wang YV, Menendez S, Morera LB, Raya A, Wahl GM, Belmonte JC. Linking the p53 tumour suppressor pathway to somatic cell reprogramming. *Nature* 2009, 460: 1140-1144.

52. Utikal J, Polo JM, Stadtfeld M, Maherali N, Kulalert W, Walsh RM, Khalil A, Rheinwald JG, Hochedlinger K. Immortalization eliminates a roadblock during cellular reprogramming into iPS cells. *Nature* 2009, 460: 1145-1148.

53. Marion RM, Strati K, Li H, Murga M, Blanco R, Ortega S, Fernandez-Capetillo O, Serrano M, Blasco MA. A p53-mediated DNA damage response limits reprogramming to ensure iPS cell genomic integrity. *Nature* 2009, 460: 1149-1153.

54. Hanna J, Saha K, Pando B, van Zon J, Lengner CJ, Creyghton MP, van Oudenaarden A, Jaenisch R. Direct cell reprogramming is a stochastic process amenable to acceleration. *Nature* 2009, 462: 595-601.

55. Banito A, Rashid ST, Acosta JC, Li S, Pereira CF, Geti I, Pinho S, Silva JC, Azuara V, Walsh M, Vallier L, Gil J. Senescence impairs successful reprogramming to pluripotent stem cells. *Genes Dev* 2009, 23: 2134-2139.

56. Liu Y, Sanoff HK, Cho H, Burd CE, Torrice C, Ibrahim JG, Thomas NE, Sharpless NE. Expression of p16(INK4a) in peripheral blood T-cells is a biomarker of human aging. *Aging Cell* 2009, 8: 439-448.

57. Liu Y, Sanoff HK, Cho H, Burd CE, Torrice C, Mohlke KL, Ibrahim JG, Thomas NE, Sharpless NE. INK4/ARF transcript expression is associated with chromosome 9p21 variants linked to atherosclerosis. *PLoS One* 2009, 4: e5027.

58. Matheu A, Maraver A, Collado M, Garcia-Cao I, Canamero M, Borras C, Flores JM, Klatt P, Vina J, Serrano M. Anti-aging activity of the Ink4/Arf locus. *Aging Cell* 2009, 8: 152-161.

59. Pan Y, Shadel GS. Extension of chronological life span by reduced TOR signaling requires down-regulation of Sch9p and involves increased mitochondrial OXPHOS complex density. *Aging* 2009, 1: 131-145.

60. Soukas AA, Kane EA, Carr CE, Melo JA, Ruvkun G. Rictor/TORC2 regulates fat metabolism, feeding, growth, and life span in *Caenorhabditis elegans*. *Genes Dev* 2009, 23: 496-511.

61. Ben-Zvi A, Miller EA, Morimoto RI. Collapse of proteostasis represents an early molecular event in *Caenorhabditis elegans* aging. *Proc Natl Acad Sci U S A*. 2009,
62. Selman C, Tullet JM, Wieser D, Irvine E, Lingard SJ, Choudhury AI, Claret M, Al-Qassab H, Carmignac D, Ramadani F, Woods A, Robinson IC, Schuster E, Batterham RL, Kozma SC, Thomas G, Carling D, Okkenhaug K, Thornton JM, Partridge L, Gems D, Withers DJ. Ribosomal protein S6 kinase 1 signaling regulates mammalian life span. *Science* 2009, 326: 140-144.
63. Enns LC, Morton JF, Treuting PR, Emond MJ, Wolf NS, McKnight GS, Rabinovitch PS, Ladiges WC. Disruption of protein kinase A in mice enhances healthy aging. *PLoS One* 2009, 4: e5963.
64. Wessells R, Fitzgerald E, Piazza N, Ocorr K, Morley S, Davies C, Lim HY, Elmen L, Hayes M, Oldham S, Bodmer R. d4eBP acts downstream of both dTOR and dFoxo to modulate cardiac functional aging in *Drosophila*. *Aging Cell* 2009, 8: 542-552.
65. McCord RA, Michishita E, Hong T, Berber E, Boxer LD, Kusumoto R, Guan S, Shi X, Gozani O, Burlingame AL, Bohr VA, Chua KF. SIRT6 stabilizes DNA-dependent Protein Kinase at chromatin for DNA double-strand break repair. *Aging* 2009, 1: 109-121.
66. Ghazi A, Henis-Korenblit S, Kenyon C. A transcription elongation factor that links signals from the reproductive system to lifespan extension in *Caenorhabditis elegans*. *PLoS Genet*. 2009, 5: e1000639.
67. Cheng Z, Guo S, Capps K, Dong X, Kollipara R, Rodgers JT, Depinho RA, Puigserver P, White MF. Foxo1 integrates insulin signaling with mitochondrial function in the liver. *Nat Med* 2009, 15: 1307-1311.
68. Li Y, Wang WJ, Cao H, Lu J, Wu C, Hu FY, Guo J, Zhao L, Yang F, Zhang YX, Li W, Zheng GY, Cui H, Chen X, Zhu Z, He H, Dong B, Mo X, Zeng Y, Tian XL. Genetic association of FOXO1A and FOXO3A with longevity trait in Han Chinese populations. *Hum Mol Genet* 2009, 18: 4897-4904.
69. Pawlikowska L, Hu D, Huntsman S, Sung A, Chu C, Chen J, Joyner AH, Schork NJ, Hsueh WC, Reiner AP, Psaty BM, Atzmon G, Barzilai N, Cummings SR, Browner WS, Kwok PY, Ziv E. Association of common genetic variation in the insulin/IGF1 signaling pathway with human longevity. *Aging Cell* 2009, 8: 460-472.
70. Anselmi CV, Malovini A, Roncarati R, Novelli V, Villa F, Condorelli G, Bellazzi R, Puca AA. Association of the FOXO3A locus with extreme longevity in a southern Italian centenarian study. *Rejuvenation Res* 2009, 12: 95-104.
71. Flachsbarth F, Caliebe A, Kleindorp R, Blanche H, von Eller-Eberstein H, Nikolaus S, Schreiber S, Nebel A. Association of FOXO3A variation with human longevity confirmed in German centenarians. *Proc Natl Acad Sci U S A* 2009, 106: 2700-2705.
72. Waskar M, Landis GN, Shen J, Curtis C, Tozer K, Abdueva D, Skvortsov D, Tavaré S, Tower J. *Drosophila melanogaster* p53 has developmental stage-specific and sex-specific effects on adult life span indicative of sexual antagonistic pleiotropy. *Aging* 2009, 1: 903-936.
73. Donehower LA. Longevity regulation in flies: a role for p53. *Aging* 2009, 1: 6-8.
74. Kang HJ, Feng Z, Sun Y, Atwal G, Murphy ME, Rebbeck TR, Rosenwaks Z, Levine AJ, Hu W. Single-nucleotide polymorphisms in the p53 pathway regulate fertility in humans. *Proc Natl Acad Sci U S A* 2009, 106: 9761-9766.
75. Gravina S, Lescai F, Hurteau G, Brock GJ, Saramaki A, Salvioli S, Franceschi C, Roninson IB. Identification of single nucleotide polymorphisms in the p21 (CDKN1A) gene and correlations with longevity in the Italian population. *Aging* 2009, 1: 470.
76. Flores I, Blasco MA. A p53-dependent response limits epidermal stem cell functionality and organismal size in mice with short telomeres. *PLoS One* 2009, 4: e4934.
77. Begus-Nahrman Y, Lechel A, Obenauf AC, Nalapareddy K, Peit E, Hoffmann E, Schlaudraff F, Liss B, Schirmacher P, Kestler H, Danenberg E, Barker N, Clevers H, Speicher MR, Rudolph KL. p53 deletion impairs clearance of chromosomal-instable stem cells in aging telomere-dysfunctional mice. *Nat Genet* 2009, 41: 1138-1143.
78. Su X, Paris M, Gi YJ, Tsai KY, Cho MS, Lin YL, Biernaskie JA, Sinha S, Prives C, Pevny LH, Miller FD, Flores ER. TAp63 prevents premature aging by promoting adult stem cell maintenance. *Cell Stem Cell* 2009, 5: 64-75.
79. Aggarwal M, Brosh RM, Jr. Premature aging syndrome gene WRN genetically interacts with a topoisomerase. *Cell Cycle* 2009, 8: 2143.
80. Aggarwal M, Brosh RM. WRN helicase defective in the premature aging disorder Werner syndrome genetically interacts with topoisomerase 3 and restores the top3 slow growth phenotype of *sgs1 top3*. *Aging* 2009, 1: 219-233.
81. Dhawan S, Tschen SI, Bhushan A. Bmi-1 regulates the Ink4a/Arf locus to control pancreatic beta-cell proliferation. *Genes Dev* 2009, 23: 906-911.
82. Chen H, Gu X, Su IH, Bottino R, Contreras JL, Tarakhovskiy A, Kim SK. Polycomb protein Ezh2 regulates pancreatic beta-cell Ink4a/Arf expression and regeneration in diabetes mellitus. *Genes Dev* 2009, 23: 975-985.
83. Wong ES, Le Guezennec X, Demidov ON, Marshall NT, Wang ST, Krishnamurthy J, Sharpless NE, Dunn NR, Bulavin DV. p38MAPK controls expression of multiple cell cycle inhibitors and islet proliferation with advancing age. *Dev Cell* 2009, 17: 142-149.
84. Young AR, Narita M, Ferreira M, Kirschner K, Sadaie M, Darot JF, Tavaré S, Arakawa S, Shimizu S, Watt FM, Narita M. Autophagy mediates the mitotic senescence transition. *Genes Dev*. 2009, 23:798-803
85. Ren C, Finkel SE, Tower J. Conditional inhibition of autophagy genes in adult *Drosophila* impairs immunity without compromising longevity. *Exp Gerontol* 2009, 44: 228-235.
86. Hansen M, Chandra A, Mitic LL, Onken B, Driscoll M, Kenyon C. A role for autophagy in the extension of lifespan by dietary restriction in *C. elegans*. *PLoS Genet* 2008, 4: e24.
87. Morselli E, Galluzzi L, Kepp O, Criollo A, Maiuri MC, Tavernarakis N, Madeo F, Kroemer G. Autophagy mediates pharmacological lifespan extension by spermidine and resveratrol. *Aging* 2009, 1: 961-970.
88. Alvers AL, Wood MS, Hu D, Kaywell AC, Dunn WA, Jr., Aris JP. Autophagy is required for extension of yeast chronological life span by rapamycin. *Autophagy* 2009, 5: 847-849.
89. Dwivedi M, Song HO, Ahnn J. Autophagy genes mediate the effect of calcineurin on life span in *C. elegans*. *Autophagy* 2009, 5: 604-607.
90. Tavernarakis N, Pasparaki A, Tasdemir E, Maiuri MC, Kroemer G. The effects of p53 on whole organism longevity are mediated by autophagy. *Autophagy* 2008, 4: 870-873.

- 91.** Armour SM, Joseph A, Baur, Sherry N, Hsieh SN, Land-Bracha A, Thomas SM, Sinclair DA. Inhibition of mammalian S6 kinase by resveratrol suppresses autophagy. *Aging* 2009, 1: 515-528.
- 92.** Stephan JS, Yeh YY, Ramachandran V, Deminoff SJ, Herman PK. The Tor and PKA signaling pathways independently target the Atg1/Atg13 protein kinase complex to control autophagy. *Proc Natl Acad Sci U S A* 2009, 106: 17049-17054.
- 93.** Chang YY, Neufeld TP. An Atg1/Atg13 complex with multiple roles in TOR-mediated autophagy regulation. *Mol Biol Cell* 2009, 20: 2004-2014.
- 94.** Jung CH, Jun CB, Ro SH, Kim YM, Otto NM, Cao J, Kundu M, Kim DH. ULK-Atg13-FIP200 complexes mediate mTOR signaling to the autophagy machinery. *Mol Biol Cell* 2009, 20: 1992-2003.
- 95.** Wang T, Lao U, Edgar BA. TOR-mediated autophagy regulates cell death in *Drosophila* neurodegenerative disease. *J Cell Biol* 2009, 186: 703-711.
- 96.** Eisenberg T, Knauer H, Schauer A, Buttner S, Ruckenstein C, Carmona-Gutierrez D, Ring J, Schroeder S, Magnes C, Antonacci L, Fussi H, Deszcz L, Hartl R, Schraml E, Criollo A, Megalou E, Weiskopf D, Laun P, Heeren G, Breitenbach M, Grubeck-Loebenstein B, Herker E, Fahrenkrog B, Frohlich KU, Sinner F, Tavernarakis N, Minois N, Kroemer G, Madeo F. Induction of autophagy by spermidine promotes longevity. *Nat Cell Biol* 2009, 11: 1305-1314.
- 97.** Soda K, Dobashi Y, Kano Y, Tsujinaka S, Konishi F. Polyamine-rich food decreases age-associated pathology and mortality in aged mice. *Exp Gerontol* 2009, 44: 727-732.
- 98.** Madeo F, Eisenberg T, Kroemer G. Autophagy for the avoidance of neurodegeneration. *Genes Dev* 2009, 23: 2253-2259.
- 99.** Marasa BS, Srikantan S, Masuda K, Abdelmohsen K, Kuwano Y, Yang X, Martindale JL, Rinker-Schaeffer CW, Gorospe M. Increased MKK4 abundance with replicative senescence is linked to the joint reduction of multiple microRNAs. *Sci Signal* 2009, 2: ra69.
- 100.** Masuda K, Marasa B, Martindale JL, Halushka MK, Gorospe M. Tissue- and age-dependent expression of RNA-binding proteins that influence mRNA turnover and translation. *Aging* 2009, 1: 681-698.
- 101.** Sanduja S, Kaza V, Dixon DA. The mRNA decay factor tristetraprolin (TTP) induces senescence in human papillomavirus-transformed cervical cancer cells by targeting E6-AP ubiquitin ligase. *Aging* 2009, 1: 803-817.
- 102.** Brennan SE, Kuwano Y, Alkharouf N, Blackshear PJ, Gorospe M, Wilson GM. The mRNA-destabilizing protein tristetraprolin is suppressed in many cancers, altering tumorigenic phenotypes and patient prognosis. *Cancer Res* 2009, 69: 5168-5176.
- 103.** Kang TH, Sancar A. Circadian regulation of DNA excision repair: implications for chrono-chemotherapy. *Cell Cycle* 2009, 8: 1665-1667.
- 104.** Sahar S, Sassone-Corsi P. Metabolism and cancer: the circadian clock connection. *Nat Rev Cancer* 2009, 9: 886-896.
- 105.** Krishnan N, Kretzschmar D, Rakshit K, Chow E, Giebultowicz JM. The circadian clock gene period extends healthspan in aging *Drosophila melanogaster*. *Aging* 2009, 1: 937-948.
- 106.** Nakahata Y, Sahar S, Astarita G, Kaluzova M, Sassone-Corsi P. Circadian control of the NAD+ salvage pathway by CLOCK-SIRT1. *Science* 2009, 324: 598-599.
- 107.** Hirayama J, Miyamura N, Uchida Y, Asaoka Y, Honda R, Sawanobori K, Todo T, Yamamoto T, Sassone-Corsi P, Nishina H. Common light signaling pathways controlling DNA repair and circadian clock entrainment in zebrafish. *Cell Cycle* 2009, 8: 2794-2801.
- 108.** Vinogradova IA, Anisimov VN, Bukalev AV, Semenchenko AV, Zabezhinski MA. Circadian disruption induced by light-at-night accelerates aging and promotes tumorigenesis in rats. *Aging* 2009, 1: 855-865.
- 109.** Kondratov RV, Vykhovanets O, Kondratova AA, Antoch MP. Antioxidant N-acetyl-L-cysteine ameliorates symptoms of premature aging associated with the deficiency of the circadian protein BMAL1. *Aging* 2009, 1: 979-987.
- 110.** Safdie FM, Dorff T, Quinn D, Fontana L, Wei M, Lee C, Cohen P, Longo VD. Fasting and cancer treatment in humans: A case series report. *Aging* 2009, 1: 988-1007.
- 111.** Hinkal G, Parikh N, Donehower LA. Timed somatic deletion of p53 in mice reveals age-associated differences in tumor progression. *PLoS One* 2009, 4: e6654.
- 112.** Huang B, Vassilev LT. Reduced transcriptional activity in the p53 pathway of senescent cells revealed by the MDM2 antagonist nutlin-3. *Aging* 2009, 1: 845-854.
- 113.** Boily G, He XH, Pearce B, Jardine K, McBurney MW. SirT1-null mice develop tumors at normal rates but are poorly protected by resveratrol. *Oncogene* 2009, 28: 2882-2893.
- 114.** Ikeno Y, Hubbard GB, Lee S, Cortez LA, Lew CM, Webb CR, Berryman DE, List EO, Kopchick JJ, Bartke A. Reduced incidence and delayed occurrence of fatal neoplastic diseases in growth hormone receptor/binding protein knockout mice. *J Gerontol A Biol Sci Med Sci* 2009, 64: 522-529.
- 115.** Finch CE. Update on slow aging and negligible senescence--a mini-review. *Gerontology* 2009, 55: 307-313.